

**An Account of the Taxonomy of North American Wolves from Morphological and Genetic**

**Analyses**

**Steven M. Chambers,\* Steven R. Fain, Bud Fazio, Michael Amaral**

**S.M. Chambers**

Division of Ecological Services, U.S. Fish and Wildlife Service, PO Box 1306, Albuquerque,  
New Mexico 87103

**S.R. Fain**

National Forensics Laboratory, U.S. Fish and Wildlife Service, 1490 East Main Street, Ashland,  
Oregon 97520-1310

**B. Fazio**

Mexican Wolf Recovery Program, New Mexico Ecological Services Field Office, U.S. Fish and  
Wildlife Service, 2105 Osuna Road, NE, Albuquerque, New Mexico 87113. Current address:  
Wildlife and Sport Fish Restoration Program, U.S. Fish and Wildlife Service, PO Box 1306,  
Albuquerque, New Mexico 87103.

**M. Amaral**

New England Field Office, U.S. Fish and Wildlife Service, 70 Commercial Street, Suite 300  
Concord, New Hampshire 03301-5087

24 \*Corresponding author: [steve\\_chambers@fws.gov](mailto:steve_chambers@fws.gov)

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## **Abstract**

The available scientific literature was reviewed to assess the taxonomic standing of North American wolves, including subspecies of the gray wolf, *Canis lupus*. The recent scientific proposal that the eastern wolf, *Canis lupus lycaon*, is not a subspecies of gray wolf, but a full species, *Canis lycaon*, is well-supported by both morphological and genetic data and should be accepted. This species' range extends westward to Minnesota, and it hybridizes with gray wolves where the two species are in contact in eastern Canada, the Upper Peninsula of Michigan, Wisconsin, and Minnesota. Genetic data support a close relationship between eastern wolf and red wolf (*Canis rufus*), but do not support the proposal that they are the same species; it is more likely that they evolved independently from different lineages of a common ancestor with coyotes. The Mexican wolf (*Canis lupus baileyi*) is a well-supported subspecies. The available genetic and morphometric data do not provide clear support for the recognition of the Arctic wolf (*Canis lupus arctos*), but the available genetic data are almost entirely limited to one group of genetic markers (microsatellite DNA) and are not definitive on this question. The northern timber wolf (*Canis lupus occidentalis*) and the plains wolf (*Canis lupus nubilus*) are valid subspecies. Their recognition is supported by morphological data and extensive studies of microsatellite DNA variation where both subspecies are in contact in Canada. The wolves of coastal areas in southeastern Alaska and British Columbia should be assigned to *Canis lupus nubilus*. There is scientific support for the taxa recognized here, but delineation of exact geographic boundaries presents challenges. Rather than sharp boundaries between taxa, boundaries should generally be thought of as intergrade zones of variable width.

## **Introduction**

The taxonomy of North American members of the genus *Canis* has a complicated and controversial history. This is not surprising in light of their variability in size, proportions, and pelage; large geographic ranges; tendency of various forms to interbreed; and their extirpation over large areas beginning early in the period of colonization by Europeans. Members of North American *Canis*, exclusive of coyotes (*Canis latrans*), are commonly referred to as “wolves.” For these North American wolves, 31 published names for subspecies or species are available (Hall and Kelson 1959, Table 1 of this paper). The two most recent comprehensive taxonomic reviews based on morphology both recognize two species, *Canis lupus* (gray wolf) and *Canis rufus* (red wolf), but differ in that they recognize as many as 27 (Hall [1981], based primarily on Goldman [1944]) or as few as eight subspecies (Nowak 1995) for the two species collectively.

The first of many studies of *Canis* using molecular genetic markers (Wayne and Jenks 1991, Lehman et al. 1991) raised new challenges to the general taxonomic scheme (Goldman 1944) that had stood for almost 50 years. In particular the possible role of coyotes in the ancestry of both the red wolf and what had been considered gray wolves in the Great Lakes region generated new controversy. Development of even more powerful genetic markers has led to new, highly controversial interpretations, such as the distinctiveness of wolves of the Great Lakes region from gray wolves and the possibility that they are conspecific with red wolves (Wilson et al. 2000), a proposal rejected by others based on genetics (e.g., Koblmüller et al. 2009a) and morphometrics (e.g., Nowak 2009). Other controversies include whether the current Great Lakes wolf population is evolutionarily representative of the historical population (Leonard and Wayne 2008), the taxonomic identity of Minnesota wolves (Nowak 2009), the

historical northern boundary of the Mexican wolf (*Canis lupus baileyi*) (Leonard et al. 2005), and the taxonomic identity of wolves of Pacific coastal regions (Muñoz-Fuentes et al. 2009).

The extreme lack of consensus among researchers on so many important issues related to the taxonomy of North American wolves prompted the present review.

## **Scope and intent**

The purpose of this review is to explore the scientific support in the currently available scientific literature for: (1) recognizing any taxonomic subdivisions, including species and subspecies, of North American wolves; (2) recommending at least general geographic boundaries for any recognized taxa, either species or subspecies; and (3) recommending additional research and analysis that would improve the scientific basis for evaluating the taxonomy of wolves.

This review provides the authors' views only on whether the validity of each taxon is supported by a preponderance of evidence from the relevant, available scientific literature. It is important to emphasize the following points about the scope of this review:

(1) It is an evaluation and synthesis of the available scientific literature. It is not intended to generate and report results of new research.

(2) It does not evaluate or make any recommendation on whether any subspecies that is found to be valid should be used as a management unit, as the object of management action, or preferred to an alternative legal classification for protection, such as a distinct vertebrate population segment recognized under the Endangered Species Act (USFWS and NOAA 1995). Suitability of a subspecies as a unit for any of these purposes requires further, separate analysis weighing legal and policy considerations.

(3) It is not a review of the conservation status of any of the taxa considered; as such, it does not review threats to or the population status of any entity; and

(4) It represents the views of the authors and not necessarily those of the U.S. Fish and Wildlife Service.

### **Species concepts and criteria**

There is no single species concept or set of criteria accepted by all taxonomists. Phylogenetic relationships and reproductive relationships represent two major approaches to defining species, but there have been attempts to combine them in identifying species and species limits. Brief descriptions of some general approaches are provided below.

(1) The biological species concept (BSC): This concept is based on reproductive relationships among populations. The ability to interbreed and realize gene flow between two populations is the indication that they belong to the same species. The concept is most commonly associated with Mayr (1963, 1970), but has antecedents during the development of evolutionary biology in the 20<sup>th</sup> century. According to a brief definition by Mayr (1970), a species is a “reproductively isolated aggregate of interbreeding populations.” A major difficulty in applying the BSC is encountered when assessing allopatric populations, where reproductive relationships cannot be assessed directly and must be inferred from other information.

(2) Phylogenetic species concepts (PSC): Species are identified by their genealogical or phylogenetic relationships and diagnosability. The many variations of these concepts and others are reviewed by Wiley (1981), Avise (2004), and Coyne and Orr (2004).

(3) Avise and Ball (1990, Avise 2004) proposed an integration of concepts from the BSC and PSC into “concordance principles.” Their approach accepts intrinsic reproductive barriers as

basic to species recognition, but incorporates “evidence of concordant phylogenetic partitions at multiple independent genetic attributes.”

Establishing species limits by assessing reproductive barriers according to the BSC and concordance principles does not require absolute reproductive isolation for recognition of species limits and boundaries. Mayr (1942) provided many examples of inter-specific hybridization, including species of *Canis*, and recognized that there may only be occasional hybrids, or areas where hybridization is common within hybrid zones. He recognized that the stability of some hybrid zones was important in maintaining the overall distinctness of the species involved, and that different habitat preferences are among the mechanisms that can contribute to the stability of hybrid zones. Inter-specific hybridization is now known to be more frequent than understood at the time of the development of the BSC, and it is the reproductive fate of hybrid individuals that is important in determining whether introgression is occurring to the extent that the formerly separate gene pools are merging (Coyne and Orr 2004).

(4) The cohesion species concept was proposed by Templeton (1989) to at least partly deal with situations such as those in canids where there is naturally-occurring hybridization among species and reproductive isolation is difficult to evaluate. He provided this definition: “The cohesion species concept is the most inclusive population of individuals having the potential for phenotypic cohesion through intrinsic cohesion mechanisms.” Cohesion mechanisms include promoting genetic identity with gene flow and constraints from selective, ecological, developmental, and historical factors.

(5) Some recent taxonomists (e.g., Sites and Marshall 2004, de Queiroz 2007) have distinguished between species concepts and the operational criteria for empirically determining species limits, or delimiting species. A “separately evolving metapopulation lineage” has been

suggested by de Queiroz (2007) as a feature common to all species concepts, with the criteria from various concepts serving as operational criteria for assessing lineage separation. Operational criteria include fixation of character states, correlated divergence between morphology and genetics or between different genetic marker systems, character divergence, monophyly, diagnosability, ecological divergence, and behavioral differences. Different operational criteria can lead to different conclusions because their necessary properties for species diagnosis develop at different times during the process of lineage divergence and speciation (de Queiroz 2007). Sites and Marshall (2004) and de Queiroz (2007) advocate an eclectic approach that uses the appropriate operational criteria for all available classes of scientific information.

## **Subspecies concepts**

There is no scientific consensus on what constitutes a subspecies, and some authorities (e.g., Wilson and Brown 1953, Zink 2003) have questioned the utility of the subspecies level of classification. Following is a description of various subspecies criteria that have been proposed and applied in the taxonomic literature. Because some criteria are more stringent than others, a putative subspecies may meet the criteria and be recognized following one concept, but found to be invalid under a more stringent concept.

Nowak (1995, p. 394) discussed the standards he used in revising the subspecies of *Canis lupus*: “My investigation largely disregarded such questions [concerning use of very localized characters] and concentrated on general trends in measurable size and proportion that could be evaluated on a continent-wide or worldwide basis. Substantive statistical breaks in such trends, as discussed above, were taken as evidence of taxonomic division.”



184           In *The Mammals of North America* (Second Edition), Hall (1981, p. viii) included the  
185 following in his “Criteria for Species versus Subspecies.” “If crossbreeding occurs in nature at a  
186 place or places where the geographic ranges of two kinds of mammals meet, the two kinds are to  
187 be treated as subspecies of one species. If no crossbreeding occurs, the two kinds are to be  
188 regarded as two distinct, full species.”

189           Mayr (1963, glossary) defined subspecies as: “An aggregate of local populations of a  
190 species inhabiting a geographic subdivision of the range of the species, and differing  
191 taxonomically from other populations of the species.” Mayr (1963, page 348) explains  
192 “differing taxonomically:” ‘Therefore subspecies are to be named only if they differ  
193 “taxonomically,” that is, by diagnostic morphological characters.’ Mayr (1969, p. 190) describes  
194 a quantitative method for determining whether populations differ taxonomically: ‘A so-called  
195 75-percent rule is widely adopted. According to this, a population is recognized as a valid  
196 subspecies if 75 percent of the individuals differ from “all” (= 97 percent) of the individuals of a  
197 previously recognized subspecies. At the point of intersection between the two curves where this  
198 is true, about 90 percent of population A will be different from about 90 percent of the  
199 individuals of population B (to supply a symmetrical solution).’

200           Patten and Unitt (2002, p. 27) define subspecies as: “diagnosable clusters of populations  
201 of biological species occupying distinct geographic ranges.” They do not require that  
202 diagnosability be absolute, but advocate 90 percent separation as a more stringent criterion than  
203 the 75-percent rule.”

204           Avice (2004, p. 362) attempted to incorporate phylogenetic information within a  
205 biological species concept in providing the following guidance on recognizing subspecies:  
206 ‘Within such units [=species], “subspecies” warranting formal recognition could then be

conceptualized as groups of actually or potentially interbreeding populations (normally mostly allopatric) that are genealogically highly distinctive from, but reproductively compatible with, other such groups. Importantly, the empirical evidence for genealogical distinction must come, in principle, from concordant genetic partitions across multiple, independent, genetically based molecular (or phenotypic; Wilson and Brown 1953) traits.’

The most stringent criterion that has been proposed for subspecies recognition is reciprocal monophyly (Zink 2004). Application of a monophyly criterion requires that all individuals in a taxon be genealogically closer to one another than to any individual in another taxon. A number of objections to monophyly as a subspecies criterion have been raised, perhaps foremost is that in phylogenetic classifications it is a species-level criterion and inappropriate for application below the species level (Goldstein et al. 2000, Patten and Unitt 2002). Its application using genetic data is limited to genetic sequences that do not recombine, such as mitochondrial DNA, and therefore usually depends on one type of marker rather than multiple markers that can be tested for concordance, as in the Avise (2004) criterion. In addition, there are many instances of related species that have achieved reproductive isolation but are not reciprocally monophyletic (Avise 2004); it takes many generations (on the order of four to seven times the effective population size) after putative taxa are separated for reciprocal monophyly to be achieved (Hudson and Coyne 2002).

A common feature of all of the above definitions is that they recognize that subspecies are groups of populations, and most recognize that subspecies can be variable and overlap in distinguishing characters to some degree.

## **Limitations of the available data**

Several factors concerning the available scientific information bearing on wolf relationships complicate the assessment of taxonomic relationships.

Wolves have been extirpated over large portions of North America, particularly most of the conterminous United States (Figure 1), so there are large gaps in geographic coverage, particularly for genetic data. Recent studies (discussed in later sections of this review) of DNA markers from museum specimens have attempted to address these gaps, but as yet they represent relatively few individuals.

For evaluating continent-wide patterns of variation and their potential taxonomic implications, it would be ideal to have comprehensive sampling across the landscape. This would allow for more rigorous testing and formulation of evolutionary scenarios, and for application of increasingly sophisticated methods of landscape genetics. Regrettably, sampling of wolf populations is far from even over North America. Sampling patterns can influence the interpretation of the genetic structure of populations and lead to erroneous conclusions (Schwartz and McKelvey 2009). Sampling may be relatively intensive in areas that still have large wolf populations, such as Alaska and northern Canada, but information on other areas may be limited to a few, widely spaced individuals. Some published studies (e.g., Koblmüller et al. 2009a) report results from large data sets, but without sufficiently explicit geographic information to permit the reader to evaluate genetic population structure and interactions among populations.

Comparable sets of data are not available for many areas of taxonomic interest. For example, some areas may have detailed data on microsatellite DNA variation, but lack information about lineage markers (mitochondrial DNA and Y-chromosome variation).

Very few of the reviewed studies were designed to address taxonomic questions. Studies designed for other purposes may not be informative on specific taxonomic issues and the evaluation of putative taxa.

The nature of the available data do not permit the application of many of the subspecies criteria reviewed above. For example, the “75-percent rule” is for individual character analysis, but most available analyses of morphological data for wolves use multivariate statistics that summarize variation in many characters. Furthermore, the available data on a particular taxonomic question comprise a variety of very different types of information that must be integrated. The approach to subspecies of Avise (2004), described above, is the most applicable to the disparate data sets available on wolves.

For decades the subspecies classification of gray wolves, *Canis lupus*, was the 24 subspecies recognized by Hall and Kelson (1959) and Hall (1981), which was largely based on the work of Goldman (1937, 1944). There is very little information for some of these named subspecies, especially from genetic studies. Nowak (1995) has reduced the number of recognized subspecies to five; the subspecies and their geographic ranges recognized by Nowak (1995, 2002) are presented in Figure 2 of this paper.

#### **Approach taken in this review**

*Species limits.* Certain attributes of North American *Canis* present special challenges in evaluating species-level taxa, including the ability of different forms to hybridize under certain conditions. There is also a temporal dimension to wolf relationships; some lineages that maintained their distinctiveness from one another are now in secondary contact where earlier ecological and other factors that had formerly inhibited hybridization have been altered. On the

other hand, there is detailed genetic information from some areas, and information on variation in ecology and behavior is available for explaining historic and modern relationships of different populations.

This review generally follows concordance principles in recognizing species (Avice and Ball 1990, Avice 2004). This is an eclectic approach that seeks to identify species as separate lineages supported by concordant data from various classes of genetic markers, morphometric analysis, behavior, and ecology. This approach is appropriate for North American *Canis* because populations of the putative species are or have been in contact with one another and there is considerable genetic information bearing on reproductive relationships. At the same time, there are extensive data from genetic lineage markers (mitochondrial DNA and Y-chromosome haplotypes) that provide phylogenetic information that needs to be considered to understand the evolutionary history, ancestral condition, and taxonomic relationships of North American wolf populations. Lineage markers are essential for inferring possible pre-contact population differences that became complicated by contact and admixture between formerly separate populations or lineages (Cathey et al. 1998, Hannote et al. 2000, Feng et al. 2000, Pidancier et al. 2006).

As discussed later in this review, North American *Canis* comprise two major lineages or clades: one including most gray wolves, and the other includes eastern wolf, red wolf, and coyote. The species-level taxonomic implications of these two clades is first evaluated. With respect to *Canis lupus*, the most controversial question about its species limits raised subsequent to Goldman's (1937, 1944) consolidation of various North American species names (discussed in the later section on taxonomic background) under *C. lupus* is the proposal that the eastern wolf, *Canis lupus lycaon* is a separate species and outside the species limits of *C. lupus*. Reproductive

relationships of populations representing the eastern wolf and other *C. lupus*, in the narrow sense, can be assessed because there has been interbreeding and admixture, and data are available from genetic markers suitable for evaluating the extent of admixture and alternative interpretations of the origin of coyote clade markers within the eastern wolf. Whether the red wolf is within the species limits of *C. lupus* is less controversial.

Because there has been at least historical contact and informative genetic data are available, a similar approach can be taken in evaluating species limits within the coyote clade, with one exception. Extirpation, limited geographic sources for genetic data, and uncertainties about historical distributions do not permit a meaningful assessment of reproductive relationships between eastern wolf and red wolf. For determining species-level relationships between these two putative taxa, they must essentially be treated as allopatric populations, and operational criteria other than reproductive relationships must be applied. Some data are available for assessing non-reproductive criteria such as correlated divergence between morphology and genetics, different genetic marker systems, character divergence, cohesion, monophyly, and diagnosability.

As discussed previously under species concepts, evaluation of reproductive relationships following concordance principles and the biological species concept does not require absolute reproductive isolation for recognition of species-level differences. If absolute isolation were required, all North American *Canis*, wolves and coyotes, would be considered one species, or a “ring species” (Mayr 1942), because all component taxa are linked by evidence of interbreeding, although the incidence of such effective introgression can range from ancient and very rare in some cases to modern and ongoing in others. Requirement of absolute reproductive isolation as a standard for species delimitation would mean that coyotes and wolves in eastern North

America are the same species, despite marked differences in morphology, ecology, behavior, and genetic composition. Such a single-species classification would obscure evolutionarily important diversity. Partially because of the power of new systems of molecular genetic markers, incomplete reproductive isolation between recognized species is now known to be common, especially in certain groups (Grant and Grant 1992, Schwenk et al. 2008), and examples include familiar species such as some species pairs of Darwin's finches (Grant and Grant 2006), mallards (*Anas platyrhynchos*) and American black duck (*Anas rubripes*) (Mank et al. 2004), and Canadian lynx (*Lynx canadensis*) and bobcat (*Lynx rufus*) (Schwartz et al. 2004). The approach of this review following concordance principles will allow full consideration of the available information in evaluating the relationships among populations, the extent of interbreeding, and the likely consequences for recognizing species limits.

*Subspecies of gray wolf.* The initial approach of this analysis was to compare the distributions of the five subspecies of gray wolf (*Canis lupus*) identified in the most recent taxonomic revision of that species (Nowak 1995) with available scientific information on inter-population differences and relationships, primarily from studies of genetic variation. The scientific support for the validity of these taxa is evaluated by summarizing the results of each scientific study with a bearing on the taxonomic standing of a particular subspecies and relationships to other subspecies. A subspecies is found to be supported when the geographic distribution of specific genetic markers coincides with its general distribution based on morphological analyses. The distribution of mitochondrial DNA haplotypes associated with different clades, and presumed Old World sources, is of particular interest. This approach most closely resembles the subspecies definition and criteria of Avise (2004) in that concordance between morphometric

and genetic data is taken as evidence for the validity of a subspecies. This approach should not be interpreted as *a priori* acceptance of Nowak's (1995) subspecies classification. This is a taxonomic evaluation, and it is reasonable to consider the most recently revised taxonomy, which is that of Nowak (1995). Based on additional information, primarily genetic data, this review comes to conclusions that differ from Nowak (1995) on some taxonomic interpretations, and subspecies that Nowak (1995) reduced to synonymy are considered when additional data suggest that they deserve further evaluation.

### **Biology of the Species**

This section first provides summaries of the taxonomic history of *Canis* and some aspects of ecology and behavior that have been identified as important in explaining population structure. The major part of this section comprises summaries of individual studies of morphology and genetics that are relevant to the evaluation of the taxonomy of North American wolves.

### **Taxonomic background on wolf species and subspecies**

*History and overview of the genus Canis.* This brief summary of the global history of *Canis* is based primarily on the reviews by Nowak (1979) and Kurtén and Anderson (1980).

The genus *Canis* originated in North America by the middle Pliocene. Members of the genus probably began colonizing the Old World soon (in geological time) thereafter, where their descendents include the modern species *Canis adustus* (side-striped jackal; range: Africa), *Canis aureus* (golden jackal; Eurasia and North Africa), *Canis mesomelas* (black-backed jackal; Africa), and *Canis simensis* (Ethiopian wolf; Ethiopia) (Wilson and Reeder 2005).



Diverse lineages and species of *Canis*, including coyote, evolved in North America during the Pliocene and Pleistocene. Members of one of these North American lineages entered Eurasia in the early Pleistocene and eventually evolved into *Canis lupus*. Gray wolf later entered North America, where its fossils first appear in middle Pleistocene deposits. More than one invasion of North America by Eurasian *C. lupus* has been suggested based on morphological data and biogeographical reconstruction (Nowak 1983, 1995). This has been confirmed by genetic data that support at least three separate invasions from different Eurasian lineages to explain the patterns of genetic variation observed in modern *C. lupus* of North America (Vilà et al. 1999). An additional gray wolf lineage known only from Pleistocene individuals preserved in permafrost in Alaska became extinct without leaving modern descendents (Leonard et al. 2007).

Gray wolf is the only species of *Canis* with a range that includes portions of both Eurasia and North America. Gray wolves had very large historic distributions in both areas: throughout all of Eurasia except southeast Asia, and in North America from the Arctic to Mexico. Based on morphometric analyses, Nowak (1995) recognized five subspecies of *C. lupus* in North America. Of the 12 subspecies previously recognized in Eurasia, three were considered synonyms of other subspecies, and there was insufficient material to statistically evaluate four others. None of the modern recognized subspecies occurs or occurred in both Eurasia and North America.

*Canis in North America.* The first published name of a taxon of *Canis* from North America is *Canis lycaon*, which was published in 1775 based on the earlier description and illustration of an individual that was thought to have been captured near Quebec (Goldman 1937). The next North American taxon names were published when Say (1823) named and described *Canis nubilus* based on wolves he observed in eastern Nebraska. The coyote (*Canis latrans*) was also

described by Say (1823) from the same Nebraska locality at the same time, and his observations appear to be the first that clearly distinguish between wolves and coyotes. These and the other 28 available scientific names subsequently described from North American wolf taxa are listed in chronological order in Table 1. Wolf taxa were originally described as either subspecies (sometimes indicated as a trinomial “variety”) or species through 1912. Thereafter, all new taxa were described as subspecies. Most available wolf names were subspecies described in the 1930s and 1940s.

Earlier names were published as individual descriptions in various publications, including reports of exploratory expeditions. The first attempts to compile consolidated treatments of North American wolf taxa were the incomplete reviews by Miller (1912) and Pocock (1935). Goldman (1937, 1944) provided the first truly comprehensive treatment of North American wolf taxa, but did not include coyote. Goldman’s classification addressed uncertainties in the nomenclatural history of the taxa, and included many subspecies, many of which he himself described. A particularly notable feature of Goldman’s classification was recognition of two species of wolves in North America: red wolf (as *Canis niger*, now known as *Canis rufus*) occupying parts of the southeastern United States, and gray wolf occupying the remaining range of wolves in North America.

For decades the subspecies classification of gray wolves was the 24 subspecies recognized by Hall and Kelson (1959) and Hall (1981), which was based primarily on Goldman’s (1944) classification. The range map of subspecies from Goldman (1944, Figure 14) is reproduced here as Figure 3. Nowak’s morphometric studies led him to propose the reduction in number of the North American subspecies of gray wolf from the 24 previously recognized to five (Table 2, Figure 2). Brewster and Fritts (1995) summarized controversies concerning North

American wolves, with a concentration on western North America, based on the genetic and morphometric information available at that time.

The following sections provide more detailed taxonomic background on individual North American wolf taxa.

*Red wolf. Canis rufus* has usually been recognized as a species separate from gray wolf (Goldman 1937, 1944; Nowak 1979; Hall 1981; Baker et al. 2003), but is sometimes considered a subspecies of gray wolf (Lawrence and Bossert 1967, Wilson and Reeder 2005). Nowak (1979, p. 85) has noted that the name *Canis niger* (Bartram, 1791), which was used by Goldman (1944) and some other authors for this species, was determined by the International Commission on Zoological Nomenclature to be unavailable for nomenclatural purposes.

The three subspecies of red wolf recognized by Goldman (1937, 1944) and Hall (1981) are listed in Table 3 along with their general historical ranges. The red wolf survives only in captive breeding facilities and reintroduced populations in North Carolina (Phillips et al. 2003). All surviving individuals are descendants of red wolves captured within the historic range of the subspecies *C. r. rufus*, so that nearly all genetic data on *C. rufus* are derived from individuals attributable to that subspecies. Because all living red wolves are derived from this single subspecies, the subspecies classification will not be treated in this review.

*Gray wolf subspecies.* The more expansive subspecies classification of Goldman (1944) Hall and Kelson (1959), and Hall (1981) as well as the simplified classification of Nowak (1995) are presented in Table 2. The recognized names in this table will be used in the following discussion of their taxonomic treatment.

The taxonomic status of the eastern wolf is controversial. It has been considered a full species, *Canis lycaon* (Wilson et al. 2000, Baker et al. 2003); a subspecies of gray wolf, *Canis lupus lycaon* (Goldman 1937, 1944; Nowak 1995, 2002, 2003); the result of coyote introgression into gray wolf (Lehman et al 1991); the same species as the red wolf (Wilson et al. 2000); or a result of hybridization between red wolf and gray wolf (Nowak 2002, 2003, 2009).

Goldman (1937, 1944) considered the eastern wolf to be a subspecies, *C. l. lycaon*, found from southern Quebec and Ontario to Minnesota. He also described *Canis lupus labradorius* from northern Quebec, stating that it was similar to eastern wolf, but larger. Goldman (1944) mapped a geographic range for eastern wolf that extended from northeast Florida to eastern Minnesota and States to the east, and Ontario and southern Quebec in Canada (Nowak [2002] now places the Florida location at “vicinity of Miami”). He recognized the following three neighboring subspecies:

*Canis lupus nubilus* (plains wolf): bordering eastern wolf on the west from southern Illinois to Manitoba. Goldman (1944, p. 444) notes, however, “[s]pecimens from eastern Minnesota and Michigan seem more properly referable to *lycaon*, but relationship to *nubilus* is shown in somewhat intermediate character.”

*Canis lupus hudsonicus* (Hudson Bay wolf): bordering eastern wolf range west of Hudson Bay in northern Manitoba.

*Canis lupus labradorius* (Labrador wolf): bordering eastern wolf range in northern Quebec.

The general ranges of these three subspecies were followed by Hall and Kelson (1959) and Hall (1981).

Relevant to the boundary of eastern wolf, Standfield (1970) observed that a larger type of wolf occurred in boreal areas north of Lake Superior, and a smaller wolf occurred in deciduous forests to the east and southeast. Mech and Frenzel (1971) suggested that some wolves in northeastern Minnesota were *C. l. nubilus* based on color forms of black and white that Goldman (1944) had reported as common for *C. l. nubilus* but not for eastern wolf.

Corresponding to varying views on its taxonomic standing, the geographic range of eastern wolf has not been consistently defined: The very broad ranges in eastern North America recognized by Goldman (1944) and Hall (1981) were reduced by Nowak (1995) to a much smaller area centered on southern Ontario and Quebec.

Wilson et al. (2000) proposed that eastern wolf be restored to full species status based on its genetic distinctness from gray wolf. They also proposed that it is the same species as red wolf, and that this combined taxon be recognized under the earlier published name, *Canis lycaon*.

The Mexican wolf was described by Nelson and Goldman (1929) as *Canis nubilus baileyi*, with a type locality identified in Chihuahua, Mexico. Its distribution was described as: “Southern and western Arizona, southern New Mexico, and the Sierra Madre and adjoining tableland of Mexico as far south, at least, as southern Durango.” The specimens examined included a wolf from Kendrick Peak on the Coconino Plateau in north-central Arizona and several individuals from the Sacramento Mountains, New Mexico.

Goldman (1937) re-classified Mexican wolf as a subspecies of the species *Canis lupus*, so that its name became *Canis lupus baileyi*. He also included the Kendrick Peak, Arizona, specimen with his newly described *Canis lupus mogollonensis* (Goldman 1937, 1944), which shifted the northern limits of Mexican wolf further south in Arizona. Goldman (1933) mapped

the eastern boundary of Mexican wolf as contiguous with the western boundary of *C. l. monstrabilis* in southeastern New Mexico, far western Texas, and eastern Mexico. This view of the boundary of Mexican wolf in Arizona was followed by Hall and Kelson (1959), Nowak (1979), and Hall (1981).

Based on their morphometric analysis of wolves of the southwestern United States and adjacent Mexico, Bogan and Mehlhop (1983) referred wolves formerly assigned to *C. l. mogollonensis* and *C. l. monstrabilis* to Mexican wolf.

Nowak (1995) included Mexican wolf as one of five North American subspecies that he recognized in his revision of gray wolf subspecies, but contrary to Bogan and Mehlhop (1983), referred *C. l. mogollonensis* and *C. l. monstrabilis* to *C. l. nubilus*.

Arctic wolf, *Canis lupus arctos*, was described from skulls from Melville Island and Ellesmere Island in the Canadian Arctic (Pocock 1935). *Canis lupus arctos* was subsequently recognized by Goldman (1944), Hall and Kelson (1959), and Hall (1981).

Based on morphometric analysis, Nowak (1995) placed *Canis lupus orion* and *Canis lupus bernardi* as synonyms of *C. l. arctos*. Both were recognized as separate subspecies by Goldman (1944), Hall and Kelson (1959), and Hall (1981). The range of Nowak's expanded *C. l. arctos* generally includes Greenland and all the Canadian Arctic Islands, except Baffin Island, which was included within the range of *Canis lupus nubilus*.

*Canis lupus nubilus* and *Canis lupus occidentalis* are the most geographically widespread of the five subspecies of gray wolf recognized by Nowak (1995), and share long and complex borders. They also have the largest synonymies of the five species, with 11 synonyms recognized for *C. l. nubilus* and six for *C. l. occidentalis* (Table 3).

## **Summary of relevant literature**

*Ecology, behavior, prey and habitat.* Based on its extraordinarily large historical range, gray wolf has been one of the most successful large, terrestrial vertebrate species to occupy the earth. In North America, it occurred in nearly all natural types of habitats from the Arctic to tropical Mexico. This section summarizes aspects of habitat, prey, and behavior that have been invoked to explain certain patterns of variation in wolves.

In North America, wolves can be successful in all natural habitat types (Carroll 2003, Carroll et al. 2003, 2006, Oakleaf et al. 2006), except the most extreme deserts. Differences in habitat have been correlated with variations in behavior, including migration and prey selection. For example, Kolenosky and Stanfield (1975) have described variation in Ontario wolves, where larger wolves of boreal forests specialize on moose (*Alces americanus*) and caribou (*Rangifer tarandus*) as prey, while smaller wolves in deciduous forest habitats specialize on white-tailed deer (*Odocoileus virginiana*). Carmichael et al. (2001) and Musiani et al. (2007) have proposed that differences in migratory behavior and prey have influenced genetic differences between wolves that follow migratory caribou on the tundra and wolves that prey on more sedentary caribou in forested areas. These studies are further discussed in the following sections on morphology and genetics. North American wolves specialize on large mammals as prey. In addition to caribou, moose, and deer, they feed on muskox (*Ovibos moschatus*), American bison (*Bison bison*), elk (*Cervus elaphus*), mountain sheep (*Ovis* sp.), and mountain goat (*Oreamnos americanus*). They also consume domestic ungulates: cattle, sheep, and goats. Mech (1974) indicated that American beaver (*Castor canadensis*) are the smallest prey to be consistently reported for wolves. Lagomorphs and smaller rodents are consumed opportunistically.

Pack structure, thought to be key to the ability of gray wolves to catch and kill large ungulate prey, has been alternatively explained by kin selection and benefits of sharing prey with offspring. Wolves are cursorial animals capable of traveling long distances (e. g., Mech 1987, Musiani et al. 2007). Wolves can range from one habitat type to another, and are capable swimmers (Mech 1974). Even where rivers are too wide for regular crossing, wolves can cross when sufficient ice forms (Carmichael et al. 2001). Mountains are generally not a barrier to wolf movement, and in some portions of their range, mountains are where wolves are most common. Particularly steep and high ranges have, however, been invoked to explain the partial isolation and genetic divergence of coastal wolves in southeastern Alaska and British Columbia from inland populations (Weckworth et al. 2005, 2010).

The factors briefly discussed above have been invoked as ad hoc explanations to explain certain patterns of morphological or genetic variation in wolves. Wolves are large, vagile animals that have few natural limitations in areas that they can colonize. There do not appear to be any general rules predicting where wolves will be found or where geographic variation can be expected. There can, however, be combinations of behavior, prey, and habitat that can contribute to the partial isolation of populations, and foster interpopulational differences. Instances will be identified in the following sections summarizing studies on morphology and genetics.

*Summaries of relevant studies.* Studies of the morphology and genetics of North American *Canis* are summarized in the following sections, which are organized into categories by the type of data or genetic marker used. Each discussion of a category of information is followed by a brief summary of areas of agreement or disagreement among the studies. The species and



subspecies names used in these summaries are those used by the authors of these papers; use of these names in these sections should not be interpreted as our acceptance of any author's nomenclature. Similarly, the conclusions stated in the summaries are those of the author(s) of each paper being summarized. Our analysis and conclusions are presented in the Analysis and Discussion section later in this paper.

*Morphology.* Nearly all recent studies of morphological variation among taxa of North American *Canis* employed the multivariate statistical methods of principal components analysis, discriminant function analysis, or both. Discriminant function analysis requires that groups be known or distinguished a priori by other data and is most useful for evaluating the affinity of unknown individuals (e. g., Maldonado et al. 2004), and for identifying characters most useful for distinguishing among groups (James and McCulloch 1990). Its use in determining inter-group differences has been criticized because the a priori identification of groups is sometimes based on the same data that are used to generate the distance measures, which introduces circularity into the analysis, and differences that can readily discriminate between groups may be relatively small and of little biological significance (James and McCulloch 1990, Lance et al. 2000). For measuring inter-group differences for taxonomic analysis there are more objective methods, such as principal components analysis; an example is discussed later in the analysis of the standing of the Mexican wolf, *Canis lupus baileyi*. In the following summaries of statistical studies of morphology, the a priori groups subjected to discriminant function analysis are identified.

Jolicoeur (1959) carried out an analysis of the distribution of coat color and bivariate and multivariate discriminant function analyses of skull features of 499 *C. lupus* from western

Canada. Samples were grouped for the discriminant function analyses by regions within Canadian provinces. He found general patterns of the incidence of pale pelage increasing towards the Arctic, and skulls trending from shorter and broader in the northeast to longer and narrower to the southeast portion of the study area. Although the study was not framed in a taxonomic context, he suggested that “far too many subspecific designations are now in use,” referring to the classification of Goldman (1944). A notable result was that the samples from Vancouver Island were more like individuals from further north than to wolves on the neighboring mainland of British Columbia (Jolicoeur 1959, p. 297).

In a study of North American canids using discriminant function analysis, Lawrence and Bossert (1967) included a comparison of groups classified as “*Canis lupus*,” *Canis rufus* (under the name *Canis niger*), and *Canis lupus lycaon*. The *C. lupus* sample was found to be intermediate between *C. l. lycaon* (from Algonquin Provincial Park, Ontario) and *C. rufus*. The validity of this result is difficult to evaluate because the geographic source of the *C. lupus* sample of 20 wolves was not indicated and “large individuals were avoided,” thus biasing the sample. Another factor limiting comparison to subsequent morphometric studies was the determination of character values relative to the length of the skull rather than actual measurements. This removed size as a character, and size is generally considered an important character in evaluating variation among wolves in North America (Kolenosky and Stanfield 1975, Nowak 1979, Schmitz and Kolenosky 1985).

Kolenosky and Standfield (1975) studied skulls of two types of wolves from within the Ontario range of *C. l. lycaon* (as broadly defined by Goldman [1944]) using discriminant function analysis and provided comparisons of whole body mass (n = 594) and coat color (n = 1404). The samples included 105 “boreal-type” (from areas of boreal forests) wolves and 122

“Algonquin-type” (from deciduous forest regions). Over 75% of boreal-type skulls could be distinguished from those of the Algonquin, or eastern wolf, type. Boreal males averaged 34.5 kg compared to 27.5 kg for Algonquin-type males. Coat colors of Algonquin-type individuals were nearly all gray-fawn, while boreal-type were mostly gray-fawn, but many individuals were black or cream. They suggested that the size and color of boreal-type individuals were more like *C. l. nubilus*, and that the two Ontario forms may not be interbreeding. They associated these types of wolves with different ungulate prey species, with the larger boreal-type wolves preying on moose and caribou, and the Algonquin-type wolves on white-tailed deer.

Skeel and Carbyn (1977) performed principal components and discriminant function analyses on 311 wolf skulls from widely-spaced localities in central and northern North America, including several Canadian national parks. Samples were grouped by subspecies or by park for discriminant function analysis. For addressing the question of the relationship of *C. l. lycaon* to other *C. lupus*, the relevant samples were from the southwestern corner of Ontario (referred to *C. l. lycaon*), historical *C. l. nubilus* (primarily from the north-central United States), *C. l. hudsonicus* from southeast Northwest Territories, and *C. l. griseoalbus* from Manitoba and Saskatchewan. The last two subspecies were later treated as synonyms of *C. l. nubilus* and *C. l. occidentalis*, respectively, in Nowak’s (1995) taxonomic revision, which is discussed later. Strong geographic patterns were not obvious, except that *C. l. lycaon* is generally more similar to *C. l. nubilus* and its synonyms than to samples from further to the northwest, which would probably be attributable to *C. l. occidentalis* in Nowak’s (1995) revised classification. Skeel and Carbyn’s (1977) general conclusion was that there is large overlap in characters among individuals, but that wolves in “boreal-subalpine forest regions” are larger.

In comparing *C. l. occidentalis*, primarily from Canadian national parks, to other subspecies of *C. lupus*, three-dimensional principal components plots for males and females (Skeel and Carbyn 1977, Figure 2 and Figure 3, respectively) showed a clear separation of *C. l. occidentalis* (codes W, R, J, and P in the figures) from a grouping that included *C. l. nubilus*, *C. l. hudsonicus*, and *C. l. irremotus* (the latter two are synonyms of *C. l. nubilus* in Nowak's [1995] classification) samples on the first principal component, which can be attributed to the larger size of *C. l. occidentalis* (Skeel and Carbyn's Figure 2 is reproduced here as Figure 4). Discriminant function plots (Skeel and Carbyn 1977, Figure 4 and Figure 5) showed polygons for *C. l. occidentalis* and *C. l. hudsonicus* overlapping minimally, and more substantially between *C. l. occidentalis* and both *C. l. nubilus* and *C. l. irremotus*. Further analysis with clustering (Skeel and Carbyn 1977, Figure 6) showed discontinuity between *C. l. occidentalis* and the other subspecies, which grouped closer together. A multi-dimensional scaling analysis (Skeel and Carbyn 1977, Figure 7) also separated *C. l. occidentalis* and *C. l. nubilus* samples, with the exception that Wood Buffalo National Park *C. l. occidentalis* were closest to *C. l. hudsonicus* from Northwest Territories on Hudson Bay; these are the two northern-most areas included in the study.

Nowak (1979) reviewed the taxonomic history and carried out discriminant function analyses of North American *Canis*. Groups for the initial analysis were gray wolf, red wolf, coyote, and domestic dog. Various samples representing specific populations, time periods when specimens were collected, or extinct species of *Canis* were then plotted and compared to the positions of the samples from the initial analysis. Of particular relevance to the present review was his treatment of the relationships and taxonomic standing of *C. rufus* and *C. l. lycaon*.

Nowak (1979, p. 87) found little statistical overlap between early (before extensive introgression by coyotes) *C. rufus* (n = 74 males, 55 females) and western *C. lupus* (n = 233 males, 146 females) skulls, although a few specimens were difficult to assign. He noted (p. 29) that *C. rufus* resembled *C. l. lycaon* more than it did any other subspecies of *C. lupus*.

Nowak (1979, Figure 7) found substantial, but not complete statistical overlap between skulls of *C. l. lycaon* and other *Canis lupus* from western North America as *Canis lupus lycaon* individuals were generally smaller. Nor were the boreal type (Ontario type of Standfield [1970]) wolves of Michigan, Minnesota, Wisconsin, and western Ontario and the deciduous type (Algonquin type of Standfield [1970]) of southeastern Ontario and southern Quebec sharply delineated. Some characters in wolves from the western range of *C. l. lycaon* were found to be intermediate between the eastern *C. l. lycaon* and *C. l. nubilus* from the Great Plains (Nowak 1979, p. 20), thus lending some support to Mech and Frenzel's (1971) suggestion that some eastern Minnesota wolves were *C. l. nubilus*. Nowak's (1979, p. 21) general conclusion was that individuals that he referred to *C. l. lycaon* were no more distinctive than other subspecies of *C. lupus*.

Nowak (1983, Figure 6) performed a preliminary bivariate analysis of skulls of various subspecies of North American *C. lupus*, generally following Goldman's (1944) classification. The two studied characters generally reflect the length and width of skulls. This analysis indicated a cline in the two characters, with size increasing from south to north in central North America, but with a break or discontinuity at approximately the U.S.-Canada border in central North America that divided southern and northern *C. lupus*.

Based on these data, he proposed new geographic groupings of North American wolves. In this scheme (Nowak 1983, Figure 7b), wolves from Minnesota, Wisconsin, the Upper

Peninsula of Michigan, and southwestern Ontario were grouped with southern wolves of the U.S. Great Plains to the west, rather than with *C. l. lycaon* as in Goldman (1944). The Hudson Bay sample (*C. l. hudsonicus*) was also grouped with the southern wolves. The sample of *C. l. columbianus* of far western Canada was intermediate but closer to the northern group.

Skulls from the Canadian Arctic Islands (subspecies *Canis lupus arctos* and *C. l. bernardi*) were outliers to this general north-south trend in overall size and were distinguished by having skulls that were wide relative to their length. Nowak (1983, Figure 7a, b) suggested that at the maximum extent of Pleistocene glaciations, the ancestors of *Canis lupus arctos* were isolated north of the ice sheet in a refugium in northern Greenland, and then spread westward to the Arctic Islands following withdrawal of the glaciers. He also speculated that *C. l. bernardi* and *C. l. orion*, an Arctic subspecies from Greenland, may have declined and their former ranges occupied by *C. l. arctos*.

Based on this information and historical factors, Nowak (1983, Figure 7a, b) suggested a “hypothetical” new evolutionary scenario and configuration of subspecies. At the maximum extent of Pleistocene glaciations, south of the ice sheet were wolves that had already evolved in or colonized North America: *C. l. lycaon* in the east, *C. l. baileyi* in the southwest, and a “southern group.” The southern group corresponds to *C. l. nubilus* in Nowak’s (1995) eventual reclassification. North of the ice sheet were the ancestors of *C. l. arctos* in the east, and a “northern group” isolated to the west in Alaska. This northern group corresponds to *C. l. occidentalis* in Nowak’s later revision.

Bogan and Mehlhop (1983) reported the results of principal component and discriminant function analyses of 253 wolf skulls from Mexico and the southwest region of the United States, including specimens from Kansas, Oklahoma, Texas, Colorado, New Mexico, and Arizona.

Samples were grouped by subspecies according to Goldman's (1944) classification. They considered *Canis lupus mogollonensis* and *Canis lupus monstrabilis* to be synonyms of *C. l. baileyi* based on broad morphological overlap of their skulls (Bogan and Mehlhop 1983, Figure 2 and Figure 3; Figure 2 is reproduced here as Figure 5). This effectively expanded the range of *C. l. baileyi* north to central Arizona and New Mexico, and east into central Texas. They also acknowledged that specimens previously referred to *C. l. mogollonensis* represented intergrades between *C. l. baileyi* and *C. l. youngi*, the subspecies then recognized for the southern Rocky Mountains, and did not detect the abrupt break between *C. l. baileyi* and *C. l. mogollonensis* noted by Goldman (1944). They recognized three subspecies in the area covered by their study: *C. l. baileyi*, *C. l. youngi*, and in the Great Plains, *C. l. nubilus*.

Schmitz and Kolenosky (1985) reported clinal variation in *C. l. lycaon* (following Goldman's [1944] delineation of the range of that subspecies) in Ontario based on discriminant function analysis of skull and body characters. Canids were assigned to six groups for the discriminant function analysis: boreal, Algonquin, and southern Ontario wolves; and Algonquin, southeast Ontario, and southwest Ontario coyotes. From larger wolves in boreal regions in the north, size declined to the smaller, Algonquin-type wolves in southern Ontario. They found that the boreal wolves more resembled Minnesota wolves than Algonquin wolves in body characters, but boreal more resembled Algonquin in skull characters. Their explanation was that the resemblance between boreal and Minnesota wolves (which they viewed as derived from *C. l. nubilus*) was owing to convergence based on similar prey size, and that resemblance between boreal and Algonquin in skull characters was owing to taxonomic affinity and clinal variation within *C. l. lycaon*.

Nowak (1995) performed discriminant function analyses using 580 male skulls grouped by subspecies as delineated by Hall (1981), with *C. l. lycaon* further divided into three groups: Minnesota, western Ontario, and Algonquin Provincial Park. Based on statistical distances ( $D^2$  of Mahalanobis), Nowak (1995, Figure 20) revised the subspecies taxonomy of North American *Canis lupus* in a manner generally consistent with the geographic groupings that he had proposed in his 1983 paper and reduced the number of subspecies from the 23 recognized by Goldman (1944) to five. In this classification, most of the North American range of *C. lupus* is occupied by *C. occidentalis* and *C. nubilus*, which corresponded to the respective northern and southern groups identified earlier (Nowak 1983). The other three subspecies had smaller ranges on the periphery of the North American range of *C. lupus*: *C. l. lycaon* in the east, *C. l. baileyi* in the southwest, and *C. l. arctos* in the islands of the High Arctic.

Nowak's (1995) analysis included 105 individuals of *C. l. occidentalis* and 119 individuals of *C. l. nubilus*. An additional 46 specimens from subspecies that he included as synonyms of *C. l. nubilus* were also included in the study. Statistical distances and discriminant function plots (Nowak 1995, Figure 5, Figure 7; the latter is reproduced here as Figure 6) separate *C. l. nubilus* and *C. l. occidentalis*. Polygons on the discriminant function plots overlapped, but were mostly non-overlapping, with *C. l. occidentalis* being larger than *C. l. nubilus*. *Canis lupus ligoni* (attributed by Nowak to *C. l. nubilus*) of coastal southeastern Alaska was intermediate. Nowak (1995, p. 383) acknowledged that *C. l. ligoni* has probably been affected by hybridization with *C. l. occidentalis*, but that it is partly isolated from *C. l. occidentalis* to the east by "mountains, glaciers, and waterways," and has closer statistical distance to *C. l. nubilus* to the south. He also found that samples from coastal British Columbia, including Vancouver Island, were closer to *C. l. nubilus*.



Nowak (1995, p. 386, Figure 10) found that *C. l. hudsonicus* (a synonym of *C. l. nubilus* in his classification), found west of Hudson Bay, was within the statistical limits of *C. l. nubilus*, although it overlapped with the discriminant function polygon for *C. l. occidentalis*. He suggested that Skeel and Carbyn's (1977) finding that *C. l. hudsonicus* was closer to *C. l. occidentalis* from Wood Buffalo National Park may have involved inclusion of females, which are smaller, in the male sample of the latter. He also suggested that habitat and prey preferences may contribute to differences between *C. l. hudsonicus* and *C. l. occidentalis* and their coexistence without merging or displacement, stating that *Canis lupus occidentalis* is mainly in the taiga in this area, while *C. l. hudsonicus* is mainly in the less productive tundra.

The range delineated for *C. l. lycaon* included southeastern Ontario and southern Quebec. Areas formerly considered by Goldman (1944) to be within the western range of *C. l. lycaon* (Minnesota, the Upper Peninsula of Michigan, Wisconsin, and southwestern Ontario) were included within the range of *C. l. nubilus*. Nowak (1995, Figures 5, 10) based this on the low statistical distance between Minnesota wolves and historical *C. l. nubilus*, and on the occurrence of individuals from western Ontario within the range of variation of his southern group, which is equivalent to *C. l. nubilus*. *Canis lupus lycaon* from or near to Algonquin Provincial Park overlapped to some degree with the western Ontario specimens, but were mostly outside the polygon describing variation in *C. l. nubilus*.

Nowak (1995, Figures 4, 8) recognized *C. l. baileyi* as a subspecies, but did not adopt Bogan and Mehlhop's (1983) inclusion of *C. l. mogollonensis* and *C. l. monstrabilis* as its synonyms. These different interpretations are discussed later in the "Analysis and Discussion" under *C. l. baileyi*.

The geographical ranges of *C. l. baileyi* and other subspecies of *C. lupus* may never have had definite or stationary boundaries. Nowak (1995, p. 385) suggested that *C. l. baileyi*, “regularly dispersed into the range of populations to the north and vice versa.” He also suggested that extirpation of more northern wolves facilitated the dispersal of *C. l. baileyi* from Mexico to areas formerly occupied by other subspecies.

*Canis lupus arctos* partially overlapped both *C. l. nubilus* and *C. l. occidentalis* on Nowak’s (1995, Figure 9) discriminant function plot for some northern wolves. He recognized *C. l. arctos* as a subspecies based on these results, along with the observation that their large carnassial teeth were “the most consistent distinguishing character.” He included *C. l. bernardi* in *C. l. arctos* based on this character, and included *C. l. orion* based on examination of two specimens and reported free movement (Dawes et al. 1986) of wolves between the northern Greenland range of *C. l. orion* and the Ellesmere Island portion of the range of *C. l. arctos*.

The relatively small size of *C. l. baileyi* and *C. l. lycaon* invites their comparison with *C. rufus*. Nowak’s (1995) *C. rufus* sample was selected to exclude specimens that reflected hybridization with coyotes, *Canis latrans*. These skulls were a series collected before 1930 in southern Missouri, and another collected before 1940 from southeastern Texas to Florida. They were compared with Algonquin Provincial Park *C. l. lycaon* from southeastern Canada and his “southern group” of gray wolves, which is equivalent to *C. l. nubilus* and includes Minnesota wolves collected after 1960. Nowak’s (1995, Figure 11) discriminant function analysis of these samples indicated that the areas of the discriminant function plot occupied by the Algonquin *C. l. lycaon* and *C. rufus* individuals approach one another. He suggested the possibility that coyote hybridization could have contributed to the closeness of *C. l. lycaon* to *C. rufus*. He observed that there were too few specimens to evaluate whether *C. l. lycaon* and *C. rufus* intergraded. He

described the historical range of *C. rufus* as extending north to Pennsylvania and that of *C. l. lycaon* as extending south from Quebec and Ontario “to an undetermined point in the eastern United States.”

Nowak’s (1995, Figure 11) discriminant function analysis of *C. rufus* (n = 33) and *C. baileyi* (n = 21) showed no overlap, although one *C. baileyi* individual was just outside the polygon representing *C. rufus* individuals. Statistical distance values (Nowak 1995, Figure 12) showed pronounced differences between 27 *C. l. monstabilis* (a synonym of *C. l. baileyi* according to Bogan and Mehlhop [1983]) and *C. rufus*. Nowak (1995, p. 389) observed that, “there are no specimens to show that the gray wolf was sympatric with unmodified populations of red wolf,” although *C. lupus* was in the vicinity of areas of central Texas where *C. rufus* and coyotes were hybridizing.

In recent papers, Nowak (2002, 2003, 2009) repeated his view that *C. l. lycaon* is a subspecies of *C. lupus* and may be the result of hybridization that occurred when *C. rufus* advanced north into Canada following the last Pleistocene glacial retreat and came into contact with *C. lupus*, more specifically the subspecies *C. l. nubilus*. His discriminant function plots showed that specimens attributed to *C. l. lycaon* (n = 10) are intermediate between western *C. lupus* (n = 97) and *C. rufus* (n = 13) and slightly overlap *C. lupus* (Nowak 2002, figure 8; 2003, figure 9.9).

In his review and discriminant function analysis of eastern *C. lupus* and *C. rufus*, Nowak (2002) explored relationships between *C. l. lycaon* and *C. l. rufus*, as well as *C. lupus* from the western Great Lakes region and some localities in the western United States. Historical *Canis lupus lycaon* (n = 10) from southeastern Canada (Nowak 2002, Figure 8) overlapped only slightly with a “western series” of *C. lupus* (ranging from Minnesota to Idaho and Arizona). A series

797 from the Upper Peninsula of Michigan had substantial overlap with both the eastern *C. l. lycaon*  
798 and the western *C. lupus*. Based on the intermediacy of *C. l. lycaon* from southeastern Canada  
799 between *C. l. nubilus* and *C. rufus*, Nowak (2002, 2003) suggested that *C. l. lycaon* may be the  
800 result of hybridization as *C. l. nubilus* invaded from the west and encountered *C. rufus* invading  
801 from the south following retreat of the terminal Pleistocene glaciations.

802 Nowak (2002, Figure 6) compared Minnesota *C. lupus* skulls (n = 23) taken after 1970  
803 with five series of historical specimens from within the western range of *C. l. nubilus* (n = 78).  
804 The Minnesota wolves overlapped strongly with the series from the northern Rocky Mountains  
805 (of the United States), and less so with the southern Rocky Mountain series. There was slight  
806 overlap with the Nebraska-Kansas-Oklahoma series, and none with the small sample from Texas.

807 To determine whether western *C. lupus* and *C. rufus* became more morphologically  
808 similar where their ranges approach one another, Nowak (2002) performed a discriminant  
809 function analysis that included seven skulls from western Texas. These individuals were from  
810 within the range of *C. l. monstrabilis*, which is considered a synonym of *C. l. baileyi* by Bogan  
811 and Mehlhop (1983), but is considered a synonym of *C. l. nubilus* by Nowak (1995). The  
812 resulting discriminant function plot (Nowak 2002, Figure 6) indicated no morphometric  
813 convergence between the Texas *C. lupus* and pre-1918 *C. rufus* (n = 6), which although based on  
814 a limited sample of *C. lupus* specimens provided no evidence that interbreeding between these  
815 species was occurring in western and central Texas.

816 Nowak (2003, Table 9.3; 2009, Figure 3) described the historical range of *C. l. lycaon* as  
817 extending south to northern and western New York. Nowak (2003, p. 247) noted that a few  
818 Pleistocene specimens indicate that *C. l. baileyi* once extended to Kansas and southern  
819 California.

Nowak (2009) focused on the relationships among wolves of the Great Lakes area in a discriminant function analysis of historical skulls intended to portray patterns of variation before wolves were exterminated from much of the area. In the initial comparison of series representing western *C. l. nubilus* (collected before 1930, n = 27), northern Minnesota wolves (1970-1975, n = 23), and *C. l. lycaon* from Algonquin Provincial Park, Ontario (1964-1965, n = 20), there was no overlap between *C. l. lycaon* and either the northern Minnesota or the western *C. l. nubilus*. Most Minnesota series, however, overlapped the polygon for *C. l. nubilus* (Nowak 2009, Figure 1). Nowak (2009, Figure 15.2) then compared these relationships with two series that are geographically intermediate between Minnesota and Algonquin Provincial Park: Upper Peninsula of Michigan (collected prior to 1966), and a series collected between the Upper Peninsula and Algonquin Provincial Park. Each of these two series overlapped both Minnesota and Algonquin on the discriminant function plots, thus bridging the morphological gap between them.

Mulders (1997) used principal components and discriminant function analyses to study skulls of 525 *Canis lupus* from Canada. He found the wolves of the Canadian Arctic Islands (*C. l. arctos* and *C. l. bernardi*) to be distinct from mainland wolves, but not from each other. He interpreted his findings as supporting recognition of the subspecies *C. l. arctos* and Nowak's (1995) treatment of *C. l. bernardi* as its junior synonym. He found support for *C. l. occidentalis* and *C. l. nubilus*, but with boundaries different from those proposed by Nowak (1995). He characterized *C. l. occidentalis* as "mainland tundra wolves," with a range including Yukon, Northwest Territories, Baffin Island, and portions of Manitoba and western Ontario in the vicinity of Hudson Bay. He characterized *C. l. nubilus* as "central boreal wolves," with a range

south of *C. l. occidentalis*, including eastern British Columbia, Alberta, and nearly all of Saskatchewan; these areas were included within the range of *C. l. occidentalis* by Nowak (1995). Mech and Paul (2008) accepted the recognition of *C. lycaon* as a species separate from *C. lupus*. Based on their analysis of body mass of 950 female and 1006 male adult wolves from across northern Minnesota, they describe an increasing trend in body mass from east to west for both sexes. They concluded that this trend supports the view that the two species meet and hybridize in northern Minnesota. *Canis lupus* in this study would represent *Canis lupus nubilus* according to Nowak's (1995) distribution map.

*Summary of studies on morphology.* Studies with comparable geographic coverage agree in indicating smaller wolves in the Great Lakes region (eastern wolf), with size increasing to the north and west of that region. The study of Lawrence and Bossert (1967) is not comparable because the influence of size was reduced or eliminated by the selection of specimens and the use of ratios rather than direct measurements of skull characters.

Studies involving *Canis lupus bailey* came to different conclusions as to its northern boundary, with Bogan and Mehlhop (1983) favoring a more northerly boundary than Nowak (1995).

*Autosomal genetic markers.* Broad patterns of variation in North American *Canis* were investigated (Roy et al. 1994, 1996) using microsatellite DNA. Samples of *C. lupus* from Minnesota and southern Quebec as well as red wolves (*Canis rufus*) were intermediate between two large multi-dimensional scaling clusters (Roy et al. 1996, Figure 3, which is reproduced here as Figure 7) representing five populations each of *C. lupus* and coyotes (*C. latrans*). The red

wolf samples (n = 40, Roy et al. 1994) were from the captive breeding program (derived from red wolves captured in southeastern Texas and southwestern Louisiana [Roy et al. 1994]) and pre-1940 individuals (n = 16, Roy et al. 1996) from Texas, Oklahoma, Arkansas and Missouri. Roy et al. (1994, 1996) attributed the intermediate placement of these red wolves as well as Minnesota and southern Quebec *C. lupus* to extensive hybridization between *C. lupus* and coyotes. The possibility of an original evolutionary affinity between *C. rufus* and wolves from southern Quebec and Minnesota was not discussed. The *C. lupus* of northern Quebec (n = 20, Roy et al 1994) were closer to western *C. lupus* from Vancouver Island (n = 20), Alberta (n = 20), and Kenai Peninsula, Alaska (n = 19). A neighbor-joining analysis (Roy et al. 1994, Figure 7) of Nei's (1978) genetic distance found northern Quebec wolves to be intermediate between southern Quebec and Minnesota wolves and western *C. lupus* from Vancouver Island, Alberta, Northwest Territories and Alaska. Red wolves were intermediate to southern Quebec and Minnesota wolves and coyotes.

García-Moreno et al. (1996) compared microsatellite DNA variation in *C. l. baileyi* from the captive breeding program with 42 dogs and the gray wolf, coyote and red wolf data presented by Roy et al. (1994). A multi-dimensional scaling plot of the microsatellite data (García-Moreno et al. 1996, Figure 4), showed pronounced separation of *C. l. baileyi* from all the other canids, including other *C. lupus*, although the authors acknowledged that the effects of small founder size and genetic drift in the captive *C. l. baileyi* population may have contributed to their genetic distinctiveness (Paetkau et al. 1997). Red wolves were not included in the Minnesota/Quebec wolf cluster in the MDS plot, but appeared within the confidence ellipse of coyotes (García-Moreno et al. 1996, Figure 4).

A neighbor-joining tree (García-Moreno et al. 1996, Figure 5) based on Nei's (1978) genetic distance displayed the captive *C. l. baileyi* lineages close together on a well-supported branch distinct from other *C. lupus*. The same neighbor-joining tree placed the *C. lupus* samples from Minnesota basal to the clade comprised of *C. l. baileyi*, western gray wolves and domestic dogs while the southern Quebec wolves were basal to the coyote – red wolf clade. Regardless, both the Minnesota and southern Quebec populations were described as “hybridizing gray wolves.”

Several detailed studies have used autosomal microsatellite DNA to characterize the population genetics of wolf recovery in the Northern Rocky Mountains of the United States (Forbes and Boyd 1996, 1997; vonHoldt et al. 2008, 2010). The Montana population is descended from wolves that naturally dispersed from southern Alberta and British Columbia, whereas the Idaho and Yellowstone National Park populations were founded with re-introductions from central Alberta and northern British Columbia. These populations represent the single taxon, *Canis lupus occidentalis* in Nowak's (1995) classification. These studies provide fascinating illuminations of pack structure, reproductive behavior, and migration but are not informative on the taxonomic questions that are the subjects of this paper and will not be further considered.

In a study concentrating on the evolutionary relationships of the wolves of eastern Canada, Wilson et al. (2000) reported microsatellite variation at eight of the loci used by Roy et al. (1994) in comparisons of *Canis* from the vicinity of Algonquin Provincial Park, Ontario (putatively *C. lycaon*), suspected hybridizing (wolf/coyote) wolves from southern Quebec and Minnesota, *C. rufus* from the red wolf captive breeding program, and *C. lupus* from Northern Ontario, Alberta, and the Northwest Territories. This study also described mitochondrial DNA



(mtDNA) control region sequence variation that will be discussed later. Issues of particular interest were the relationship of *C. lycaon* to *C. rufus* and how hybridization with coyotes may have contributed to the genetic similarities observed between them.

Neighbor-joining trees based on Nei's genetic distance (1972) (Wilson et al. 2000, Figures 1 and 2) grouped *C. rufus* with wolves from Algonquin Provincial Park, southern Quebec, and Minnesota, but separate from both coyotes and western *C. lupus*. They concluded that the similarity between eastern Canadian wolves and *C. rufus* was not due to shared introgression from coyotes, because alleles found in the coyote populations were either absent or found at low frequency in *C. rufus* (Wilson et al. 2000, Table 1). At these loci captive *C. rufus* were more similar to Algonquin wolves than to coyotes from Texas, an expected source of introgression into the founders of the captive red wolf population.

Individual assignment tests also indicated Algonquin Provincial Park wolves and *C. rufus* were distinct (probability of identity measure) or nearly distinct (individual index) from Texas coyotes (Wilson et al. 2000, Figure 3). These analyses also supported the mutual distance of Algonquin wolves and *C. rufus* from western *C. lupus* (Wilson et al 2000, Figure 4).

Hedrick et al. (2000) compared major histocompatibility complex variation (MHC) among Mexican wolves and red wolves from the respective recovery programs with western coyotes. They found that Mexican wolves did not share alleles with red wolves or California coyotes, but one allele in the Aragon lineage of Mexican wolves was shared with other gray wolves (Hedrick et al. 2000). Red wolves shared one of their alleles with gray wolves. Further study of MHC variation with additional coyote samples (Hedrick et al. 2002) found that three of the four red wolf haplotypes were shared with coyotes, consistent with their recent history prior to the establishment of the captive population (Wayne and Jenks 1991).

Carmichael et al. (2001) studied microsatellite variation in 491 *C. lupus* from nine locations in the Northwest Territories and Yukon of Canada. Analysis of genetic distance using  $F_{ST}$ , the fixation index of Wright (1951), and assignment tests all indicated restricted gene flow between wolves on different sides of the Mackenzie River ( $F_{ST} = 0.04$ ) and little differentiation among wolves on the same side ( $F_{ST}$  ranged 0.01 to 0.02). Rather than topological isolation, the authors associated this barrier with predator-prey specialization to different caribou herds with seasonal migratory patterns that were exclusive to either side of the river. Genetic distances observed between Arctic Island (*C. l. arctos* in Nowak 1995) and mainland (*C. l. occidentalis* in Nowak 1995) wolves ( $F_{ST}$  ranged 0.09 to 0.19) were twice that observed among mainland wolves ( $F_{ST}$  ranged 0.01 to 0.1) (Carmichael et al. 2001, Table 2).

In a study designed to explore pack composition among wolves in Algonquin Provincial Park, Grewal et al. (2004) assessed parent-offspring relationships from mtDNA control region sequence as well as Y-chromosome and autosomal microsatellite variation. Wolves at proximal locations in Frontenac Axis ( $n = 74$ ) and Magnetawan region ( $n = 26$ ), as well as Northeast Ontario ( $n = 33$ ), Abitibi-Temiscamingue region ( $n = 13$ ), and La Verendrye Reserve in southeast Quebec ( $n = 13$ ) were also assessed. The data from the mtDNA sequence and Y-chromosome microsatellite portions of this study will be discussed in following sections.

In addition to breeding adults and offspring, most packs were found to have at least one additional, non-breeding and unrelated adult. STRUCTURE analysis identified five immigrants into the Algonquin Provincial Park: three from Frontenac Axis, one from Magnetawan region, and one from north of the park. Wolves from the Frontenac Axis and Magnetawan region locations exhibited significant introgression from coyote, whereas the genotypes of wolves from north of the park (within the range of *C. l. nubilus* in Nowak 1995) predominantly exhibited *C.*

*lupus* alleles. Algonquin Provincial Park wolves (within the range of *C. l. lycaon* in Nowak 1995 and *C. lycaon* in Wilson et al. 2000) were differentiated from proximal populations ( $F_{ST}$  ranged 0.024 to 0.055), but were approximately twice as divergent from wolves from the more distant Abitibi-Temiscamingue region ( $F_{ST} = 0.089$ ), La Verendrye Reserve ( $F_{ST} = 0.091$ ) and Northeastern Ontario ( $F_{ST} = 0.076$ ) localities.

Weckworth et al. (2005) compared microsatellite variation among wolf populations from coastal southeast Alaska ( $n = 101$ ; *C. l. nubilus* in Nowak 1995) and coastal south-central Alaska, interior Alaska, Northwest Territories and British Columbia ( $n = 120$ ; *C. l. occidentalis* in Nowak 1995) and found that the coastal and continental groups were distinguished by significant differences in allele frequencies. Mean distance between coastal and continental wolves (average  $F_{ST} = 0.16$ ) was nearly twice that of the mean distance within groups (average  $F_{ST} = 0.09$ ). However, wolves from coastal south-central Alaska and coastal southeast Alaska were similarly distinguished (average  $F_{ST} = 0.16$ ). The authors attributed the observed genetic divergence to the waterways, high mountains, and glaciers barring the dispersal of wolves (Weckworth et al. 2005, Figure 2). They also hypothesized that southeast Alaska was colonized from the south with the retreat of the last Pleistocene glaciation whereas interior Alaska had been colonized by wolves from the Beringian refugium to the north (Nowak 1983).

Musiani et al. (2007) compared microsatellite DNA variation in gray wolves from seven localities in tundra/taiga habitat in Northwest Territories, Canada ( $n = 337$ ) and four localities in boreal forest habitat in Northwest Territories and northern Alberta ( $n = 67$ ) to examine the effects of habitat specializations on population structure. GENELAND analysis combined the wolves from tundra/taiga and boreal forest localities into respective populations, and found significant genetic differentiation ( $F_{ST} = 0.03$ ) between the two. As genetic and geographic

distances were not significantly correlated, and topological barriers were not in evidence, the authors concluded that prey and habitat specialization had promoted the genetic differentiation. The autosomal and Y-chromosome microsatellite data obtained in this study are discussed at length elsewhere.

Carmichael et al. (2007, 2008) explored geographic variation in wolves inhabiting the North American Arctic. Carmichael et al. (2007) focused on the structuring of mainland populations, Carmichael et al. (2008) on Canadian Arctic Islands wolves. The wolves genotyped in these studies included individuals from the Canadian Arctic Islands ( $n = 342$ ), southeast Alaska coastal islands ( $n = 35$ ) and mainland locations ( $n = 1648$ ) that were attributable to three different subspecies following Nowak's (1995) reclassification and general subspecies boundaries: *C. l. occidentalis*, *C. l. nubilus*, and *C. l. arctos*. Ten populations (Carmichael et al. 2007, Figure 2(b)) were identified through the combined results of STRUCTURE and GENELAND analyses. Carmichael et al. (2007) explained this population divergence in terms of reduced dispersal due to topographic, habitat selection, and prey preference barriers. They noted that the geospatial distribution of the recognized populations did not correspond to the morphological subspecies boundaries in Nowak (1983, 1995), but reflected contemporary factors affecting gene flow.

Sampling areas attributable to *C. l. occidentalis* were generally grouped together in a neighbor-joining tree of genetic distance (Carmichael et al. 2008, Figure 3B). However, the sample group from southeast Alaska coastal islands (*C. l. nubilus*, after Nowak 1995) was more similar to proximal interior populations in Alaska, Yukon, Northwest Territories and British Columbia (*C. l. occidentalis*, after Nowak 1995) than to the cluster of other *C. l. nubilus* sampling areas from Nunavut (Qamianirjuaq and Bathurst) and Atlantic Canada. The Manitoba

and British Columbia samples that straddled the eastern and western boundaries between *C. l. occidentalis* and *C. l. nubilus* were both included within the *C. l. occidentalis* portion of the tree. Of the *C. l. nubilus* sampling areas, Baffin Island and an adjacent mainland area occurred together on a well-supported branch, but the wolves of the Atlantic sampling area (including Labrador, New Foundland, New Brunswick, and Nova Scotia) were as divergent from the Baffin NE Mainland population as they were from the Victoria/Banks/High Arctic Islands wolves (Carmichael et al 2008, Table 3).

Consistent with recent re-population from the south rather than having occupied a separate glacial refuge north of the ice sheet as proposed by Nowak (1983), Canadian Arctic Island wolves exhibited few unique alleles. The lack of isolation of island and mainland wolf populations was further supported by the observation of high gene flow between mainland and island wolf populations (Carmichael et al 2008, Table 2) and annual over-ice, island-mainland migrations of island wolves (Carmichael et al. 2001) and their caribou prey (Carmichael et al. 2008, Figure 4). Although genetic distance between island and mainland populations was generally much higher ( $D_s$  ranged 0.08 to 0.63) than among mainland populations ( $D_s$  ranged 0.01 to 0.30) (Carmichael et al. 2008, Table 3), the effects of small founder size and genetic drift may have contributed to the genetic distinctiveness of Arctic Islands wolves (Paetkau et al 1997). For example, the High Arctic and Victoria Island populations as well as the southeast Alaska coastal islands populations exhibited evidence of inbreeding ( $F_{IS}$  ranged 0.18 to 0.63). As a result, the authors recommended that conclusions about, “the taxonomic validity of *C. l. arctos* should be deferred,” until data are available from mtDNA and Y-chromosome sequences (Carmichael et al. 2008, p. 886).

Wheeldon and White (2009) successfully genotyped three historical wolf specimens from Minnesota and Wisconsin (1899-1908) at six microsatellite loci and characterized them in a STRUCTURE analysis along with wolf and coyote samples from throughout Canada. The microsatellite profiles of both modern and historical western Great Lakes wolves exhibited similar admixture proportions that were attributed to *C. lupus*-*C. lycaon* hybridization. These results suggested that *C. lupus* and *C. lycaon* were sympatric and interbreeding prior to their extirpation from the region in the early twentieth century.

Kobl Müller et al. (2009a) used autosomal microsatellite, Y-chromosome microsatellite, and mtDNA sequence analysis to compare modern Great Lakes wolves with eastern ( $n = 49$ ) and western ( $n = 78$ ) coyotes, western *C. lupus* ( $n = 58$ ), and historical Great Lakes wolves (collected prior to the modern establishment of coyotes in the region). The modern Great Lakes wolf sample consisted of some ( $n = 64$ ) of the same Minnesota animals analyzed by Lehman et al. (1991) and Roy et al. (1994), recovered Wisconsin ( $n = 16$ ) and Upper-Peninsula Michigan wolves ( $n = 63$ ), as well as wolves from Ontario ( $n = 51$ ) and Quebec ( $n = 9$ ). The historical sample ( $n = 19$ ) included wolves from Minnesota, Wisconsin, Michigan, Ontario, Quebec, and New York. The Y-chromosome and mtDNA data are discussed separately.

STRUCTURE analysis of the Northwest Territories and Great Lakes wolf samples from Minnesota, Wisconsin, and Michigan revealed little evidence of admixture, whereas, the authors interpreted the significant admixture observed in wolves from Ontario and Quebec as the result of recurrent hybridization between gray wolves and Great Lakes wolves and between Great Lakes wolves and coyotes. Genetic divergence was reported in this paper using the notation  $\Theta_{ST}$ , which is simply  $\Theta$ , the coancestry parameter, in the original notation of Weir and Cockerham (1984). Little difference was detected between the modern recovered population of Great Lakes

wolves and the historical sample ( $\Theta = 0.036$ ). Great Lakes wolves were equally divergent from eastern ( $\Theta = 0.142$ ) and western ( $\Theta = 0.133$ ) coyote, but less so from western *C. lupus* ( $\Theta = 0.078$ ). The authors did not conclude that Great Lakes wolves constituted a separate species, but rather a unique population or ecotype of *C. lupus*.

Wolf-dog hybridization in the wolf population of Vancouver Island was the subject of study by Muñoz-Fuentes et al. (2009b) using microsatellites and mtDNA control region sequence. Wolves were thought to have been extirpated from the island between 1950 and 1970, after which they re-colonized from the adjacent mainland of British Columbia. In contrast to the mtDNA data, the microsatellite data did not reveal evidence of dog introgression. The authors concluded that these results were consistent with a single hybridization event occurring early in the re-establishment of wolves on the island.

Wilson et al. (2009) explained the genetic relationships of three “races” of wolves living in Ontario, Canada: the larger Ontario and smaller Algonquin types of the eastern timber wolf (*C. l. lycaon*, Kolenosky and Standfield 1975) and the Tweed wolf that is thought to be a wolf-coyote hybrid. The authors assessed the relationships of wolves from across Ontario for evidence of hybrid admixture from autosomal microsatellite variation. The study included wolves from both southern regions of Ontario: Frontenac Axis (n = 74), Magnetawan region (n = 26) and Algonquin Provincial Park (n = 92); and northern regions: northeast Ontario (n = 33), northwest Ontario (n = 30) and Pukaskwa National Park (n = 13).

STRUCTURE analysis segregated the sample into three populations (Wilson et al. 2009, Figure 3): wolves from the northern regions, which also exhibited *C. lupus* mtDNA (Old World type, Wilson et al. 2009, Table 2) at high frequency (24-85%), eastern wolves from Algonquin Provincial Park (3% *C. lupus* mtDNA), and Tweed wolves from the Magnetawan region and

Frontenac Axis in the south (100% *C. latrans*/*C. lycaon* mtDNA, New World type, Wilson et al. 2009, Table 2). The microsatellite genotypes of Frontenac Axis and Magnetawan region wolves were significantly introgressed with coyote alleles (> 50% of population), whereas those from north of the park (within the range of *C. l. nubilus* in Nowak 1995) exhibited genotypes of predominantly gray wolf alleles (> 70% of population) (Wilson et al. 2009, Figure 4). Algonquin Provincial Park wolves (within the range of *C. l. lycaon* in Nowak 1995 and *C. lycaon* in Wilson et al. 2000) were differentiated from southern Magnetawan region and Frontenac Axis populations ( $F_{ST}$  ranged 0.022 to 0.055, respectively), but were twice as divergent from northern region wolves ( $F_{ST}$  ranged 0.071 to 0.117) (Wilson et al. 2009, Table 5). The authors concluded that eastern wolves of the smaller Algonquin type (Kolenosky and Standfield 1975) are *C. lycaon*, eastern wolves of the larger Ontario type (Kolenosky and Standfield 1975) are *C. lycaon*-*C. lupus* hybrids, and Tweed wolves are *C. lycaon*-coyote hybrids.

Fain et al. (2010) tested the influence of hybridization on wolf recovery in the western Great Lakes states. Microsatellite DNA variation was compared in the recovered Great Lakes states wolf population from Minnesota, Wisconsin and Upper Peninsula Michigan (n = 112); western gray wolves from Alaska, British Columbia and Alberta (n = 103); Wisconsin coyotes (n = 36) and domestic dogs (n = 39).

STRUCTURE analysis clearly segregated Great Lakes states wolves, western gray wolves, coyotes and dogs (Fain et al. 2010, Figure 6); however, the Great Lakes sample included *C. lupus* x *C. lycaon* hybrids (25%). Divergence between Great Lakes wolves (within the range of *C. l. nubilus* in Nowak 1995) and western *C. l. occidentalis* (Nowak 1995) was high ( $F_{ST}$  = 0.125), and Great Lakes wolves were equally divergent from domestic dogs ( $F_{ST}$  = 0.123).



Moreover, the analysis showed that western Great Lakes wolves and sympatric coyotes were also highly differentiated ( $F_{ST} = 0.159$ ). The authors concluded that this result was inconsistent with recent hybridization. The mtDNA and Y-chromosome data also obtained in this study are discussed separately.

Rutledge et al. (2010b) compared microsatellite DNA variation in gray wolves from northeast Ontario ( $n = 51$ ), eastern wolves from Algonquin Provincial Park ( $n = 128$ ) and coyotes from Frontenac Axis ( $n = 38$ ) in eastern Ontario in order to examine the effects of hybridization on population structure. GENELAND analysis segregated the three localities (separated by 700km) into genetically differentiated populations ( $F_{ST}$  ranged 0.052 to 0.120), but there was evidence of admixture. Algonquin Provincial Park wolves were admixed with both northeast Ontario ( $n = 8$ , 6%) and Frontenac Axis ( $n = 14$ , 11%) wolves, and over 15% of northeast Ontario and Frontenac Axis wolves were admixed with Algonquin Provincial Park wolves. Principle components analysis of individual autosomal microsatellite genotypes placed Algonquin Provincial Park wolves closest to Frontenac Axis wolves and revealed a south-north cline in allele frequencies. These results were considered supportive of the hypothesis that eastern wolves in Algonquin Provincial Park represent a conduit of gene flow between gray wolves to the north and coyotes to the south. In addition, mtDNA and Y-chromosome haplotype frequency distributions were consistent with the hypothesis that introgression was gender-biased with females of the smaller species mating males of the larger species. Conspecific pairings at Algonquin Provincial Park were more common than predicted by random mating. The mtDNA and Y-chromosome data obtained in this study are discussed further in following sections.

Wheeldon et al. (2010) evaluated the species distinction of the wolves from the western Great Lakes region. The authors analyzed species-specific mtDNA and Y-chromosome

sequence haplotypes in addition to autosomal microsatellite variation. The Y-chromosome and mtDNA sequence haplotype data are discussed elsewhere. The sample included 410 wolves and coyotes from the western Great Lakes states (Minnesota, Michigan, and Wisconsin) and western Ontario. Wolves and coyotes were distinguished as separate populations in both STRUCTURE and factorial correspondence analysis with little indication of hybridization. Y-chromosome and mtDNA haplotypes of both eastern and gray wolves were found in the wolf sample. The lack of subdivision of the wolf sample in the STRUCTURE analysis was cited as evidence that the wolves of the western Great Lakes states and western Ontario are members of a single interbreeding population.

*Summary of studies of autosomal genetic markers.* Studies of autosomal microsatellite DNA generally distinguish groups representing western gray wolves, eastern wolves (alternatively referred to as Great Lakes wolves in some studies), red wolves, Mexican wolves, and coyotes.

Most studies in the western Great Lakes region, found the wolves to comprise an admixed population. An exception is the study of Koblmüller et al. (2009a), which claimed little admixture in the western Great Lakes region. There is disagreement on the source of the admixture: those who claim it represents gray wolf-coyote interbreeding (Roy et al. 1994, 1996; García-Moreno et al. 1996), others gray wolf-eastern wolf interbreeding (Wilson et al. 2000, Wheeldon and White 2009, Wilson et al. 2009, Fain et al. 2010, Wheeldon et al. 2010).

Autosomal microsatellite DNA data were interpreted by Carmichael et al. (2007, 2008) as failing to support Nowak's interpretation, based on his morphometric analysis, of long isolation and subspecific validity of the arctic wolf, *Canis lupus arctos*.

*Single nucleotide polymorphisms (SNPs).* vonHoldt et al. (2011) report the results of an analysis of 48,036 single nucleotide polymorphism (SNP) markers in worldwide samples of dogs and wild canids. Samples of wild canids from North American include 57 coyotes, 12 red wolves, 19 Great Lakes wolves, and 70 gray wolves. In a principal components analysis (vonHoldt et al. 2011, Figure 3), the first component (accounting for 10% of the total variance) separated domestic dogs from the wild canids, and the second principal component (accounting for 1.7% of the variance) separated coyote and red wolf from other North American wolves. A successive series of STRUCTURE analyses first distinguished dogs from wild North American canids ( $K = 2$ ), coyote and red wolf from other wolves ( $K = 3$ ), Mexican wolf from other gray wolves ( $K = 6$ ), red wolf from coyotes ( $K = 9$ ), and Great Lakes wolves from gray wolves ( $K = 10$ ). A SABER analysis was performed on Great Lakes wolves (including two individuals from Algonquin Provincial Park) and red wolves using western gray wolves, dogs, and western coyotes as ancestral reference populations.

Conclusions by vonHoldt et al. (2011) include: red wolf is an admixed variety derived from coyote; Great Lakes wolf is an admixed variety derived from gray wolves; Great Lakes wolf is genetically distinct from other gray wolves; Mexican wolf is a distinct form of gray wolf. Within gray wolves, geographic variation in SNP composition is attributed to geographic variation in ecological conditions rather than to taxonomic distinctions.

*Mitochondrial DNA.* Lehman et al. (1991) used restriction analysis of the entire mtDNA genome to describe haplotype variation in wolves and coyotes in areas of sympatry as well as isolated regions of allopatry in order to assess the occurrence of inter-species hybridization. The sample included wolves from Minnesota, Isle Royale in Lake Superior, western Ontario, Algonquin

Provincial Park, Quebec, as well as selected western localities extending to Nome, Alaska. A maximum parsimony tree (Lehman et al. 1991, Figure 3) separated western *Canis lupus* from coyotes and wolves from the Great Lakes region. Although some Great Lakes wolves exhibited *C. lupus* haplotypes, most shared haplotypes with coyotes, or had “coyote-type” haplotypes (i.e., phylogenetically similar but not observed in coyotes). All wolves sampled from southern Quebec and southeastern Ontario exhibited either coyote or coyote-type haplotypes. Lehman et al. (1991) concluded that Great Lakes wolves have a history of repeated hybridization with coyotes with the consequent introgression of coyote mtDNA haplotypes. They suggested that “Boreal type” (in the sense of Kolenosky and Standfield [1975]) and *C. l. lycaon* from north of the portion of their range also occupied by coyotes may represent “pure wolf lines” while the “Algonquin type” wolves to the south have been subject to coyote introgression. Because the most common coyote-type haplotypes found in Minnesota wolves were not found in coyotes, the authors suggested that hybridization involving those wolves occurred “in the distant past.” They also stated that, as both *C. lupus* and coyote-type haplotypes occurred in individuals within the same packs, the Great Lakes region may contain a complex mix of *C. lupus*, coyotes, and their hybrids.

Wayne and Jenks (1991) evaluated the genetic integrity of the source population from which the founders of the captive red wolf breeding program were selected. Seventy-seven canids were captured in southeastern Texas and southwest Louisiana in the 1974-1976 effort to rescue the last remaining wild *C. rufus*, and included the four matrilineal lines used to found the red wolf captive breeding program (USFWS 1990). These animals were characterized morphologically as coyotes (58%), *C. rufus*-coyote hybrids (31%), and *C. rufus* (11%). Genetic characterization of these same animals with whole mtDNA genome restriction analysis found

that they exhibited either coyote (84%) or gray wolf (16%) mtDNA and that the morphological and genetic classifications often did not correspond. Moreover, seven individuals (9% of the sample) exhibited a mtDNA restriction type previously observed only in Mexican wolves (*C. l. baileyi*), and parsimony analysis placed the mtDNA restriction type observed in captive red wolves within the phylogenetic clade composed exclusively of coyote types. The captive red wolf type was indistinguishable from a haplotype (i.e., C32) found in coyotes from Louisiana. In another part of this study, a portion of the mtDNA cytochrome *b* gene sequence was determined from historical (1905-1930) museum skins identified as *C. rufus* from Texas (n = 1), Louisiana (n = 1), Arkansas (n = 2), Oklahoma (n = 1), and Missouri (n = 1). Sequences were compared to similarly characterized gray wolf (n = 2), Mexican wolf (n = 1), coyote (n = 6), dog (n = 1), and golden jackal (n = 1) reference samples. The *C. l. baileyi* haplotype differed by only a single substitution from sequences of three historical red wolf specimens from Louisiana, Oklahoma, and Texas, which had a haplotype identical to a *C. lupus* individual from Minnesota. The *C. l. baileyi* haplotype was 10 to 19 nucleotide substitutions different from those of *C. rufus* from Arkansas, Missouri, and the captive population.

Wayne et al. (1992) used whole mtDNA genome restriction analysis to study variation in *Canis lupus*, mostly from North America (n = 204), but included Eurasian samples for comparison (n = 35). North American wolves exhibited five haplotypes, Eurasian wolves seven, and none were shared. Subspecies assignments were not identified, but the distributions of wolf haplotypes W1 through W4 among North American wolves may have a bearing on the standing and relationships of the subspecies *C. l. nubilus* and *C. l. occidentalis*. The W3 haplotype was the most common and widespread, from Alaska to Montana, but not Manitoba, which all (n = 4) had the unique W2 haplotype. Haplotype W1 was found from the Northwest Territories to

Montana (and later also found in Minnesota and central Ontario [Wayne et al. 1995]), and W4 was found in scattered locations from Alaska to Montana. These haplotypes all differed by one or two restrictions sites (about 0.1 to 0.2% sequence divergence). The fifth haplotype (W14) was found only in the *C. l. baileyi* samples from the Mexican wolf captive breeding program. This haplotype was most closely associated with Eurasian *C. lupus* in the phylogenetic analysis (Wayne et al. 1992, Figure 2). Altogether, eastern wolves (n = 106) from Manitoba, Minnesota, Michigan, Ontario and Quebec exhibited seven “coyote-derived” haplotypes.

Roy et al. (1996) tested the hypothesis that red wolves are evolutionarily ancestral to both gray wolves and coyotes but had undergone a recent interval of interbreeding with coyotes at the time of their near extermination in the wild. A portion of the mtDNA cytochrome *b* gene was sequenced from historical (1919-1943) museum skins of *C. rufus* (n = 11) and compared to those described by Wayne & Jenks (1991) from additional historical *C. rufus* (n = 6), captive *C. rufus* (n = 1), *C. lupus* from Alaska (n = 1), *C. lupus* from Minnesota (n = 1), *C. l. baileyi* (n = 1), *C. latrans* (n = 5), and *C. aureus* (n = 1). The observed haplotypes comprised two major clades, one including all coyotes, the other gray wolves. Most *C. rufus* were placed in the “coyote clade” (n = 8), but others from Missouri and Oklahoma (n = 3) were included in the wolf clade. Only haplotype CruOK3 was common to the different sets of historical red wolf samples.

Sequence divergences were consistent within species: *C. rufus* (0.4 to 0.9%), *C. latrans* (0.4 to 1.7%) and *C. lupus* (0.4 to 1.3%). Sequence divergence between *C. lupus* and *C. latrans* was about four-fold greater (3.2-5.6%) and *C. lupus* were over 3.2% divergent from *C. rufus* from Arkansas, Oklahoma, and Texas, but less than 2.1% divergent from *C. rufus* from Missouri and Oklahoma. The authors concluded that introgression of *C. lupus* into *C. rufus* had occurred in Oklahoma and Missouri but not in Arkansas and Texas. The divergence observed between the

1231 16 historical *C. rufus* sequences and gray wolf and coyote references was consistent with the  
1232 amount of within species variation found in gray wolves and coyotes. The results of the  
1233 microsatellite analysis performed in this study are discussed in the preceding section.

1234 Vilà et al. (1999) included data from North American *C. lupus* (n = 24) in a broad-scale  
1235 phylogeographic study of mtDNA control region sequence variation in wolves (Vilà et al. 1999,  
1236 Figure 1, which is reproduced here as Figure 8). The phylogenetic tree that resulted from a  
1237 neighbor-joining analysis placed *Canis lupus baileyi* basal to all other wolf clades, while the five  
1238 other North American haplotypes sorted into three different clades that each included Eurasian  
1239 *C. lupus*. There was no obvious geographic pattern in the distribution of these haplotypes, but  
1240 the scale of sampling was too coarse for this purpose.

1241 The single haplotype shared by the six individuals of *C. l. baileyi* was unique and was  
1242 more similar to certain Eurasian wolves than to other North American *C. lupus*. Vilà et al.  
1243 (1999, p. 2099) suggested that *C. l. baileyi* may represent an early invasion of North America by  
1244 Eurasian wolves, before the arrival of *C. lupus* with other haplotypes. The phylogenetic analysis  
1245 also included the mtDNA control region sequence of a single Texas *C. rufus*. The *C. rufus*  
1246 sequence was not found in coyotes, although it was within the coyote clade of the neighbor-  
1247 joining tree.

1248 Wilson et al. (2000) examined the origin and taxonomy of the wolves of eastern Canada,  
1249 and whether they are the result of hybridization between gray wolves and coyotes as has been  
1250 suggested for the red wolf (Roy et al. 1996). Wilson et al. (2000, Figures 5A and 5B, reproduced  
1251 here as Figure 9 and Figure 10, respectively) compared the mtDNA control region sequences of  
1252 historical wolves (1960-1965) from the vicinity of Algonquin Provincial Park (n = 13) and

southern Ontario (n = 8), Texas coyotes (n = 27), captive *C. rufus* (n = 12), and *C. lupus* from Manitoba, northern Ontario, northern Quebec and the Northwest Territories (n = 9).

The authors found no gray wolf sequences in historical Algonquin wolves or in captive *C. rufus*. Moreover, captive red wolves and historical Algonquin wolves exhibited unique haplotypes not found in either *C. lupus* or coyotes. Phylogenetic analyses revealed a similarity between the unique haplotypes C1 of Algonquin wolves, C3 found in a single modern Manitoba wolf and C2 of captive red wolves (Wilson et al. 2000, Figure 5). This group was strongly separated from the wolves of western and northern Ontario as well as the Northwest Territories. Other eastern Canadian wolves had haplotypes grouped by the analyses with coyote haplotypes, although most were not found in western coyotes (Lehman et al. 1991, Wilson et al. 2000).

Wilson et al. (2000) attributed the existence of the unique “coyote-like” haplotypes in Algonquin wolves and *C. rufus*, as well as the ability of these canids to hybridize with coyotes, to their having shared a more recent common ancestor with coyotes in the New World rather than with the more distantly related, Old World-evolved *C. lupus*. Based on this relationship, they concluded that eastern Canadian wolves should not be considered a subspecies (*C. l. lycaon*) of gray wolf, but as a full species, *C. lycaon*, closely related to, if not conspecific with *C. rufus*. If conspecific with *C. rufus*, Wilson et al. (2000) suggested that the name that might best apply would be *Canis lycaon*, based on “historical taxonomic classifications.” Wilson et al. (2000) maintained that *C. lycaon* ranged into Minnesota, northwestern Ontario, and Manitoba, and suggested that *C. lycaon* and *C. lupus* may be interbreeding in these western reaches.

Eight *C. lupus* individuals from five different North American localities were also characterized in this study. Although three haplotypes (i.e., C22, C23 and C24) were found, the scale of sampling was too coarse to detect any obvious geographic pattern in their distribution.



For example, Ontario and Manitoba wolves should represent *C. l. nubilus* but haplotype C23 was found there as well as in the Northwest Territories where wolves represent *C. l. occidentalis* (Nowak 1995, Table 4 in this paper). As discussed in the preceding section, Wilson et al. (2000) also characterized microsatellite variation in wolves from across Ontario, Quebec and western Canada.

To test the hypothesis that eastern wolves evolved in North America, Wilson et al. (2003) obtained mtDNA control region sequences from the hides of two historical wolves, one from New York state (collected in the 1890s) and the other from Penobscot County, Maine (in the 1880s). Both animals were presumed unaffected by coyote hybridization as they had been collected prior to the invasion of the eastern U.S. by western coyotes. The authors suggested that the exhibition of “coyote-like” haplotypes by these animals would indicate a most recent common ancestor between eastern wolves and coyotes rather than gray wolves.

In both neighbor-joining and maximum parsimony analyses (Wilson et al. 2003, Figure 1) neither specimen clustered with *C. lupus* samples from northern Ontario, northern Quebec, Manitoba, the Northwest Territories, or Europe. The haplotype of the Maine individual was identical to the unique *C. lycaon* haplotype C1 previously identified from Algonquin Provincial Park wolves. The New York sample clustered with modern western coyotes, although this haplotype (C13) was not identical to any reported from coyotes and was referred to by the authors as “coyote-like.” The authors interpreted these findings as supporting both the presence of *C. lycaon* in Maine and New York in the 1880s and their earlier contention (Wilson et al. 2000) that *C. lycaon* and *C. rufus* represent an eastern wolf with an evolutionary history separate from that of *C. lupus*.

Adams et al. (2003) compared the mtDNA control region sequences of the four matriline of the captive red wolf breeding program (n = 8), red wolf-coyote hybrids (n = 40) collected from the Alligator River National Wildlife Refuge in northeastern North Carolina (Adams et al. 2003, Figure 1), and coyotes collected from locations in California, Kentucky, Nebraska, North Carolina, Texas and Virginia (n = 66).

The authors found that the four matriline of the 14 red wolf founders of the captive breeding program had identical control region sequences, haplotype Cru (same as C2, Wilson et al. 2000), and when compared to haplotypes observed in coyotes proximal to the red wolf experimental population (i.e., 14 haplotypes found in North Carolina and Virginia coyotes, n = 50) and the published literature (i.e., 12 coyote haplotypes), Cru was unique. The average sequence divergence between haplotype Cru and these 26 coyote haplotypes was 3.24% (range = 1.2-10%). The average sequence divergence among these coyote haplotypes was 2.79% (range = 0.3-11%).

In a study designed primarily to explore the pack composition of wolves in Algonquin Provincial Park, Grewal et al. (2004) reported on the variation of mtDNA control region sequence haplotypes (n = 261), Y-chromosome microsatellite haplotypes (n = 248) and autosomal microsatellite genotypes (n = 261) among wolves at various locations in Ontario and Quebec. The data from the autosomal and Y-chromosome microsatellite portions of this study are discussed in the preceding and following sections.

Mitochondrial DNA haplotypes were identified as being of *C. lupus* (C22), *C. lycaon* (C1, C9), or coyote (C13, C14, C16, C17, C19; but see the summary of the study of mtDNA variation of Wheeldon and White (2009) for an alternative interpretation of C13). The species source of the rare haplotypes C3 and C36 were not identified. *Canis lupus* mtDNA haplotypes

were uncommon (four of 102 individuals) in wolves at Algonquin Provincial Park, but were 10 times more common to the north (La Verendrye Reserve) and west (northeastern Ontario) of Algonquin Park (Table 4 of this paper). *Canis lycaon* mtDNA haplotypes were observed at higher frequency and coyote haplotypes at lower frequency to the south (Frontenac Axis) of Algonquin Provincial Park (Grewal et al. 2004, Figure 2, Table 1). Grewal et al. (2004) suggest that coyote haplotypes probably introgressed into Algonquin Park wolves during the 1960s when the population was reduced and coyote/wolf hybrids (“Tweed wolves” of Kolenosky and Standfield 1975) expanded into the park.

Leonard et al. (2005) obtained mtDNA control region sequences from 34 museum specimens of *C. lupus* collected from the conterminous U.S. and Labrador prior to the extirpation of wolves (i.e., 1917 or earlier) to determine the proportion of the variation that has been retained by extant populations of wolves in Alaska, Canada, and northeast Minnesota. The sample of historical wolves included individuals identified morphologically and by geographic provenance as *C. l. baileyi* (n = 8), *C. l. labradorius* (n = 4), *C. l. nubilus* (n = 16), and *C. l. youngi* (n = 6). The 13 mtDNA haplotypes observed in these historic wolves were sorted into northern and southern clades by phylogenetic analysis. The northern clade, representing *C. l. nubilus/youngi/labradorius* (combined under *C. l. nubilus*, Nowak 1995), included nine haplotypes, three of which are common in modern wolves (i.e., lu28, lu32 and lu38). The southern clade, representing *C. l. baileyi*, included four haplotypes, only one of which remains in modern Mexican wolves (i.e., lu33). An historical *C. l. baileyi* sample from Chihuahua, Mexico, (USNM98313/JAL474) exhibited an mtDNA haplotype (lu60) that is otherwise known only from coyotes. Overall, 58% of the historical wolf samples exhibited the same four haplotypes observed in the modern wolf sample. Eight haplotypes found in historical wolves were not

observed in modern wolves. Individuals with southern clade haplotypes were found as far north as Utah, Colorado, and Nebraska, whereas, northern haplotypes were found as far south as Arizona. The authors explained this observation as genetic evidence of population interaction: “In highly mobile species, large zones of intergradation may characterize subspecies boundaries . . .”

Musiani et al. (2007) used variation in mtDNA control region sequence, and Y-chromosome and autosomal microsatellites to examine the effects of habitat specialization on population structure in wolves from northern Canada. The autosomal and Y-chromosome microsatellite results are discussed in respective sections.

Mitochondrial DNA control region haplotype frequency and diversity were analyzed in gray wolves from tundra/taiga ( $n = 337$ ) habitat in Northwest Territories, and boreal forest habitat ( $n = 67$ ) in Northwest Territories and northern Alberta. Nine different haplotypes were characterized, two predominated in tundra/taiga wolves (89% of sample). While eight haplotypes were found in boreal forest wolves, the two most common comprised only 42% of the sample (Musiani et al. 2007, Figure 5). Haplotype frequencies differed between habitat types. For example, haplotype lu32 was observed in both samples but was far more common among tundra/taiga wolves (71% of sample) than in boreal forest wolves (22% of sample). Similarly, mitochondrial gene diversity was three times higher in wolves from boreal forest habitat than in tundra/taiga wolves (0.051 and 0.153 respectively). Genetic differentiation between wolves of different sexes from boreal forest and tundra/taiga habitats was three times higher for females ( $F_{ST} = 0.353$ ) than males ( $F_{ST} = 0.138$ ). Consistent with the autosomal microsatellite analysis, the mtDNA control region sequence data also supported the characterization of boreal forest and tundra/taiga wolves as separate populations (but see results obtained from Y-chromosome

microsatellite variation). The authors concluded that sex bias, likely due to natal habitat-biased dispersal, had contributed to the population differentiation reflected by mtDNA variation ( $F_{ST} = 0.28$ ).

Leonard and Wayne (2008) investigated the controversy surrounding wolf-coyote hybridization and its impact on the recovery success of western Great Lakes wolves. They compared mtDNA control region sequences from historical wolves ( $n = 12$ , collected 1905-1916) and 18 modern wolves ( $n = 18$ ) from portions of Michigan, Wisconsin, and Minnesota. All twelve historical specimens exhibited what the authors termed “Great Lakes” haplotypes (i.e., GL1, GL2, GL3, GL5, and GL8) that were basal to modern coyote haplotypes in a parsimony consensus cladogram (Leonard and Wayne 2008, Figure 1). No historical sample exhibited a haplotype from the clade that included western gray wolf haplotypes. By comparison, the composition of the haplotypes in the modern samples was: Great Lakes: 50%; *C. lupus*: 6%; coyote-like: 44%. In the modern samples, coyote-like haplotypes occurred in Minnesota and Michigan, but the Michigan sample was small ( $n = 2$ ) and no modern Wisconsin wolves were sampled. Both *C. lupus* and Great Lakes haplotypes were found in modern Minnesota wolves. The authors concluded that historical Great Lakes wolves were characterized by phylogenetically distinct haplotypes representing an endemic “American wolf” and that these had been replaced by a modern population of mixed *C. lupus* and coyote ancestry.

Hailer and Leonard (2008) explored the relationships of three canid species, historically sympatric in central Texas, for evidence of hybridization with an analysis of mtDNA control region sequence and Y-chromosome microsatellite haplotypes. The Y-chromosome data are discussed in a following section. The sample included wolves from the captive breeding programs for *C. l. baileyi* ( $n = 16$ ) and *C. rufus* ( $n = 5$ ) as well as wild *C. latrans* from Texas ( $n =$

53) and Nebraska (n = 75). These were compared to each other and with similarly characterized reference animals from previously-published studies (Vilà et al. 1999, Adams et al 2003, Leonard et al. 2005).

All *C. rufus* had coyote-like control region haplotype ru1 (Vila et al. 1999, same as Cru in Adams et al. 2003) that was distinct from *C. l. baileyi*. Although possibly of coyote origin, haplotype ru1 was not observed in any of the coyotes (n = 131) in the study. The lu60 haplotype found in an historical *C. l. baileyi* skull (Leonard et al. 2005) differed by two base changes from the Texas coyote haplotype la86 and was interpreted as possible evidence of past coyote introgression into *C. l. baileyi*. A single Texas coyote exhibited the mtDNA haplotype Cla12 which was phylogenetically most similar to the gray wolf haplotype lu32 (Figure 2, Hailer and Leonard 2008). Haplotype lu32 has also been observed in historical Mexican wolves (Leonard et al 2005) and the authors suggested that introgression between Mexican wolves and coyotes may have been the route by which this marker entered the Texas coyote population.

Wheeldon and White (2009) addressed the attestation (Leonard and Wayne 2008) that phylogenetically distinct historical Great Lakes wolves have been replaced by a modern population of mixed *C. lupus* and coyote ancestry. In addition to the microsatellite DNA data summarized previously, Wheeldon and White (2009) compared the mtDNA control region sequences of three historical specimens (1899-1908) from Minnesota and Wisconsin. The Wisconsin specimen displayed the *C. lycaon* C1 haplotype identified by Wilson et al. (2000) in Algonquin wolves, whereas the two Minnesota specimens exhibited the coyote-like C13 haplotype also found in a historical (circa 1890s) wolf hide from New York (Wilson et al. 2003). Wilson et al. (2003) and Wheeldon and White (2009) considered C13 to be a *C. lycaon* haplotype related to coyote haplotypes through introgression either before European settlement

or much earlier in the Pleistocene. Both haplotypes were identical to those observed in other historical wolves from the Great Lakes region (Koblmüller et al. 2009a, Figure 2a). Wheeldon and White (2009) interpreted these data as supporting an historical western Great Lakes wolf population with genetic composition similar to the wolves currently populating the region. The authors' analyses of autosomal microsatellite DNA data are discussed in the preceding section.

Wheeldon et al. (2010) analyzed species-specific mtDNA sequence haplotypes and Y-chromosome and autosomal microsatellite variation in 410 wolves and coyotes from the western Great Lakes states (Minnesota, Michigan, and Wisconsin) and western Ontario. The Y-chromosome and autosomal microsatellite data are discussed separately.

Both eastern wolf and gray wolf haplotypes were found in the wolf sample, however, no coyote haplotypes were found. The authors used the species attribution criteria of Wilson et al. (2000) to describe the observed haplotypes. In the combined sample of western Great Lakes states wolves from Minnesota, Wisconsin and Michigan (n = 203), *C. lycaon* mtDNA control region sequence haplotypes C1, C3 and C13 were more prevalent (n = 134, 66%) than *C. lupus* haplotypes C22, C23 and C97 (34%). In contrast, wolves from northern Ontario (n = 135) more commonly exhibited *C. lupus* haplotypes (56%) than *C. lycaon* haplotypes (44%). Geospatial maps of the distribution of *C. lycaon*, *C. lupus* and *C. latrans* mtDNA haplotypes revealed that they occur together throughout the western Great Lakes region (Wheeldon 2010, Fig. 1).

Furthermore, 44% of male western Great Lakes states wolves (n=43) displayed both *C. lupus* and *C. lycaon* mtDNA and Y-chromosome markers. The authors concluded that the wolves of the western Great Lakes states region are hybrids of gray wolves *C. lupus* and eastern wolves *C. lycaon*. Unlike *C. lycaon* in southeastern Ontario, which has hybridized extensively with coyotes (Grewal et al. 2004, Rutledge et al. 2010b), no wolves were found to exhibit coyote

mtDNA haplotypes and only one coyote was found with a wolf mtDNA haplotype (Wheeldon et al. 2010, Table 1).

Koblmüller et al. (2009a) addressed questions regarding the evolutionary origin of Great Lakes wolves as well as historical and ongoing hybridization between Great Lakes wolves, western gray wolves and coyotes. Koblmüller et al. (2009a) compared the mtDNA control region sequence haplotypes of modern and historical (n = 19, 1892-1916) wolves from the Great Lakes region as well as reference populations of coyotes and western gray wolves (*C. lupus*). Although these data were not presented in such a way as to be able to determine the respective haplotype frequencies observed in wolves from the various modern (n = 6) and historical (n = 7) Great Lakes region sampling localities, the authors summarized their results with a phylogenetic analysis (neighbor-joining tree), and identified coyote (containing all coyote individuals) and wolf clades (containing all western *C. lupus* individuals). The analysis placed most Great Lakes wolves (n = 142) in the coyote clade, but they were also well-represented in the wolf clade (n = 75). The authors concluded that Great Lakes wolves are gray wolves (*C. lupus*) that have been influenced by repeated hybridization with coyote in both ancient and recent times. They also concurred with the conclusions of Leonard and Wayne (2008) that historical Great Lakes wolves were characterized by phylogenetically distinct haplotypes representing an endemic “American wolf” and that these have been replaced by a modern population of mixed *C. lupus* and coyote ancestry. The autosomal and Y-chromosome microsatellite DNA data obtained in this study are summarized elsewhere.

Muñoz-Fuentes et al. (2009, Figure 3, which is reproduced here as Figure 11) observed a pronounced differentiation in mtDNA control region sequence variation between coastal and inland wolves in British Columbia and other localities in western Canada and Alaska. They



attributed this distinction to local discontinuities in vegetation cover and prey composition and preferences of dispersing wolves for habitats similar to where they were reared.

Although genetic differentiation between all localities was significant, coastal and inland populations were most different and haplotype frequency and composition among coastal wolves were markedly different (Table 4 of this paper). Differentiation between historical samples of *C. l. nubilus* (n = 19, Leonard et al. 2005) and modern *C. l. occidentalis* (n = 118, Muñoz-Fuentes et al. 2009, Table 2) was high ( $\Phi_{ST} = 0.124$ , where  $\Phi_{ST}$  is a measure of DNA haplotype divergence [Excoffier et al. 1992] ), but differentiation between coastal British Columbia wolves (n = 75) within the range of *C. l. nubilus* in Nowak (1995) and historical *C. l. nubilus* was even greater ( $\Phi_{ST} = 0.550$ ). Muñoz-Fuentes et al. (2009) observed that this differentiation supported the recognition of coastal British Columbia wolves as a discrete management unit (Moritz 1994).

Assessments of mtDNA control region sequence variation by Muñoz-Fuentes et al. (2010) detected evidence of wolf-dog hybridization in wolves that recolonized Vancouver Island after extirpation in the 1950s. The single mtDNA haplotype lu68 found in all 13 available pre-extirpation specimens (collected 1910-1950) was observed in only 5% of the recovered Vancouver Island population and 25% of the coastal mainland population (Muñoz-Fuentes et al. 2009). The recovered population primarily exhibited haplotype lu38 (95%), which was also common (68%) on the mainland coast of British Columbia (Muñoz-Fuentes et al. 2009a). Although the mtDNA data indicated evidence of dog introgression, the microsatellite data that were also collected in this study did not (discussed in the preceding section). These results were found to be consistent with a rare hybridization event that occurred early in the re-establishment of wolves on the island.

Rutledge et al. (2010a) tested the hypothesis that hundreds of years before the eastward expansion of coyotes, the wolf of the temperate forests of eastern Canada was a gray wolf. Four historical (circa 1500 to 1600) *Canis* skull fragments (i.e., three teeth and a lower mandible with two in situ teeth) were obtained from a pre-European contact Iroquois village archaeological site in southern Ontario, Canada. None of the remains exhibited gray wolf mtDNA, but tooth samples L1Va3 and L1Va5 had mtDNA sequence haplotypes previously found in ancient old world dogs (Vila et al 1997, Leonard et al. (2002), and the remaining tooth L1Va4 and mandible L1Va6 exhibited mtDNA haplotypes similar to coyotes from Saskatchewan (Rutledge et al. 2010a, Figure 5). Tooth sample L1Va4 had been found in context with the mandible L1Va6, and morphometric analysis identified this bone specimen as being outside the size range of coyotes and likely of eastern wolf origin. The authors concluded that the combined genetic and morphometric data suggest that the archaeological remains are of eastern wolf, *C. lycaon*, origin and that the historical wolf of eastern Canada was not the gray wolf *C. lupus*. However, they added that the data were also consistent with a *C. lycaon*–*C. lupus* hybrid origin (Wheeldon and White 2009).

Rutledge et al. (2010b) tested the hypothesis that eastern wolves have mediated gene flow between gray wolves and coyotes in the region of Algonquin Provincial Park, Ontario. Mitochondrial DNA control region sequence haplotypes were developed for wolves from northeastern Ontario (n=51), Algonquin Provincial Park (n=127) and Frontenac Axis (n=38). The autosomal and Y-chromosome microsatellite data also obtained in this study are discussed in respective sections.

The frequency and composition of the mtDNA haplotypes observed were consistent with the observations of Wilson et al. (2000) and Grewal et al. (2004) for wolves in Algonquin

Provincial Park (Rutledge et al. 2010b, Table 3). Wolf clade mtDNA haplotypes were observed at high frequency in northeastern Ontario (53%), low frequency in Algonquin Provincial Park (7%) and were absent in the southern-most, Frontenac Axis sample. Coyote clade mtDNA haplotypes attributed to *Canis lycaon* were common in both northeastern Ontario (14%) and Algonquin Provincial Park (16%), and occurred at high frequency in Frontenac Axis wolves (61%). Coyote haplotypes were observed at moderate frequency in northeastern Ontario (33%) and Frontenac Axis (39%) but occurred at highest frequency in Algonquin Provincial Park wolves (77%; Table 4 of this paper). Female breeders at Algonquin Provincial Park had a higher frequency of *C. lycaon* haplotypes (36%) than the total sample. These results are in general agreement with the separation of *C. l. nubilus* and *C. l. lycaon* range along a transect from the juncture of Lake Superior and Lake Huron eastward to the Quebec border (Nowak 1995).

Fain et al. (2010) examined the taxonomic relationships of wolves in the western Great Lakes states and tested the influence of coyote hybridization on wolf recovery in the region. Mitochondrial DNA control region sequence haplotypes were developed for wolves sampled from Minnesota (n = 42), Wisconsin (n = 65), Upper Peninsula Michigan (n = 17), and western wolves from Alaska (n = 40), British Columbia (n = 25), and Alberta (n = 25). The study also included Wisconsin coyotes (n = 132). The autosomal and Y-chromosome microsatellite data also obtained in this study are discussed in the preceding and following sections, respectively.

The authors found the data from all three marker types to support the recognition of *Canis lycaon* as a unique species of North American wolf in the western Great Lakes states. The frequency and composition of the mtDNA haplotypes observed were consistent with the observations of Grewal et al. (2004) for wolves in northwest Ontario as fully two-thirds of the combined western Great Lakes states sample exhibited mtDNA haplotypes of *C. lycaon* (see

Fain et al. 2010, Figure 3 for species attribution of haplotypes). Also similar to northwest Ontario, there was substantial contribution from *C. lupus* as haplotypes lu28 and lu32 were observed in a third of the population (see Table 4 of this paper).

The geographic scale of the *C. lycaon*-*C. lupus* hybrid zone was indicated by the observation of *C. lycaon* mtDNA haplotypes as far west as northwestern Minnesota and *C. lupus* mtDNA haplotypes in eastern Upper Peninsula, Michigan (Fain et al. 2010, Fig. 2). The *C. lupus*- and *C. lycaon*-derived mtDNA haplotypes observed in western Great Lakes wolves were exclusive of sympatric *C. latrans* and inconsistent with ongoing hybridization with coyotes.

Stronen et al. (2010) combined morphological characters (body mass and skull features) and species-specific mtDNA control region sequence haplotypes to assess wolf-coyote hybridization in wolves from Riding Mountain National Park (n = 19) and Duck Mountain Provincial Forest (n = 8) in Manitoba. Additional wolf samples from northern Manitoba (n = 13) and Prince Albert National Park, Saskatchewan (n = 24) were characterized for mtDNA haplotype only.

The eastern wolf mtDNA haplotype C3 was observed in 50% of the Duck Mountain Provincial Forest sample but was not observed elsewhere in Manitoba or Saskatchewan (see Stronen et al. 2010, Figure 3, for species attribution of haplotypes). The *C. lupus* haplotypes C22 and C23 predominated in Manitoba (25% and 63% respectively) whereas only *C. lupus* haplotypes C23 and 16 were observed in wolves from Prince Albert National Park, Saskatchewan (17% and 83% respectively). This study provided further definition to eastern wolf range; the authors concluded that the western range of eastern wolves did not extend to Saskatchewan (Table 4 of this paper).

In a previous study of Alaskan wolves, Weckworth et al. (2005) found that coastal and continental groups were distinguished by significant differences in allele frequencies at autosomal microsatellite loci, a criterion used to identify management units for conservation (Moritz 1994). Additional criteria are the exhibition of significant differences in mtDNA haplotype frequency for management units or reciprocal monophyly in the inheritance of mtDNA variants by the members of an evolutionarily significant unit. To test for compliance under these criteria, Weckworth et al. (2010) evaluated the mtDNA of the same individuals characterized in their earlier study.

Haplotype variation was assessed in coastal southeast Alaska wolves (n = 129), coastal Gulf of Alaska wolves (n = 73) and wolves from interior localities in Alaska (n = 64), Yukon (n = 12), and British Columbia (n = 24). Although the mtDNA haplotypes identified in this study comprised a portion of the cytochrome *b* gene, tRNAs for proline and threonine as well as the control region, only the control region portion was used in this review. The authors observed a pronounced differentiation in mtDNA control region sequence variation between coastal wolves in the southeast archipelago of Alaska and coastal wolves from the Kenai Peninsula and Copper River delta in the Gulf of Alaska. Similar differences were found in comparisons to populations in interior Alaska and British Columbia. Although genetic differentiation among all localities was significant, differentiation between coastal and inland populations was greatest and haplotype frequency and composition among coastal wolves was unique. As evidence, haplotype H (Weckworth et al. 2010) also known as lu68 (Muñoz-Fuentes et al. 2009, 2010) is unique to southeast Alaska and coastal British Columbia (Table 4 of this paper). The authors attributed this distinction to independent evolutionary histories for coastal and continental wolves in North America.

1572

1573 *Summary of studies of mtDNA.* The initial interpretation of mtDNA variation in the Great Lakes  
1574 region as resulting from coyote-wolf introgression (Lehman et al. 1991) was reinterpreted by  
1575 Wilson et al. (2000) and in later studies (Fain et al. 2010, Wheeldon et al. 2010) as the result of  
1576 gray wolf-eastern wolf hybridization. Regardless of disagreements over the identity of the taxa  
1577 involved, there is general agreement that there was a unique historical wolf population in the  
1578 Great Lakes region that has subsequently been affected by hybridization (Wilson et al. 2000,  
1579 2003; Leonard and Wayne 2008; Koblmüller et al. 2009a; Fain et al. 2010; Wheeldon et al.  
1580 2010).

1581       Early reports indicated red wolves had either gray wolf or coyote mtDNA and  
1582 cytochrome *b* gene haplotypes (Wayne and Jenks 1991, Roy et al. 1996). However, surviving  
1583 red wolves all have a mtDNA control region haplotype unique to red wolves (Wilson et al. 2000,  
1584 Adams et al. 2003, Hailer and Leonard 2008).

1585       The initial observation (Wilson et al. 2000) of the phylogenetic clustering of red wolves  
1586 and eastern wolves has not been found in subsequent studies (Wilson et al. 2003, Leonard and  
1587 Wayne 2008, Fain et al. 2010).

1588       There is agreement on the phylogenetic uniqueness of the mtDNA control region  
1589 haplotypes of historical and contemporary Mexican wolves (Vilà et al. 1999, Leonard et al.  
1590 2008), but a single exception is discussed in the Analysis and Discussion.

1591

1592 *Y-chromosome DNA.* In a study designed primarily to explore the pack composition of wolves in  
1593 Algonquin Provincial Park, Grewal et al. (2004) reported on Y-chromosome microsatellite  
1594 haplotype variation among wolves (n=248) at various locations in Ontario and Quebec.

Haplotypes were developed for the Y-linked microsatellite loci MS41A, MS41B, MS34A, MS34B characterized by Olivier et al. (1999). Sixteen different haplotypes were identified (Table 5 of this paper), but species assignments were not made for any of the observed haplotypes. Four to eight haplotypes were observed in each of the six locality samples. No haplotype was common to all localities, but the haplotypes AA and CE were each found at five localities. Seven haplotypes were specific to single localities.

Consistent with the separation of *C. l. nubilus* and *C. l. lycaon* range (Nowak 1995), differentiation between wolf populations from northern (i.e., northeastern Ontario, Abitibi-Temiscamingue Region, La Verendrye Reserve) and southern (i.e., Algonquin Provincial Park, Frontenac Axis, Magnetawan Region) localities was high ( $\Phi_{ST}$  ranged 0.111 to 0.325), while differentiation of wolves from within the respective regions was low ( $\Phi_{ST}$  ranged -0.057 to 0.073).

Bannasch et al. (2005) used paternally inherited Y-chromosome haplotypes from the Y-linked microsatellite loci MS34A, MS34B and MS41B (Olivier et al. 1999), and 650-79.2, 650-79.3 and 990-35 described in their own study, to evaluate the genetic structure of purebred dogs. A total of 824 unrelated male domestic dogs from 50 different breeds were analyzed. Sixty-seven different haplotypes were observed and many provided genetic distinction between breeds. Twenty-six breeds exhibited breed-specific haplotypes indicating lengthy independent histories. Y-chromosome haplotype sharing between a number of breeds reflected a shared origin, and many of these relationships were confirmed by known breed histories. Neither wolves nor coyotes were included in this study.

Sundqvist et al. (2006) characterized Y-chromosome haplotype variation in a study of the origin of domestic dog breeds. Four Y-linked microsatellite loci (Olivier et al. 1999) were

analyzed in wild gray wolves from Alaska ( $n = 12$ ) and Northwest Territories ( $n = 13$ ). Eleven different haplotypes were observed, only one of which (H32) was found in both samples (Table 5 in this paper).

Musiani et al. (2007) analyzed Y-chromosome haplotype variation in male gray wolves from boreal forest ( $n = 32$ ) and tundra/taiga ( $n = 170$ ) habitat in Alberta and Northwest Territories, Canada ( $n = 202$ ). Four Y-linked microsatellite loci (Olivier et al. 1999) were analyzed, nineteen different haplotypes were observed and nine occurred in  $> 5\%$  of the sample (Table 5 in this paper). Although haplotype frequencies differed between habitat types, genetic differentiation ( $F_{ST} = 0.03$ ) was not significant between male boreal forest and tundra/taiga wolves, contrary to the significant differentiation in mtDNA ( $F_{ST} = 0.28$ ) observed between habitat types. In addition, Y-chromosome gene diversity was similar in wolves from boreal forest and tundra/taiga habitat (i.e., 0.897 and 0.891 respectively). In contrast to results obtained in this study from mtDNA control region sequence and autosomal microsatellite variation (discussed in previous sections), the Y-chromosome haplotype data did not support the characterization of boreal forest and tundra/taiga wolves as separate populations.

Hailer and Leonard (2008) assessed cross-species introgression in red wolf, Mexican wolf, and Texas coyotes using Y-chromosome microsatellite haplotypes from four Y-linked microsatellite loci (Olivier et al. (1999) as species markers. The sample included male wolves from the captive breeding programs for *C. l. baileyi* ( $n = 16$ ) and *C. rufus* ( $n = 5$ ) as well as male *C. latrans* from Texas ( $n = 34$ ) and Nebraska ( $n = 36$ ). These were compared to each other and with similarly characterized reference gray wolves ( $n = 226$ ) from previously-published studies (Sundqvist et al. 2001, 2006; Musiani et al. 2007).



Two different Y-haplotypes were identified among *C. rufus*, the coyote-like type H15 that was also observed in 6% of the Texas coyote sample, and H1 that the authors recognized as wolf-like and most similar to haplotypes H28 and H29 found in captive Mexican wolves (Hailer and Leonard 2008, Figure 3). They suggested that hybridization with Mexican wolves may have been the route by which this marker entered Texas red wolves. Haplotype H28 was also found in wolves from Alaska (equals haplotype G, Table 5), but both H1 and H29 were unique among wolves (Table 5 in this paper) and coyotes.

A total of 59 unique Y-chromosome haplotypes were identified in the Texas and Nebraska coyote samples, but only three haplotypes were common to both samples. A single Texas coyote exhibited haplotype H2, which was phylogenetically most similar to gray wolf haplotype H44 (Hailer and Leonard 2008, Figure 3). Although this haplotype was also found in a Northwest Territories wolf (Musiani et al. 2007), it may also have been present in historical wolves through their southern range. The authors suggest that introgression between Mexican wolves and coyotes may have been the route by which this marker entered the Texas coyote population.

Koblmüller et al. (2009a) also studied Y-chromosome haplotype variation in wolves of the Great Lakes region. Y-chromosome haplotypes from the four Y-linked microsatellite loci characterized by Olivier et al. (1999) and 650-79.3 and 990-35 characterized by Bannasch et al. (2005) were analyzed in male gray wolves ( $n = 30$ ) and Great Lakes wolves ( $n_{\text{Modern}} = 111$ ,  $n_{\text{Historic}} = 4$ ). The autosomal microsatellite and mtDNA sequence data also obtained in this study are discussed in preceding sections. It was not possible to determine from the presented information how the observed haplotype frequencies were distributed across the different

collection localities in the study (i.e., Minnesota, Wisconsin and Michigan, U.S., and Northwest Territories, Alberta, Ontario, and Quebec, Canada).

Seventeen different haplotypes were observed in the western gray wolf sample, 41 in the Great Lakes wolf sample, with nine haplotypes common to both samples (Koblmüller et al. 2009a, Table 2). None of these haplotypes were named, nor did the authors provide size information about the alleles from which the haplotypes were compiled. As a result, it was not possible to equate the Y-chromosome haplotypes identified by Koblmüller et al. (2009a) with those identified in other similar studies (Table 5 in this paper). A median-joining network of Y-chromosome haplotypes identified a major divide between coyote and wolf clades. Nearly all coyotes were found in the coyote clade and all western wolves occurred in the wolf clade, but haplotypes of Great Lakes wolves were distributed among both clades. The four historical Great Lakes wolves for which Y-chromosome haplotypes were determined were all from the Upper Peninsula of Michigan (J. Leonard, Uppsala University, personal communication). The analysis placed all of these specimens in the wolf clade, suggesting that *C. lupus* was interbreeding with Great Lakes wolves prior to their near-extirpation (also see Wheeldon & White 2009, Rutledge et al 2010a).

Rutledge et al. (2010b) tested the hypothesis that eastern wolves have mediated gene flow between gray wolves and coyotes in the region of Algonquin Provincial Park, Ontario. Y-chromosome microsatellite haplotypes were developed from four Y-linked microsatellite loci (Olivieri et al. 1999) for wolves from northeastern Ontario (n=26), Algonquin Provincial Park (n = 53), and Frontenac Axis (n = 19). The autosomal microsatellite and mtDNA sequence data also obtained in this study are discussed in preceding sections.

The frequency and composition of the Y-chromosome haplotypes observed were consistent with the observations of Grewal et al. (2004, Table 2) for wolves in Algonquin Provincial Park (Rutledge et al. 2010b, Table 4). The authors used the species assignment method of Shami (2002) to describe the observed haplotypes. *Canis lupus* Y-chromosome haplotypes occurred at high frequency in northeastern Ontario (96%) and moderate frequency among Algonquin Provincial Park (17%) and Frontenac Axis wolves (21%). *Canis lycaon* Y-chromosome haplotypes were common in both Algonquin Provincial Park (75%) and Frontenac Axis (47%) but only one wolf from northeastern Ontario (4%) exhibited a *C. lycaon* Y-chromosome haplotype. Breeding males at Algonquin Provincial Park had the highest representation (88.9%) of *Canis lycaon* haplotypes. *C. latrans* haplotypes were common in the Frontenac Axis sample (32%), rare in Algonquin Provincial Park wolves (8%) and were absent among northeastern Ontario wolves.

Similar to the results from mtDNA comparisons, the Y-chromosome results also agreed with the separation of *C. l. nubilus* and *C. l. lycaon* range along a transect from the juncture of Lake Superior and Lake Huron eastward to the Quebec border (Nowak 1995). Y-chromosome haplotypes of *C. lupus* origin were five-fold more frequent in northeastern Ontario than to the south where *C. lycaon* haplotypes predominated.

Fain et al. (2010) analyzed Y-chromosome and mtDNA haplotypes in addition to autosomal microsatellite variation to evaluate various genetic aspects of the recovered wolf population in the western Great Lakes states. Chief among these was their species distinction. The mtDNA and autosomal microsatellite data are discussed in preceding sections.

The Y-chromosome-linked microsatellite loci characterized by Olivier et al. (1999) and 650-79.2A, 650-79.2B, and 990-35 characterized by Bannasch et al. (2005) identified species-

specific Y-chromosome haplotypes in wolves and coyotes from Minnesota, Wisconsin, and Michigan (Upper Peninsula). Haplotypes were designated as being of either *C. lupus* or *C. lycaon*/*C. latrans* origin after the allele sizing method of Hailer & Leonard (2008). In the combined sample of male western Great Lakes states wolves (n = 61), half exhibited unique *C. lycaon* haplotypes and the other half *C. lupus* haplotypes (Fain et al. 2010, Table 1). A median-joining network identified a major divide between coyote and wolf haplotypes (Fain et al. 2010, Figure 4, which is reproduced here as Figure 12). The 30 coyote Y haplotypes observed were all in their own clade, the fourteen *C. lupus* Y haplotypes were placed together in a second clade, and the five Y haplotypes unique to Great Lakes wolves were primarily distributed in a third clade intermediate between gray wolves and coyotes. Although Y haplotype FWSClyR was placed in the coyote clade, the authors interpreted this to be the result of ancient introgression and that this haplotype was being transmitted as a *C. lycaon* lineage marker.

Forty-six percent of male western Great Lakes states wolves displayed both *C. lupus* and *C. lycaon* mtDNA and Y-chromosome lineage markers, 41% of males exhibited both *C. lycaon* mtDNA and Y-chromosome haplotypes, but only 13% exhibited both *C. lupus* mtDNA and Y-chromosome haplotypes. The authors concluded that the western Great Lakes states wolf population was composed of gray wolves, *C. lupus*, eastern wolves, *C. lycaon*, and their hybrids. Moreover, the *C. lupus* and *C. lycaon* derived mtDNA and Y-chromosome haplotypes observed in western Great Lakes states wolves were exclusive of *C. latrans* and inconsistent with ongoing hybridization with coyotes.

Wheeldon et al. (2010) evaluated species distinction among the wolves and coyotes of the western Great Lakes region. They investigated Y-chromosome haplotypes based on four Y-chromosome-linked microsatellite DNA loci (Olivieri et al. (1999) in 209 male wolves and

coyotes from the western Great Lakes states (Minnesota, Michigan, and Wisconsin) and western Ontario (Table 5 in this paper). Haplotypes were designated as being of either gray wolf or coyote/eastern wolf types after the allele sizing method of Hailer & Leonard (2008), 11 were gray wolf types and 18 were coyote/eastern wolf types. Two of the 29 haplotypes observed were shared by wolves and coyotes.

Forty-four percent of male Great Lakes states wolves displayed both *C. lupus* and coyote/eastern wolf lineage markers. Twenty-nine percent of males exhibited both coyote/eastern wolf mtDNA and Y-chromosome markers, and 27% exhibited both *C. lupus* mtDNA and Y-chromosome markers. The authors concluded that the wolves of the western Great Lakes region are hybrids of gray wolves (*C. lupus*) and eastern wolves (*C. lycaon*). Autosomal microsatellite DNA variation and mtDNA haplotypes from this study were summarized in previous sections.

*Summary of studies of Y-chromosome DNA.* Studies of wolves in the Great Lakes region have found haplotypes distributed in either a wolf clade (*C. lupus*), or a clade that includes eastern wolf and coyote. These studies differ in characterizing eastern wolf haplotypes as alternatively *Canis lycaon* (Fain et al. 2010, Rutledge et al. 2010a, Wheeldon et al. 2010), Great Lakes wolf (Koblmüller et al. 2009a), or coyote (Koblmüller et al. 2009a). Studies covering the western Great Lakes region differ in concluding that wolf-coyote hybridization has been rare (Fain et al. 2010, Wheeldon et al. 2010), or that that it has occurred recently and is currently ongoing (Koblmüller et al. 2009a).

Studies that presented data on both the mtDNA and Y-chromosome haplotypes of individual male wolves of the western Great Lakes region agree that the population consists of

gray wolves, eastern wolves, and individuals that are admixed with respect to these two kinds of wolves (Fain et al. 2010, Wheeldon et al. 2010).

In contrast to the unique, coyote clade mtDNA haplotype of captive red wolves, one of the two Y-chromosome haplotypes found in red wolves was identical to a haplotype found in Texas coyotes (Hailer and Leonard 2008), and the other was identical to a haplotype found in domestic dogs (Bannasch et al. 2005).

## **Analysis and Discussion**

Views vary on the number and identity of modern species of *Canis* in North America. There is general agreement only that coyote is a separate species, and that dogs are derived from *Canis lupus* (Vilà et al 1997,1999). The predominant view of the taxonomy of those surviving North American *Canis* known by the general vernacular name of “wolves” follows Goldman (1944) in recognizing two species: *Canis lupus* (including the subspecies *Canis lupus lycaon*) and *Canis rufus* (Nowak 1979, 1985; Hall 1981). A variant of this view incorporates the contention that *Canis rufus* is the result of hybridization between *C. lupus* and coyote (Wayne and Jenks 1991). Wilson et al. (2000) also recognize two wolf species, but propose that eastern wolf, *Canis lycaon*, is the same species as *C. rufus*, and that these together are specifically distinct from *C. lupus*. Baker et al. (2003) recognize three species: *C. lupus*, *C. lycaon*, and *C. rufus*. Wilson and Reeder (2005) include all North American wolves in *C. lupus*, which includes subspecies *C. l. rufus* and *C. l. lycaon*.

The following analysis and discussion will first address the number of species of wolf in North America. It will begin with probably the most contentious question of whether the eastern wolf is within the species limits of *C. lupus*. The taxonomic status of red wolf and eastern wolf

with respect to each other and coyote will then be addressed. Last, the subspecies classification within *C. lupus* will be evaluated.

#### **Species limits of *Canis lupus* relative to eastern wolf**

The most contentious issue related to the species limits of *C. lupus* in North America is the placement of the eastern wolf, which has also been referred to as the Great Lakes wolf (Leonard and Wayne 2008): Is the eastern wolf within the species limits of *C. lupus* as either a subspecies, *C. l. lycaon* (Goldman 1937) or a unique ecotype (Koblmüller et al. 2009a), or does it represent a different species, *C. lycaon*, outside the species limits of *C. lupus*? This section assesses whether populations referred to as eastern wolves should be considered members of *C. lupus*.

The positions of various authors of taxonomic studies on the geographic range of the eastern wolf were summarized earlier in the section on taxonomy. All extant wolves that might be assigned to the eastern wolf occur in the general area from southern Ontario and Quebec, west to Minnesota and Manitoba. Wolves in this range were nearly exterminated and, by the 1970s, the only known wolves remaining in the conterminous United States were in northeastern Minnesota (Figure 1 of this paper, Mech 1974). At about this time, wolves had been eliminated from southern Ontario and Quebec (Mech 1974) and replaced by a population of coyotes that had been influenced by hybridization with wolves (Kolenosky and Standfield 1975, Rutledge et al. 2010b). Wolves have subsequently expanded their range in Minnesota and re-occupied Wisconsin and the Upper Peninsula of Michigan.

The eastern wolf and gray wolf are known to interbreed within the Great Lakes region. Where available data permit, the approach here will be to first evaluate divergence between

eastern wolves and gray wolves, then to attempt to evaluate the implications of admixture for determining potential inter-species barriers.

*Morphometrics.* Trends of increasing size among wolves to the north and west of southern Ontario and Quebec have been noted in morphometric studies covering the Great Lakes region. The association of smaller wolves with white-tailed deer in deciduous forests and larger wolves with larger prey, such as moose and caribou, in boreal forests has been frequently cited (Kolenosky and Standfield 1975, Skeel and Carbyn 1977, Schmitz and Kolenosky 1985). When wolf skulls were divided by source habitat into deciduous forest (eastern wolf) and boreal forest (*C. lupus*), discriminant function analysis distinguished 75% of boreal wolves from eastern wolves, and boreal wolves were over 25% larger in body mass (Kolenosky and Standfield 1975). This size difference cannot be entirely attributed to interbreeding of eastern wolves with coyotes because the “Tweed wolves” of southern Ontario, which have been influenced by recent coyote introgression, were excluded from these samples. Discriminant function analysis of additional skull and body measurements have confirmed the smaller size of eastern wolves relative to gray wolves and distinguish them from coyotes and coyote-wolf hybrids (Schmitz and Kolenosky 1985). Moreover, gray-fawn coat color was most common in both samples, but black, cream, and white colors found in boreal wolves were rare in eastern wolves.

Skeel and Carbyn’s (1977, Figures 2, 3) principal components analysis places eastern wolves intermediate between *C. l. nubilus* and *C. l. occidentalis* (following Nowak’s [1995] revision) on the first principle component axis, but are closer to *C. l. nubilus*. Their eastern wolf sample was from extreme southwestern Ontario in an area where influence by *C. l. nubilus* can be expected.



Nowak's (1979, Figure 7) discriminant function analysis of skull features found the individuals he attributed to *C. l. lycaon* (eastern wolf) to be generally smaller than northern and western *C. lupus*. Within this eastern wolf sample, the individuals from the western range that he recognized for *C. l. lycaon* (Michigan, Minnesota, Wisconsin, and western Ontario) had a greater range of variation and several individuals were larger than wolves from southern Ontario and Quebec. Based on this and additional morphometric analyses (Nowak 1995, 2002, 2009), Nowak recognizes the eastern wolf as a subspecies of *C. lupus* and restricts its range to southern portions of Ontario and Quebec, while attributing Minnesota wolves to *C. l. nubilus*. His Minnesota samples, however, were taken after 1960 (Nowak 1995) or 1970 (Nowak 2002, 2009), during a period of likely increased movement into Minnesota of *C. l. nubilus* from the west and north (Mech and Frenzel 1971, Mech and Paul 2008, Mech 2010, Mech and Nowak, in review). Nowak's data for Minnesota likely reflect this substantial and recent contribution of *C. l. nubilus* to Minnesota wolves. As will be discussed later, the genetic data (Fain et al. 2010, Wheeldon et al. 2010) indicate a substantial genetic contribution from the eastern wolf throughout the western Great Lakes to Minnesota and western Ontario.

The utility of Nowak's (1995) analysis is limited in determining whether the eastern wolf is distinct from *Canis lupus* by his inclusion of recent wolves of probable *C. l. nubilus* origin in samples intended to represent the eastern wolf. The sample from Algonquin Provincial Park is the only eastern wolf sample in these studies that has not been greatly influenced by gray wolves. When Algonquin wolves are identified as such on discriminant function plots (Nowak 1995, Figures 10, 11), most individuals appear outside the polygons representing *C. lupus*. Additional eastern wolves from southern Ontario and Quebec produced a similar result (Nowak 2002, Figure 8; 2009, Figure 15.2).

The eastern wolf is smaller than other wolves in the analysis, and geographical trends in size correspond to habitat differences and support the proposal of Kolenosky and Stanfield (1975) of prey specialization of smaller wolves on white-tailed deer in deciduous forests. Morphometric data do not provide direct information on monophyly or reproductive relationships, but in this case the geographic trends in body size are consistent with habitat distributions and ecological specialization. All morphometric studies find the eastern wolf to be an outlier to *C. lupus*, and where there is some overlap in morphometric space, the eastern wolf extends well beyond the limits of *C. lupus*.

*Microsatellite DNA.* The variably admixed nature of eastern wolf populations in the western Great Lake states determined from microsatellite DNA analysis has been noted by Fain et al. (2010) and Wheeldon et al. (2010). The conclusion of Koblmüller et al. (2009a) of little admixture in wolves of this region may have been influenced by the distant (Northwest Territories and Alberta) western gray wolf samples used for comparison. STRUCTURE analysis, as well as divergence measures such as  $F_{ST}$ , can be erroneously interpreted as indicating genetic discontinuities when there are significant gaps in sampling pattern (Pritchard et al. 2000, Schwartz and McKelvey 2009). Further, these western wolves likely represent *C. l. occidentalis*, rather than *C. l. nubilus*, the subspecies of gray wolf likely to be involved in admixture with the eastern wolf in the western Great Lakes.

There are indications in the STRUCTURE analysis presented by Koblmüller et al. (2009a, Figure 3) that there is more genetic variation in the wolf sample from the Great Lakes region than suggested by the outcome (Figure 3(c)) using  $K = 4$ . The  $L(K)$  values for  $K = 4$  and  $K = 5$  are indistinguishable in their Figure 3(a), and the range of variation for Great Lakes

wolves in the factorial correspondence analysis (their Figure 3(d) is much larger than that of coyotes and western gray wolves. Elevated levels of genetic variation are consistent with an admixed population.

Wheeldon et al. (2010) included samples from northwest Ontario that clearly portray admixture, and the higher allelic richness of western Great Lakes wolves (Fain et al. 2010) is also consistent with their genetically composite ancestry. Data from historical specimens from Minnesota and Wisconsin (Wheeldon and White 2009, Koblmüller et al. 2009a) also suggest that admixture of the eastern wolf and western *C. lupus* had taken place prior to their extirpation from the region.

Both the microsatellite and morphometric data indicate the same trend from Algonquin Provincial Park in the east with increasing influence by *C. lupus* to the west. In the western Great Lakes, nearly all wolves have indications of admixture (Fain et al. 2010, Wheeldon et al. 2010), but individual wolves vary greatly in the proportion of eastern wolf influence in their microsatellite DNA genotypes. In other words, the microsatellite DNA data indicate that the wolves of the western Great Lakes region do not comprise a homogenous population, which is consistent with a composite origin and incomplete admixture.

The microsatellite DNA data distinguished eastern wolves from western *C. lupus* in neighbor-joining trees (Roy et al. 1994, García-Moreno et al. 1996, Wilson et al. 2000), multidimensional scaling (Roy et al. 1994, García-Moreno et al. 1996), log likelihood (Wilson et al. 2000), factorial correspondence (Wheeldon et al. 2010), probability of identity (Wilson et al. 2000), and STRUCTURE analyses (Koblmüller et al. 2009a, Wilson et al. 2009, Fain et al. 2010, Wheeldon et al. 2010). Eastern wolves are also distinguishable in studies that include *C. lupus* from northern Ontario (Rutledge et al. 2010b).

Despite the expectation that linearity between genetic distance measures and geographic separation is lost when samples are far apart (Paetkau et al. 1997, Schwartz and McKelvey 2009), genetic distance measures are greater for comparisons between western gray wolves and eastern wolves than for comparisons between paired samples with substantial eastern wolf composition (Table 6). There is geographic discontinuity between western gray wolf and eastern wolf samples in some studies, but greater divergence between eastern wolves and gray wolves is also found in studies that include grey wolf samples in close proximity to eastern wolves (Roy et al. 1994, Grewal et al. 2004, Wilson et al. 2009, Rutledge et al. 2010b).

Private alleles are another indication of the relative isolation of populations (Slatkin 1985). These are alleles that are found within a single locality or population, and it is inferred that they are locally restricted owing to isolation. Information on private alleles in North American wolf populations are summarized in Table 7. Private alleles are determined only with respect to other wolf populations; many of these alleles are shared with coyotes. The number of private alleles is much higher in populations including eastern wolf (southern Quebec and the western Great Lakes states) than in western gray wolves. Some private alleles are at relatively high frequencies, which is a further indication of a history of isolation. For example, four such alleles have frequencies ranging from 0.146 to 0.202 in samples of wolves from the western Great Lakes states (Fain et al. 2010), consistent with continuing isolation despite a history of admixture.

*Single nucleotide polymorphisms.* Principal components analysis of the SNP data places the Great Lakes wolf (eastern wolf) sample closer to gray wolves than to coyotes (vonHoldt 2011, Figure 3, Supplemental Figure S4).  $F_{ST}$  values between the Great Lakes sample and other North

American *Canis* samples range from 0.05 for western gray wolves to 0.11 for Mexican wolf, red wolf, and coyote. In comparison,  $F_{ST}$  values among coyotes from different regions (midwestern/southern, northeastern, and western) range from 0.02 to 0.05 (vonHoldt et al. 2011, Supplemental Table S3). Successive STRUCTURE analyses grouped the Great Lakes sample with wolves, rather than coyotes, beginning at  $K = 3$ , although substantial assignment to coyote ancestry is evident. Assignment of gray wolf ancestry to individual Great Lakes wolves ranged from about 50% to 100%. vonHoldt et al. (2011) conclude that Great Lakes wolves are genetically distinct from western gray wolves.

Features of the sample of wolves from the Great Lakes region make it difficult to evaluate the taxonomic significance of the SNP data. The Great Lakes sample is small ( $n = 19$ ), and except for two individuals from Algonquin Provincial Park, their geographic origins within the region are identified only to the level of province or state. Mitochondrial DNA and Y-chromosome lineage markers (discussed in the next section) are the primary basis for recognizing an admixed population including both eastern wolf and gray wolf in the region (Wilson et al. 2000, Fain et al. 2010, Wheeldon et al. 2010), yet the haplotype composition on the sample used for SNP analysis is not reported. This sample may include both eastern and gray wolves, which is consistent with the observation that the proportion of gray wolf ancestry varies greatly among individuals in the sample. Because eastern wolf is interpreted as having a common ancestry with coyotes (Wilson et al. 2000), the gray wolf/coyote admixture in the Great Lakes sample may alternatively represent gray wolf/eastern wolf admixture. Inclusion of gray wolves in the Great Lakes sample would increase the similarity between the Great Lakes sample and western gray wolves, just as it has in the morphometric analysis by Nowak (1995). Finally, the first two axes of the principal components analysis (vanHoldt et al. 2011, Figure 3) account

for relatively little of the variance (10% and 1.7%, respectively); The first principal component separates dogs from the wild canids, but does separate wolves from coyotes. This separation of dog from wolves, from which they were derived, indicates that signatures of genealogical history that might be detectable in this analysis can be labile over the time period since dogs were domesticated.

*Mitochondrial and Y-chromosome haplotypes.* Studies using these maternally and paternally inherited lineage markers generally agree that North American *Canis* can be attributed to three clades: one representing *Canis lupus*, one coyotes, and the third representing members of wolf populations in the Great Lakes region attributed to eastern wolf (Wilson et al. 2000, Koblmüller et al. 2009a, Fain et al. 2010, Wheeldon et al. 2010). The current wolf population of the western Great Lakes includes haplotypes from both *C. lupus* and eastern wolf clades, with eastern wolf haplotypes predominating. However the unique, “coyote-like” eastern wolf haplotypes of the Great Lakes region may have originated, there seems to be agreement that they support the existence of a unique taxon in the region, although the modern population is admixed (Wilson et al. 2000, Leonard and Wayne 2008, Koblmüller et al. 2009a). There is also disagreement on whether it should be recognized as a species (Wilson et al. 2000) or as a subspecies (Nowak 1995), and some researchers decline to apply a scientific name and refer to it as an “endemic American wolf” (Leonard and Wayne 2008) or as a unique ecotype or taxon (Koblmüller et al. 2009a).

Wilson et al. (2000) have proposed that the relatively large lineage divergence in mtDNA of the eastern wolf from gray wolves to the north and west is sufficient to recognize it as a separate species. Wilson et al (2000, p. 2159) reported control region mtDNA haplotype

sequence divergence between eastern wolves and western *C. lupus* as 8% (Wilson et al. 2000, p. 2159), compared to the average sequence divergence of 2.9 % among major clades of *C. lupus* (Vilà et al. 1999). In other studies (summarized in Table 8), sequence divergences between eastern wolves and western gray wolves are about an order of magnitude larger than within-species divergences. Using the common estimate of 2% sequence divergence per million years for mtDNA (Brown et al. 1979), the most divergent eastern wolf haplotypes diverged from ancestral gray wolf haplotypes about 4 mya, which approaches the estimated age of the genus *Canis* (Kurtén and Anderson 1980). Within-species sequence divergences this large appear to be rare in mammals (Avise et al. 1998), and are likely the result of long isolation. Following the hypothesis of Wilson et al. (2000), a specifically distinct eastern wolf evolved in North America from a common ancestor with coyotes rather than from a more immediate common ancestor with *C. lupus*, which evolved in Eurasia (Kurtén and Anderson 1980, Wayne and Vilà 2003).

Both mtDNA and Y-chromosome haplotypes of eastern wolves are members of monophyletic clades that are highly divergent from those of gray wolves. Because eastern wolves are phylogenetically more associated with coyotes, inclusion of eastern wolves in the same species with gray wolves would make the latter paraphyletic with respect to coyotes, which violates strict phylogenetic species criteria. Alternative explanations for paraphyly of a *C. lupus* that includes the eastern wolf involve incomplete lineage sorting, and hybridization with coyotes.

Incomplete lineage sorting within a broadly defined *C. lupus* is not a likely explanation for paraphyly because it is usually associated with relatively recent speciation events, and the highly divergent haplotypes of eastern wolves and coyotes indicate an ancient divergence. In addition, putative paraphyly is geographically localized to the Great Lakes region and there is no

evidence of it elsewhere within the large geographic range of overlap between *C. lupus* and coyotes.

That leaves hybridization as the remaining explanation for the paraphyletic inclusion of “coyote-like” Y-chromosome and mtDNA haplotypes in *C. lupus*. Koblmüller et al. (2009a) recognize the eastern wolf as a form of *Canis lupus* that owes its divergent genetic features to hybridization with coyotes, both ancient and ongoing, rather than to a long period of evolution as a separate lineage independent of *C. lupus*. A problem with recent coyote introgression as an explanation for the divergent mtDNA haplotypes of the eastern wolf is that these haplotypes are not found in coyotes, except those that have recently hybridized with the eastern wolf in southern Ontario and now occupy the northeastern United States (Kays et al. 2009, Rutledge et al. 2010b). The coyote-introgression hypothesis requires that the coyotes involved were of Y-chromosome and mtDNA haplotype clades that have subsequently become extinct. Although there is evidence, discussed later, of introgression of ancient coyote lineages that are now found only in the eastern wolf, such instances appear to be rare. The rarity of ancient coyote-eastern wolf introgression indicates that eastern wolves have been evolving as a separate lineage for a considerable time.

There is also disagreement among researchers on whether introgression is ongoing between coyotes and wolves in Minnesota, Wisconsin, and Michigan. Those favoring ongoing hybridization view western coyotes that have recently moved eastward as the source of the unique but “coyote-like” mtDNA and Y-chromosome haplotypes in eastern wolves (Lehman et al. 1991, Leonard and Wayne 2008, Koblmüller et al. 2009a). The contrasting view of little or no ongoing introgression from coyotes in this area interprets these haplotypes as a consequence of shared ancestry between coyotes and eastern wolves (Wilson et al. 2000, Wheeldon and White



2009, Fain et al. 2010, Wheeldon et al. 2010). The absence of western coyote Y-chromosome or mtDNA haplotypes in the current wolf population of Minnesota, Wisconsin, and Michigan is inconsistent with the hypothesis of ongoing coyote introgression in this area (Fain et al. 2010, Wheeldon et al. 2010).

The distinctive “coyote-like” mtDNA haplotypes of the eastern wolf could have resulted from ancient introgression from now-extinct coyote lineages, followed by a selective sweep. This assumes that the ancestral population of the eastern wolf had haplotypes from the wolf lineage, and therefore of Old World origin, but that it hybridized with ancient coyotes to such an extent that the original wolf-lineage mtDNA haplotypes were entirely replaced by coyote haplotypes. This requires a mating advantage of female coyotes in coyote-wolf mating, or a strong selective advantage for the mitochondrial genome of coyotes. The general process of haplotype replacement by a selective sweep is sometimes called cytoplasmic capture or mitochondrial capture. It has been reported in various plant and animal taxa (Avice 2004, Table 7.6), and can even result in total replacement of the mtDNA of one species by the mtDNA of another (Nevada et al. 2009).

Distinguishing mitochondrial capture from lineage divergence or incomplete lineage sorting can be difficult. Although it does occur, it is uncertain whether it occurs often enough to serve as a general explanation of species-level paraphyly in animal taxonomy. In a review of paraphyly in bird species, McKay and Zink (2010) found that most cases were the result of taxonomic errors or incomplete lineage sorting, with few clear instances involving hybridization. In addition to mtDNA, the eastern wolf differs from coyotes and gray wolf in that it also has distinguishing morphological, ecological, Y-chromosome, and nuclear autosomal DNA

characteristics. Moreover, there is no indication that such selective sweeps have occurred elsewhere in North American wolves.

Y-chromosome haplotypes can also be separated into wolf and coyote clades, with the distinctive haplotypes of eastern wolves basal to the coyote clade (Koblmüller et al 2009a, Fig 2(b); Fain et al. 2010, Figure 12 of this paper). As with mtDNA, inclusion of eastern wolf with *C. lupus* forms a paraphyletic group with respect to coyotes. An explanation of the high incidence of coyote-clade, Y-chromosome haplotypes in the eastern wolf through coyote introgression and displacement of wolf-clade haplotypes requires a selective advantage of coyote Y-chromosomes or a mating advantage of male coyotes over male gray wolves, which is the opposite of the usual relationship (Lehman et al. 1991) and seems unlikely.

It is reasonable based on their relative divergence from coyote mtDNA haplotypes to regard the most strongly divergent eastern wolf mtDNA haplotypes C1 and C3 (Wilson et al. 2000) and Y-chromosome types E, O, Y, and X (Fain et al. 2010) as indications of the initial divergence from their common ancestor with coyotes, and the less divergent, “coyote-like” haplotypes mtDNA C13 (Wilson et al. 2003) and Y-chromosome type R (Fain et al. 2010) as representing subsequent, ancient introgression. The more divergent Y-chromosome haplotypes of the eastern wolf appear intermediate between *C. lupus* and coyotes in the phylogenetic assessment of Fain et al. (2010, Figure 12 of this paper). In summary, species-level recognition of *Canis lycaon*, the eastern wolf, outside the species limits of *C. lupus* is supported by the phylogenetic distinctiveness of its mtDNA and Y-chromosome haplotypes. Inclusion of the eastern wolf within *C. lupus* would render the latter paraphyletic with respect to coyotes. The alternative explanation that these distinctive eastern wolf haplotypes are the result of introgression of a *Canis lupus* population by coyotes and replacement of wolf-clade haplotypes

by coyote haplotypes is not supported with evidence of these haplotypes in modern coyotes other than those that have recently hybridized with *C. lycaon* in eastern Canada (Rutledge et al 2010b).

*Conclusions on species limits of Canis lupus relative to the eastern wolf.* Available information on morphology, ecology, behavior, and genetics are concordant in supporting the existence of a unique form of wolf native to the Great Lakes region of North America. The small size of this eastern wolf in comparison to gray wolves is consistent with its association with white-tailed deer as its preferred prey among ungulate species. Genetic divergence measures at nuclear microsatellite DNA loci and a relatively high incidence of private alleles indicate that the eastern wolf has maintained cohesion and has had a long, separate evolutionary history from gray wolves. Mitochondrial DNA and Y-chromosome haplotypes indicate that eastern wolves and gray wolves evolved independently as separate lineages for a considerable time. These concordant lines of evidence and the age of lineage separation indicated by mtDNA haplotypes support the proposal of Wilson et al. (2000) that the eastern wolf and *Canis lupus* belong to separate species. The taxonomic interpretation best supported by available mtDNA and Y-chromosome haplotype data is that the eastern wolf evolved independently from a common ancestor with coyotes and independently of *C. lupus*.

The eastern wolf, as *Canis lycaon*, has also been recognized as a species-level taxon by Baker et al. (2003) in a recent revision of their checklist of North American mammals north of Mexico. The uniqueness of the Great Lakes wolf population, despite admixture, is recognized even by those who do not favor recognizing it as a separate species (Leonard and Wayne 2008, Koblmüller et al. 2009a).

It is remarkable that strong genetic signatures of separate evolutionary history remain detectable in the face of modern admixture of eastern wolves and gray wolves in the western Great Lakes region and eastern wolves and coyotes in the eastern Great Lakes region. It is notable that wolves from the two lineages have probably been in contact since *C. lupus*, an Old World lineage, appeared in North America over 500,000 years ago (Kurtén and Anderson 1980), yet mtDNA haplotypes of historical specimens (Wilson et al. 2003, Leonard and Wayne 2008, Wheeldon and White 2009, Rutledge 2010a) from the Great Lakes region are eastern wolf or coyote-like and not *C. lupus*.

The geographic range of the eastern wolf and the extent of its hybrid zone with *C. lupus* can be mapped using the geographic distribution of the mtDNA and Y-chromosome haplotypes of the two species (Wilson et al. 2000, 2009; Fain et al. 2010, Wheeldon et al. 2010). The recognition of the range of eastern wolf extending through Minnesota in the southwest and to Manitoba in the northwest (Stronen et al. 2010), reestablishes the western portion of the geographic range of eastern wolf recognized by Goldman (1944) and Hall (1981) for *C. l. lycaon*.

An historical reconstruction leading to the current relationship between the eastern wolf and *C. lupus* begins with *C. lupus* evolving in Eurasia, while the eastern wolf was evolving in North America from a common ancestor with coyotes (Wilson et al. 2000, Wheeldon and White 2009). North American *C. lupus* is inferred to have evolved in Eurasia based on fossils (Kurtén and Anderson 1980) and on the phylogenetic similarity of its haplotypes to certain Eurasian *C. lupus* (Wayne et al. 1995, Vilà et al. 1999, Wayne and Vilà 2003). At this time the eastern wolf and Eurasian *C. lupus* would have been reciprocally monophyletic, which is indicative of species-level distinction following strict phylogenetic species concepts and criteria.

When *C. lupus* subsequently invaded North America and came into proximity with the eastern wolf, the two species may have immediately started hybridizing, at least to some degree. Ecological isolation has been invoked as an explanation for the persistence of the two species or kinds of wolves (Standfield 1970). Differences in habitat and prey preference have been found to significantly affect the genetic structure of wolf populations in North America (Carmichael et al. 2001, Musiani et al. 2007) and Europe (Pilot et al. 2006). Preference of the eastern wolf for white-tailed deer in eastern deciduous forest habitats and of *C. lupus* for moose and caribou in more boreal habitats (Standfield 1970) may have limited encounters between the two species. However, with human-mediated conversion of boreal forests to deciduous forest and consequent expansion of white-tailed deer, contacts between the species would have increased. Lowered population densities of wolves as a result of persecution by people would have decreased the likelihood of encountering conspecific mates and increased the likelihood of inter-species matings. This “Allee effect” would have accelerated admixture.

There is little evidence for ongoing or recent hybridization between wolves and coyotes in the western Great Lakes states, but there are strong indications that admixture of eastern wolf and coyotes has occurred recently or is ongoing in the eastern Great Lakes region (Wilson et al. 2009, Kays et al. 2009, Rutledge et al. 2010b). The finding of 400-500 year old “wolves” in archaeological context having coyote and dog mtDNA (Rutledge et al. 2010a) suggests interbreeding between eastern wolves and coyotes had occurred long before 1919, the earliest historical record of coyotes in southern Ontario (Nowak 1979, p. 15).

An admixed population of *C. lupus* and the eastern wolf occurs across an area extending from eastern Ontario to Minnesota and into central Manitoba (Wilson et al. 2000, 2009, Grewal et al. 2004, Wilson et al. 2009, Fain et al. 2010, Stronen et al. 2010, Wheeldon et al. 2010). The

incidence in Minnesota, Wisconsin and Michigan of 36% of male wolves with the Y-chromosome marker of one species and the mtDNA marker of the other (Fain et al. 2010) indicates that inter-species hybridization has occurred. Eastern wolf mtDNA haplotypes are more common in this area, but the incidence of markers for *Canis lupus* is substantial, especially in western Minnesota. Y-chromosome (paternal) markers of the two lineages are more evenly represented.

Admixture between members of long-separate lineages introduces problems in applying a restrictive standard for reproductive isolation as a criterion for species limits. Templeton (1989) identified hybridization among species of *Canis* as an example justifying the need for his cohesion species concept as an alternative to reproductive isolation under the biological species concept to recognize species that interbreed but evolve as independent evolutionary lineages with distinct behaviors, ecologies, and morphologies. Others (e.g., Coyne and Orr 2004) adhere to reproductive isolation as a criterion, but advocate a detailed analysis of reproductive barriers without demanding absolute isolation.

The existence of such a broad hybrid zone, particularly in the western Great Lakes states, indicates that reproductive isolation is incomplete. Indications of backcrosses between the two wolves have been reported (Koblmüller et al. 2009a, Fain et al. 2010), and may indicate some degree of past or ongoing breakdown in reproductive isolation. At this time, this wolf population remains heterogeneous with respect to the contributions of the mtDNA-Y-chromosome haplotype combinations or microsatellite DNA of these two species. Members of the gray wolf and eastern wolf lineages have probably been in contact since gray wolves first appeared in North America in the late Irvingtonian Land Mammal Age about 500,000 years ago (Kurtén and Anderson 1980). Despite a long history of contact with gray wolf and near-extirpation, the

2141 distinctive genetic markers of the eastern wolf persist. Whether the rate of hybridization and  
2142 backcrossing is increasing or stable, which would indicate whether or the degree to which  
2143 reproductive isolation may be deteriorating, is unknown. Conspecific combinations of mtDNA  
2144 and Y-chromosome haplotypes are more common in male wolves of the western Great Lakes  
2145 region than expected by random mating (Wheeldon et al. 2010), which suggests some constraint  
2146 on admixture. Without this detailed information on the fitness and reproductive success of  
2147 hybrids, the population relationships of the two species and whether they are stable or tending  
2148 toward the complete merging of gene pools cannot be determined.

2149         The recently published analysis of SNP variation in *Canis* (vonHoldt et al. 2011) does not  
2150 alter our interpretation of gray wolf, eastern wolf, and their interaction in the Great Lakes region.  
2151 Difficulties with applying that study to questions concerning the taxonomic identity of Great  
2152 Lakes wolves were previously noted. The broad-brush approach of vanHoldt et al. (2011)  
2153 provides a valuable world-wide perspective on variation in *Canis*, but the taxonomic status of  
2154 wolves in the Great Lakes region requires a finer-scale analysis that explores the interactions  
2155 among individuals and packs on a more detailed geographic scale (Schwartz and Vucetich 2010).  
2156 SNP analysis can contribute to our understanding if applied at this scale and integrated with  
2157 information on mtDNA and Y-chromosome haplotypes of individual wolves. For example, the  
2158 SNP composition of males with both eastern wolf mtDNA and Y-chromosome haplotypes can be  
2159 compared with males with both gray wolf mtDNA and Y-chromosome haplotypes. SNP analysis  
2160 can be applied at a local scale to reveal a detailed understanding of important features of  
2161 interspecific hybridization (Fitzpatrick et al. 2009).

2162         Both natural and human-caused habitat changes have been implicated in other cases of  
2163 inter-specific hybridization (Mayr 1963, p. 128; Seehausen et al. 1997). Habitat degradation

together with human transport of individuals of one species into the range of another has resulted in breakdown in species integrity of the American black duck (*Anas rubripes*) through introgression from mallards (*Anas platyrhynchos*) (Mank et al. 2004), and the taxonomic integrity of some populations of California tiger salamanders (*Ambystoma californiense*) has been compromised by introgression from introduced barred tiger salamanders (*Ambystoma tigrinum*) (Fitzpatrick et al. 2009). The process by which previously isolated and divergent species experience introgressive hybridization has recently been called “reverse speciation” and identified as a concern in conservation (Seehausen 2006, Hendry 2009). Grant and Grant (2006) have termed as “despeciation” the process by which species of Galápagos Islands ground finches have lost morphological diagnosability through introgressive hybridization. They also suggest that, as environmental conditions change, reproductive isolation may be strengthened and diagnosability restored in a process they call merge-and-diverge dynamics.

Because the essential features of the hybridization process for *C. lycaon* and *C. lupus* are unknown, it is unknown whether reverse speciation or despeciation is occurring. If populations expand and ecological conditions improve, there could even be a restoration or strengthening of isolating mechanisms. If isolating mechanisms deteriorate, it raises the question of at what point the process of despeciation is considered complete and only one species should be recognized. A precedent for formal taxonomic merging as a result of hybridization is the inclusion of the Mexican duck (*Anas diazi*) with mallards (*Anas platyrhynchos*) by the American Ornithologists’ Union (1983), although that action has been questioned by the authors of a later mtDNA study of the mallard complex (McCracken et al. 2001). The possibility of merge-and-diverge dynamics (Grant and Grant 2006) suggests that a taxon should be recognized as long as individuals or populations representing its genetic distinctness remain. The long period of persistence of



distinctive eastern wolf characteristics despite long contact with both *C. lupus* and coyotes further argues for caution.

In comments on the study of Koblmüller et al. (2009a), Cronin and Mech (2009) state that taxonomy is subjective at and below the species level and propose the alternative of simply referring to the Great Lakes wolves as a population of mixed ancestry. “Mixed ancestry” encompasses diverse situations ranging from mild introgression to completely merged and homogenized populations, so that descriptor is too imprecise to characterize eastern wolves with the currently available information. We agree with Koblmüller et al. (2009b) that description of hybridization and introgression in the wolf population in the Great Lakes region does not preclude the consideration and recognition of either taxa or ecotypes, and that important information can be lost if taxonomic and ecological contexts are not considered.

#### **Species limits of *Canis lupus* with respect to the red wolf**

Goldman’s (1937) recognition of the red wolf as a distinct species (*Canis rufus*) has been followed by most taxonomic authorities, but the account for the red wolf in Wilson and Reeder (2005) accepts the conclusion of Wayne and Jenks (1991) that the red wolf is a hybrid and suggests that it should be considered of uncertain taxonomic status. As a compromise, it identifies the red wolf as a subspecies of *C. lupus*. The taxonomic status of the red wolf with respect to *C. lupus* will be evaluated in this section; its taxonomic relationship to the eastern wolf will be considered in the following section.

On the basis of several discriminant function analyses, Nowak (1979) found the red wolf to be intermediate between coyotes and gray wolves. His series of skulls from before 1930 show no overlap with coyotes, except in central Texas. Series taken after 1930 show increasing

amounts of overlap with coyotes as hybridization progressed, which he attributed to reduced densities of red wolves as a result of human control efforts.

Red wolves from the captive population share all microsatellite DNA alleles with coyotes, and 8% of their alleles are not found in gray wolves (Roy et al. 1994). Red wolves are closer to coyotes than to gray wolves in multidimensional scaling analysis (Roy et al. 1994, 1996, Figure 7 of this paper) and in neighbor-joining trees (Roy et al. 1994, 1996, Wilson et al. 2000) based on genetic distances. Eleven additional historical red wolf samples (pre-1940) extended these results (Roy et al. 1996). Unique alleles were found in red wolves, but they were few compared to populations of coyotes and gray wolves.

Two subsequent analyses of red wolf microsatellite DNA data from Roy et al. (1994, 1996) in addition to expanded coyote and gray wolf samples came to different conclusions. Reich et al. (1999) accept the premise that the red wolf originated from hybridization between coyotes and gray wolves and estimate that the event occurred as much as 12,800 years ago, but probably within the last 2500 years. Bertorelle and Excoffier (1998) found these data compatible with a model of the red wolf and coyote as sister species that diverged much more recently than their separation from the gray wolf lineage. Accordingly, red wolves are also closer to coyotes than gray wolves at the major histocompatibility complex (MHC) gene *DRB1* (Hedrick et al. 2002).

SNP data (vonHoldt et al. 2011) also place red wolf near coyotes and separate from wolves in the principal components analysis and in the STRUCTURE analysis at  $K = 3$ , which identifies three groups: wolves, dogs, and coyote combined with red wolf.  $F_{ST}$  ranged 0.08 to 0.1 between red wolf and coyote and 0.12 to 0.18 between red wolf and western and Mexican gray wolves. SABER analysis assigned about 75 to 80 % of the red wolf genome to coyotes.

The early restriction site data from mtDNA (Wayne and Jenks 1991) are difficult to interpret because samples were from canids captured while hybridization between red wolves and coyotes was underway, and individuals were classified on morphological criteria as red wolves, coyotes, or hybrids. The founders of the captive population were found to have the same haplotype as two coyotes from Louisiana. Mitochondrial cytochrome *b* gene haplotypes from six pre-1930 skins identified as from red wolves and reference samples of gray wolves and coyotes were distributed among two clades, corresponding to a wolf clade and a coyote clade (Wayne and Jenks 1991). The six historic red wolves were distributed evenly among the two clades. The three historic specimens in the coyote clade were not identical to any coyote, and one of three historic red wolves in the wolf clade had the same haplotype as a gray wolf. Eleven additional historic samples (pre-1940) extended these results (Roy et al. 1996). Eight historical samples were placed in the coyote clade in a parsimony tree, but none were identical to coyotes. One of three individuals in the gray wolf clade was identical to a gray wolf haplotype.

The mtDNA control region sequence of the red wolf captive population is unique and has an average divergence from coyotes of 3.24%, compared to an average of 2.79% divergence among coyote haplotypes (Adams et al. 2003). The red wolf is in the coyote clade of the neighbor-joining tree of haplotypes in this study.

Hailer and Leonard's (2008) study of mtDNA control region and Y-chromosome haplotypes explored potential reproductive isolation among the Mexican wolf (*C. l. baileyi*), red wolf, and coyotes, which were historically sympatric in central Texas. The single control region haplotype found in the captive population of red wolves was in the coyote clade (two base pairs different from the nearest coyote), which was strongly divergent from the wolf clade, including Mexican wolf. Of the five red wolves whose Y-chromosome haplotypes were identified, four

shared a haplotype with coyotes, while one was identical to domestic dog (Bannasch et al. 2005). Inter-species hybridization has had an influence on these three species, but the greatest effect was on the red wolf, especially on its Y-chromosome ancestry (Hailer and Leonard 2008).

Several aspects of the available data hinder clear inferences about taxonomic comparisons of the red wolf and gray wolves. The data are derived from a relatively small number of historical specimens, and from captive populations of red wolf and Mexican wolves that were derived from very small numbers of founders that are not likely to be representative of historical populations. Founder effects and genetic drift have likely affected the genetic composition of the captive populations. Further, it is difficult to separate the results of rare, and old incidences of hybridization from the undoubted introgression from coyotes that was occurring at the time the last wild red wolves were rescued (Nowak 1979).

Even with these limitations, genetic information places most individuals of the red wolf closer to coyotes than to gray wolves. This is clear in the nuclear microsatellite DNA data despite the sharing of many alleles among species and relatively few private alleles in red wolves. Some mtDNA cytochrome *b* gene haplotypes from historical red wolf samples are wolf-like, but the red wolf haplotype from the higher-resolution mtDNA control region of red wolves in the recovery program is unique and in the coyote clade. The Y-chromosome haplotypes from red wolves in the recovery program indicates introgression from coyotes (haplotype H15) and from domestic dog (haplotype H1, Table 5). The morphometric analyses of chronological series of *Canis* by Nowak (1979) documents the historical existence of a morphologically identifiable red wolf, followed by the decline in its morphological distinctiveness as hybridization with coyotes progressed. There is very limited information from historical specimens (Wayne and Jenks 1991, Roy et al. 1996), so we do not know the genetic composition of that historical red

wolf, but most red wolf individuals for which we have genetic information are closer to coyotes rather than to gray wolves. The closer relationship of the red wolf to coyotes than to gray wolves indicates it is outside of the gray wolf lineage and is not within the species limits of *Canis lupus*. It does raise the question of whether the red wolf is within the species limits of any members of the greater coyote clade, including the eastern wolf, and this will be evaluated in the next section.

### **Species limits within the coyote clade**

The mtDNA haplotypes of red wolf and some eastern wolves are part of a greater mtDNA clade that includes coyote haplotypes. This section evaluates whether red wolf and eastern wolf are within the species limits of coyotes.

*Nuclear genetic markers.* Red wolf (both from captive population and historical specimens) and eastern wolf are outside the 95% confidence ellipse encompassing coyotes based on multi-dimensional scaling analysis of 10 microsatellite loci (Roy et al. 1996, Figure 7 of this paper). A southern Quebec sample, which likely has some representation of eastern wolf, was even more distant from coyotes towards gray wolves. This pattern of divergence of red wolf and eastern wolf from coyotes is evident in a neighbor-joining tree based on genetic distances ( $D$  of Nei 1978) from the same data. The distances between eastern wolves and coyotes (0.216 and 0.341 for historical and captive red wolves, respectively) were greater than the average distances among coyote samples (0.188) (Roy et al. 1996).

Analysis of microsatellite DNA data from coyote, eastern wolf, and red wolf using neighbor-joining trees, log-likelihood analysis, and probability-of-identity analysis by Wilson et al. (2000) distinguish these groups from gray wolves, and also distinguish eastern wolf and red

wolf from coyotes. Additional information comparing red wolves and coyotes is provided by gene sequences in the major histocompatibility locus (MHC) (Hedrick et al. 2002). Three of four red wolf alleles were also found in coyotes from Texas and North Carolina. The fourth allele was a single nucleotide different from an allele found in coyotes.

STRUCTURE analyses have consistently identified the eastern wolf as a discrete group. The analysis by Wilson et al. (2009) separated Algonquin eastern wolf from the nearby population of “Tweed wolves” of the Frontenac Axis, which are primarily coyote, and from Texas coyotes as well. STRUCTURE and GENELAND analyses by Rutledge et al. (2010b) indicate that despite interbreeding, the Algonquin wolves remain genetically distinct from the Frontenac Axis population. Rutledge et al. (2010b) also reported a tendency of conspecific mating at Algonquin Provincial Park, which can contribute to the reinforcement of reproductive isolation. Other STRUCTURE analyses identified an eastern wolf (or Great Lakes wolf) cluster in comparisons with western coyotes and eastern coyotes (Koblmüller et al. 2009a), and western Great Lakes states clusters in analyses with sympatric coyotes (Fain et al. 2010, Wheeldon et al. 2010).

Comparison of genetic distance measures reported in these studies further support genetic distinctiveness of the eastern wolf.  $F_{ST}$  values between eastern wolves from Algonquin Provincial Park and coyotes are greater than between Algonquin and nearby hybridized eastern wolf-gray wolf populations (Grewal et al. 2004), and  $F_{ST}$  is high between western Great Lakes states wolves and sympatric coyotes. Genetic distance (as measured by  $\theta$ ) is greater between eastern wolf (Great Lakes wolf) and coyotes than between eastern wolf and gray wolves (Koblmüller et al. 2009a, Table 6 of this paper). A factorial correspondence analysis of these

data indicates that eastern and western coyotes are much closer to one another than either is to the great majority of eastern wolves (Figure 3(d) of Koblmüller et al., 2009a).

These data on nuclear genetic markers consistently portray a pattern indicating coyote, eastern wolf, and red wolf are related, but with the red wolf and eastern wolf as outliers to coyote.

Analyses of SNP data (vonHoldt et al. 2011) generally indicate that red wolf and eastern wolf are both more divergent from coyotes than coyote populations are from one another. The principal components analysis places the red wolf as a discrete cluster near coyotes, although the genetic bottleneck associated with the founding of the red wolf captive population has very likely contributed to this discreteness. Eastern wolf is more divergent from coyote. The possible inclusion gray wolf individuals or introgression of gray wolf into the eastern wolf population may have contributed to this divergence; however, the two individuals from Algonquin Provincial Park, where mtDNA and Y-chromosome haplotypes indicate only minimal gray wolf influence and coyote introgression is evident (Rutledge et al. 2010b), are near other eastern wolves and divergent from coyote.

STRUCTURE analysis of the SNP data (vonHoldt et al. 2011) separates eastern wolf (together with western wolves) from coyote and dog at  $K = 3$ . Red wolf is not separated from coyote until  $K = 9$ .  $F_{ST}$  among the three groupings of the coyote sample (western, midwestern/southern, and northeastern) range 0.2 to 0.5. Both red wolf and eastern wolf are more divergent from coyotes:  $F_{ST}$  between red wolf and coyotes ranges 0.08 to 0.1, and between eastern wolf and coyotes ranges 0.08 to 0.11.

*Mitochondrial DNA and Y-chromosome haplotypes.* Mitochondrial cytochrome *b* gene sequences are ambiguous with respect to red wolf-coyote divergence, as discussed earlier, with three of 11 historical specimens distributed in the gray wolf clade (Wayne and Jenks 1991, Roy et al. 1996). The three red wolf haplotypes in the coyote cytochrome *b* clade are distributed on different branches. The single control region haplotype from the captive breeding program for the red wolf appears within the coyote clade in neighbor-joining trees from relatively basal (Vila et al 1999, Figure 1; Wilson et al. 2000, Figure 5; Figures 10 and 11 of this paper; Wilson et al. 2003; Hailer and Leonard 2008) to embedded among various coyote clades (Adams et al 2003, Figure 2; Fain et al. 2010, Figure 3). The basal positions are generally not well supported.

In their study designed specifically to examine coyote and red wolf in a former area of sympatry, Hailer and Leonard (2008) found that the single red wolf control region haplotype from the captive population was not shared with any coyote, but the observed high haplotype diversity of coyotes suggests that additional coyote haplotypes may remain unsampled. It is significant that red wolf and coyote in a former area of sympatry and with documented modern hybridization do not share mtDNA haplotypes. As described earlier, the two Y-chromosome haplotypes found in red wolves in the recovery program, reflect coyote and dog introgression (Table 5 of this paper).

The early restriction site analysis of mtDNA of wolves from the Great Lakes region found haplotypes distributed among wolf and coyote clades (Lehman et al. 1991, Figure 3). Minimum spanning and neighbor-joining trees of mtDNA control region sequences group unique eastern wolf haplotypes C1 and C3 together basal to coyotes, but another eastern wolf haplotype, C13, while not identical to any coyote haplotype, is embedded with coyote haplotypes (Wilson et al. 2000, Figure 5; Figures 10 and 11 of this paper; Fain et al. 2010).



Additional mtDNA control region sequences from historical wolf specimens from the Great Lakes area confirm this pattern (Leonard and Wayne 2008). Koblmüller et al. (2009a) report sharing of haplotypes between coyotes and eastern wolves, but information is not provided on the geographical source of these individuals.

Analysis of mtDNA and Y-chromosome haplotypes from *Canis* of the western Great Lakes states (Fain et al. 2010, Wheeldon et al. 2010) indicates no sharing of haplotypes between eastern wolves and coyotes, although phylogenetic analysis indicates that there may have been past rare incidences of hybridization resulting in a low level of wolf influence on coyotes. Koblmüller et al. (2009a) reported that sharing of Y-chromosome haplotypes by eastern wolves and coyotes is rare.

Hybridization between eastern wolves and coyotes in the eastern Great Lakes regions has primarily affected eastern coyotes, including those of the northeastern United States (Kays et al. 2009), and the “Tweed wolf” of southern Ontario, which Wilson et al. (2009) consider a population of eastern coyote. In a recent study (Rutledge 2010b) coyote mtDNA was found in wolves in Algonquin Provincial Park, but shared coyote-eastern wolf Y-chromosome haplotypes were less common. Eastern wolf Y-chromosome haplotypes were more frequent in breeding males than predicted by random mating, so assortative mating may be playing a role in maintaining a cohesive eastern wolf population despite past interbreeding with gray wolves and coyotes.

The magnitude of divergence between red wolf and eastern wolf clades and other mtDNA clades within the greater coyote clade can be evaluated by comparison to divergence among clades strictly attributed to coyotes. Red wolf-coyote and eastern wolf-coyote divergences are generally greater than coyote-coyote divergences (Table 9). Mitochondrial DNA

cytochrome *b* sequences within the coyote clade had an average sequence divergence from coyotes of 1.2%, compared to an average of 1.1% among coyote haplotypes (Wayne and Jenks 1991, Roy et al. 1996, Table 9 of this paper). For mtDNA control region haplotypes, the average divergence between C2 of red wolf and coyotes is 3.2%, compared to an average of 2.8% among coyote haplotypes.

*Conclusions.* The eastern wolf forms a divergent clade basal to the greater coyote clade for both mtDNA and Y-chromosome haplotypes (Wilson et al., 2000, Fain et al. 2010), with additional clades consistent with subsequent, ancient, introgression from coyotes. Generally, sequence divergences and branching patterns of the divergent clades indicate a deeper and older divergence between eastern wolf and coyote than among coyote branches of the greater coyote clade. This agrees with various nuclear microsatellite DNA studies including gray wolves and coyotes of the Great Lakes region that show eastern wolf as an identifiable cluster (Koblmüller et al. 2009a, Fain et al. 2010, Rutledge 2010b, Wheeldon et al. 2010) and an outlier to coyotes (Roy et al. 1996). Eastern wolves and coyotes do not interbreed where they are both sympatric and abundant in the western Great Lakes states (Fain et al. 2010, Wheeldon et al., 2010). In the eastern portion of their range, eastern wolves have experienced some coyote introgression through the maternal line as indicated by mtDNA, but appear to be maintaining a cohesive population in Algonquin Provincial Park assisted by a preference for intraspecific mating (Rutledge et al. 2010b). Introgression from coyotes through the maternal line likely occurred historically when the eastern wolf population was at lower densities and the availability of conspecific mates was reduced. Reproductive isolation may not be complete in this eastern portion of the eastern wolf range, but the eastern wolf is displaying cohesion and maintaining

itself as a phylogenetically unique lineage in the Great Lakes region distinct from sympatric coyotes.

Red wolf mtDNA also forms a divergent clade within the greater coyote clade. This is consistent with microsatellite DNA data that indicate it as an outlier to coyotes. Its Y-chromosome haplotypes do not form a distinct clade, the H15 haplotype is identical to a Texas coyote haplotype, whereas the H1 haplotype is identical to domestic dog haplotypes (Bannasch et al. 2005). As a result, the Y-chromosome genetics of red wolves is not useful for phylogenetic inference. A morphologically discrete and identifiable red wolf has survived reduction in its population size and range and subsequent hybridization with coyotes and either dogs or wolf-dog hybrids (Nowak 1979). The modern descendants of that original red wolf population show the influence of hybridization as the population status of the red wolf declined. The red wolf remains an identifiable lineage separate from coyotes, despite past coyote hybridization and the continuing threat from coyotes in areas where the red wolf has been re-introduced (Adams et al. 2003).

Red wolf and eastern wolf remain identifiable lineages that have evolved in North America with coyotes. Despite long histories of sympatry with or proximity to coyote populations and the ability of *Canis* species to interbreed, instances of ancient hybridization between these two lineages and coyotes have been rare. Major threats of introgression can be associated with recent reduction of wolf population densities, and alteration of natural habitat that once contributed to the isolation of the different lineages.

#### **Relationship of red wolf and eastern wolf**

In the previous section, red wolf and eastern wolf were concluded to be lineages divergent from coyotes. This section considers whether red wolf and eastern wolf are close enough to be considered a single species under the older name of *Canis lycaon* as proposed by Wilson et al. (2000) based on their genetic similarities.

Two very different hypotheses involving hybridization have been proposed to explain the general similarities between red wolf and eastern wolf. Nowak's (1979, 1995, 2002, 2003) morphometric analyses describe eastern wolf, which he considers to be a subspecies of gray wolf, as statistically intermediate between gray wolf and red wolf, with red wolf resembling eastern wolf more than it does any gray wolf. Nowak (2002) proposed that the eastern wolf may have originated as a result of hybridization between red wolf and gray wolf. This hypothesis is not supported by the available genetic data. Samples of wolves from Algonquin Provincial Park and surrounding area reported by Wilson et al. (2000, Figure 5A) have either the C1 haplotype unique to eastern wolf, or either coyote or coyote-like haplotypes, but no haplotypes associated with gray wolf to the north and west. A sample of three individuals from "north of Algonquin Park" (Wilson et al. 2000) included both the eastern wolf C1 haplotype and gray wolf haplotype lu32, which indicates interaction between the species further to the north. Subsequent studies have reported a low incidence of gray wolf mtDNA and Y-chromosome haplotypes in Algonquin Provincial Park (Grewal et al. 2004, Rutledge et al. 2010b). The park is within the range of eastern wolf as recognized by Nowak (2003), and therefore should include a substantially greater percentage of gray wolf genetic markers if hybridization involving that species was a major factor in the origin of the eastern wolf.

Roy et al. (1994, 1996) attributed the genetic similarity between red wolf and eastern wolf to both having hybridized with coyotes. The mtDNA haplotypes initially identified as

having been derived from coyotes (Lehman et al. 1991) are different in red wolf and eastern wolf and have not been found in western coyotes, the presumed source of hybridization (Roy et al. 1996; Wilson et al. 2000, 2003; Hailer and Leonard 2008). A mtDNA haplotype is shared by eastern wolf and coyotes of the northeastern United States, but it is interpreted as an eastern wolf haplotype that has influenced this eastern coyote population and not the reverse (Kays et al. 2009). As discussed in a previous section, the similarity to coyote mtDNA haplotypes is more likely the result of a shared evolutionary history with coyotes separate from gray wolves, with some rare instances of ancient introgression.

The proposal of Wilson et al. (2000) that eastern wolf and red wolf may be the same species is based on similarities in nuclear microsatellite DNA loci and mtDNA haplotypes. Both sets of genetic markers indicate that there is genetic divergence between eastern wolf and red wolf. This divergence could represent coyote or dog introgression in red wolf, geographic variation within a single species, or it may indicate genetic discontinuity between two species. Interpretation of the taxonomic significance of this difference is confounded by the large geographic distance between the sources of samples: the red wolf samples are all from descendants of wolves captured in eastern Texas and southwestern Louisiana, while eastern wolf samples were collected at locations over 2000 km away in eastern Canada. There are no genetic samples from intervening areas, except for the historical samples from Maine and New York, which have eastern wolf mtDNA haplotypes (Wilson et al 2003). Nowak (2002) identifies a single historical skull from Maine as red wolf based on its morphological features. If these wolves are considered as belonging to two species, this gap in sampling would make it impossible to infer from existing data the past geographic ranges of the two putative species and any possible interactions between them.

Red wolf and eastern wolf are more similar to each other at autosomal microsatellite loci than either is to Texas coyotes or western gray wolves (Roy et al. 1994, 1996; Wilson et al. 2000). Microsatellite data have very limited value for this comparison because of the large geographic distances between the sources of the samples and the loss of linearity with distance of common distance measures for microsatellite DNA data (Paetkau et al. 1997). In addition, distances are based largely on allele frequency differences, which have undoubtedly been affected by severe population bottlenecks experienced as the red wolf population declined to near extinction before the founding of the captive red wolf population.

Analyses of SNP data (vonHoldt et al. 2011) generally indicate divergence between red wolf and eastern wolf. As indicated in the previous section, principal components analysis places red wolf near coyotes and eastern wolf near gray wolves. STRUCTURE analyses similarly place red wolf with coyotes and eastern wolf with gray wolves at  $K = 3$ . SABER analysis attributes more gray wolf ancestry to eastern wolf and more coyote ancestry to red wolf.  $F_{ST}$  values (vonHoldt et al. 2011, Supplemental Table S3) also indicate divergence between eastern wolf and red wolf.  $F_{ST}$  between red wolf and eastern wolf is reported as 0.11, which is comparable the divergence of each from coyotes. For comparison,  $F_{ST}$  values among coyotes in this study ranged 0.2 to 0.5. Limiting the interpretation of the SNP data are the small sample size of the eastern wolf sample ( $N = 19$ ), and the differential influences of admixture on eastern wolf and red wolf: possible inclusion of or introgression from gray wolves in the western Great Lakes region, admixture of coyotes and eastern wolves in the eastern Great Lakes region, and admixture between coyote and red wolf.

Mitochondrial DNA data could lend support to the proposal of Wilson et al. (2000) that eastern wolf and red wolf are one species if they were found to cluster together within the greater

coyote clade. In the minimum spanning network of Wilson et al. (2000, Figure 5A) the haplotype of red wolf is closer to coyotes, but appears in a basal position on the branch leading to eastern wolf. In neighbor joining and maximum parsimony trees in analyses that included historical specimens (Wilson et al. 2003), red wolf was not basal to eastern wolf, but both formed separate branches that were basal to coyote haplotypes. Red wolf and eastern wolf did not cluster together in either a parsimony consensus cladogram (Leonard and Wayne 2008, Figure 1(b)) or neighbor-joining tree (Fain et al. 2010, Figure 3) of control region mtDNA data, where they were separated by intervening coyote clades.

Red wolf and eastern wolf do not share comparable Y-chromosome haplotypes (Hailer and Leonard 2008, Fain et al. 2010, Rutledge et al. 2010b), but as discussed earlier, four of five red wolves shared a haplotype found in Texas coyotes and is likely the result of coyote introgression (Hailer and Leonard 2008) and the “gray wolf-like” haplotype reported by Hailer and Leonard (2008) is consistent with dog origin (Bannasch et al. 2005). Y-chromosome composition of the captive red wolf population is likely the result of wolf-dog hybrid and coyote introgression and is therefore not comparable to eastern wolf haplotypes.

The available information on haplotype composition and relationships indicate that eastern wolf and red wolf do not share mtDNA haplotypes and do not together form a single group exclusive of coyote lineages. Independent evolution of these two taxa from different lineages of coyote-clade ancestors is more consistent with the available genetic data and argues against combining them as a single species but retaining them as *Canis lycaon* and *Canis rufus*.

*Nomenclatural issues.* Regardless of present lack of genetic support for combining the taxa, some nomenclatural problems would need to be addressed before uniting *C. lycaon* and *C. rufus*.

A compromise approach would be to recognize *C. lycaon* and *C. rufus* as subspecies within the same species. This would require the formal publication of new name combinations for at least one subspecies. In addition, there is significant geographic variation in genetic composition within a more broadly defined taxon that includes both *C. lycaon* and *C. rufus*. Moreover, Texas wolves occupied a very different environment than wolves in eastern Canada.

The appropriate name for a single species that would encompass both *C. lycaon* and *C. rufus* remains an issue. Wilson et al. (2000) suggested that the name be *C. lycaon*. The rule of chronological priority normally applies in such cases, and the name *Canis lycaon* was published earlier than *Canis lupus* var. *rufus* (Table 1). Two reviewers of an earlier draft of this paper questioned the applicability of the older name in this case owing to uncertainty about the identity of the type specimen of *Canis lycaon* because it may have been a hybrid. As explained by Goldman (1944), the type is the individual portrayed in Schreber's illustration, which was in turn based on a figure published by Buffon in 1761. Article 73.1.4 of the International Code for Zoological Nomenclature (1999) provides for such instances: "Designation of an illustration of a single specimen as a holotype is to be treated as designation of the specimen illustrated; the fact that the specimen no longer exists or cannot be traced does not of itself invalidate the designation." Because the disposition of the remains of the illustrated specimen is unknown, and the holotype must be of the same group to which the species name is applied, the identity of the specimen portrayed is important.

The type locality was restricted by Goldman (1937) to the vicinity of Quebec, Quebec. Wolves in this region may have already been interbreeding with coyotes or dogs at the time the holotype was collected. Support for earlier hybridization is provided by the presence of either coyote or dog mtDNA in wolf (based on morphology) remains of four individuals from a 400-



500-year-old archaeological site in southern Ontario (Rutledge et al. 2010a). Article 23.8 of the International Code specifies that, “a species-group name established for an animal later found to be a hybrid must not be used as the valid name of either of the parental species, even if it is older than all other available names for them.” Pocock’s (1935) argument that the type may have been a hybrid was rejected by Goldman (1937), partially on the basis that a hybrid could still serve as the type, which is in conflict with today’s Code.

The next oldest name in Goldman’s (1944) synonymy for *C. lycaon* is *Canis lupus canadensis* (Table 1). Allen and Barbour (1937) note that the type specimen for *C. l. canadensis* is a skull illustrated by Plainville and that the locality was given only as Canada, so questions may also be raised about its identity and relation to modern wolf populations in eastern Canada. In this case the holotype of *C. rufus* (type locality: Austin, Texas) might be more appropriate because hybridization with coyotes would have been unlikely at the time it was described. Nowak (2009) believes that the holotype of *C. lycaon* is actually a specimen of *C. rufus* based in its description as black, which he describes as a well-known coat color in *C. rufus*, but unusual for *C. lycaon*. Black coat color could also indicate that the individual had dog ancestry. Current wolves nearest the *C. lycaon* type locality (vicinity of Quebec), however, have the mtDNA haplotypes of *C. lycaon*. Even if additional evidence should provide support for formally combining these taxa, this issue would need to be resolved before formal changes in taxonomy are made.

### **The subspecies of *Canis lupus***

The following analysis and discussion is organized by the remaining (less *C. l. lycaon*) subspecies of *C. lupus* recognized by Nowak (1995). This does not mean that Nowak’s

classification is accepted without consideration of alternative classifications. The analysis therefore includes consideration of formerly-recognized subspecies (e. g., Goldman [1944], Hall [1981]) that were reduced to synonymy by Nowak (1995) where patterns of variation within these four subspecies suggest that some finer-scale taxonomic subdivision might be recognizable.

*Canis lupus baileyi* (Mexican wolf). Both morphometric and genetic evidence support the distinctiveness of *C. l. baileyi* and its recognition as a subspecies. Genetic analysis of living specimens is limited to the descendents of the founders of the captive breeding population, thought to be seven individuals (Hedrick et al. 1997). Although the effects of genetic drift and a small founder population has likely increased the observed divergence of living *C. l. baileyi* from other wolves at autosomal microsatellite DNA loci (García-Moreno et al. 1996), they cannot account for the unique mtDNA haplotype (Roy et al. 1996, Vilà et al. 1999) and several private microsatellite DNA alleles (García-Moreno et al. 1996) found in *C. l. baileyi*. Additional genetic data from historical, museum specimens (Leonard et al. 2005) have corroborated the results obtained from living individuals, and further indicate that the “southern mtDNA clade” of the Mexican wolf is divergent from other North American wolves.

Comparisons of mtDNA sequence divergences among *C. lupus* haplotypes support recognition of *C. l. baileyi* as a subspecies rather than as a species distinct from other *C. lupus*. The predominant *C. l. baileyi* haplotype has a sequence divergence of 2.2% from the closest other North American *C. lupus* haplotype (Wayne and Vilà 2003, p. 228), compared to sequence divergences averaging 2.9% within *C. lupus* (Vilà et al. 1999, p. 2093), 8% between *C. lupus* and

either *C. lycaon* or *C. rufus*, and 10% between gray wolf and coyote (Wilson et al. 2000, p. 2159).

Analyses of SNP data (vonHoldt et al. 2011) from 10 Mexican wolves from the captive breeding program also indicate the distinctness of the Mexican wolf. Principal components analysis of the North American gray wolves (vonHoldt et al. 2011, Supplementary Figure S2) separates Mexican wolf from other gray wolves on the first principal components axis, which accounts for 6.6% of the variance. Mexican wolf is basal to other North American gray wolves in both a neighbor-joining tree and a phylogram (vonHoldt et al. 2011, Supplementary Figure S5).  $F_{ST}$  between Mexican wolf and other western gray wolves is 0.1, which is greater than  $F_{ST}$  values, which range 0.01 to 0.08 among western gray wolves from different regions. While these results are consistent with other genetic data, a founder effect in establishing the captive Mexican wolf population may have contributed to the measures of divergence based on both SNPs and microsatellite DNA.

There is consensus on the valid taxonomic standing of *C. l. baileyi*, but there is some controversy based on interpretation of morphometric data on the historical boundaries of the subspecies. Nowak (1995) recognized *C. l. baileyi* as a subspecies, but did not adopt Bogan and Mehlhop's (1983) inclusion of *C. l. mogollonensis* and *C. l. monstrabilis* as its synonyms. These different interpretations may be related to larger sample sizes used by Bogan and Mehlhop (1983), who studied 253 skulls of the three subspecies in question, compared to 88 skulls studied by Nowak (1995). It may also be related to Bogan and Mehlhop's (1983, p. 15) preference for principal components analysis as a more objective method for assessing overlap in characters than discriminant function analysis, which was used by Nowak (1979, p. 4). Bogan and Mehlhop (1983) also carried out discriminant function analyses on their data and found

intermediacy of skulls assigned to *C. l. mogollonensis* between *C. l. baileyi* and more northern wolves. The two different discriminant function analyses have generally comparable outcomes, so the difference is in interpreting which subspecies to assign a collection of individuals that is intermediate between recognized taxa. Bogan and Mehlhop (1983) and Nowak (1995) agree that the range of *C. l. mogollonensis* in Arizona was a transition zone where *C. l. baileyi* intergraded with more northern *C. lupus*, which is consistent with the limited available genetic data from historical specimens (Leonard et al. 2005). Wolves were long ago extirpated — perhaps by the 1940s (Parsons 1996) — within the ranges of *C. l. monstrabilis* and *C. l. mogollonensis* so the controversy is now primarily of historical interest.

The phylogenetically closer relationship of *C. l. baileyi* to certain Eurasian wolf populations than to other North American *C. lupus* (Vilà et al. 1999, Wayne and Vilà 2003) indicates that contact was secondary between *C. l. baileyi*, as delineated by Bogan and Mehlhop (1983), and the later-arriving, more northerly *C. l. nubilus*. Both morphometric (Nowak 1995, p. 385) and genetic data (Leonard et al. 2005) are consistent in indicating that, once *C. l. baileyi* came into contact with more recent *C. lupus* invaders from Eurasia, there was a broad area of reproductive interaction between them. This interaction could have been in the form of a relatively stable and broad zone of intergradation between them, or *C. l. nubilus* could have incorporated genetic elements of *C. l. baileyi* as it rapidly displaced the latter subspecies to the south. General models on plants and animals have demonstrated the process by which local genes are incorporated into an invading population (Currant et al. 2008). The interaction has been described more locally in Arizona by the morphometric data (Bogan and Mehlhop 1983, Nowak 1985), and more expansively by the mtDNA data from historical specimens, where a

northern haplotype associated with *C. l. nubilus* was found in Arizona and southern haplotypes associated with *C. l. baileyi* were found as far north as Nebraska.

*Canis lupus baileyi* and *Canis rufus* do not overlap in morphometric variation of skull features (Nowak 1979). The genetic data, particularly that of Hailer and Leonard (2008), indicate that if hybridization has occurred between these species it has apparently not affected the genetic composition of *C. l. baileyi*, with one exception. The Y-chromosome haplotype H29 that Hailer and Leonard (2008, Fig 3B) found in some *C. l. baileyi* and identified as a wolf haplotype is common in dogs (Table 5). The *C. l. baileyi* with H29 are all descended from an individual in the “Aragon lineage,” which has a nuclear DNA composition (based on microsatellites) that clusters with other *C. l. baileyi* (Hedrick et al. 1997).

*Canis lupus arctos* (Arctic wolf). The three high Arctic Islands sampling areas for *C. l. arctos* were grouped together in a neighbor-joining distance analysis based on microsatellite DNA data (Carmichael et al. 2008, Figure 3(B)), but the authors observed that the island populations exhibited only one private allele, and that their unpublished mtDNA data did not identify any unique Arctic Islands haplotypes. Based on the assumption that a long isolation in an Arctic refugium, as proposed by Nowak (1983), should have resulted in a higher proportion of unique alleles, Carmichael et al. (2008) concluded that these Arctic Islands populations are the result of recent colonization from the mainland. Their interpretation was further supported by low levels of genetic diversity in the island wolves. Low levels of diversity also affect the reliability of distance measures calculated from microsatellite loci (Paetkau et al. 1997). The genetic differences observed under these circumstances are not likely to be of taxonomic significance.

The morphometric data in support of recognition of *C. l. arctos* have limitations. The overlap with mainland subspecies (*C. l. nubilus* and *C. l. occidentalis*) is not minimal (Nowak 1995, Figure 9), and the large polygons representing the mainland subspecies are likely affected by the very large scale of geographic sampling of the mainland subspecies. A more relevant comparison for evaluating taxonomically significant discontinuity between island and mainland populations would be between island and adjacent, coastal-mainland populations. Coastal-mainland populations do not appear to be well-represented in either morphometric study (Nowak 1995, Mulders 1997).

The genetic data together with difficulties in interpreting the morphometric data do not provide clear support for subspecies recognition of *C. l. arctos*. This conclusion is tentative because it is based on lack of supportive data rather than definitive information that these populations are not taxonomically recognizable. The genetic data consist only of autosomal microsatellite DNA, and some preliminary mtDNA data that did not detect unique haplotypes in the island populations (Carmichael 2008, p. 885). Y-chromosome and additional mtDNA data could better resolve the relationship between island and mainland populations, and therefore the taxonomic standing of *C. l. arctos*.

*Canis lupus occidentalis* (northern timber wolf). When viewed in the context of Nowak's (1995) subspecies classification, Skeel and Carbyn's (1977) morphometric analysis indicates average differences between *C. l. occidentalis* and *C. l. nubilus* with some overlap.

Mitochondrial DNA haplotypes based on restriction site mapping show the same haplotypes from Alaska to Montana and Minnesota; these samples were mostly within the range of *C. l. occidentalis* (Wayne et al. 1992, 1995). An exception is a haplotype that was unique to

Manitoba, likely within the range of *C. l. nubilus*. These studies do not show strong differentiation between *C. l. occidentalis* and *C. l. nubilus*, but the samples were used for a general, world-wide review of wolf variation and are therefore small and widespread and not at a suitable scale for resolving potential subspecies differences. A similar pattern (in this case, a lack of geographic pattern) with respect to resolving these two subspecies was found in a world-wide review of variation in wolf mtDNA sequences (Vilà et al. 1999), but samples were similarly too small and widespread to be very informative about subspecies differences.

The microsatellite DNA study of Carmichael et al. (2008) from widespread localities in Canada lends support for distinguishing *C. l. occidentalis* from *C. l. nubilus*, with most sampling areas largely attributable to *C. l. occidentalis* (Qamanirjuaq, Saskatchewan, Bluenose West, Cape Bathurst, Manitoba, Alberta, Porcupine, Alaska, Yukon, British Columbia, and Mackenzie) occurring together on the neighbor-joining tree (Carmichael et al 2008, Figure 3(B)). These sampling areas were not designed to assess subspecies classification, so some straddle Nowak's (1995) boundaries between the two subspecies. The Bathurst and Qamanirjuaq sampling areas appear to also include individuals from Nowak's (1995) range for *C. l. nubilus*, and this may explain why these localities do not group closely with other *C. l. occidentalis*.

The SNP study of vonHoldt et al. (2011) includes samples from coastal areas of British Columbia, which is within the range of *C. l. nubilus* as mapped by Nowak (1995). It also includes samples within the general range of *C. l. occidentalis*: "boreal forest" (Alaska), "tundra/taiga" (inland Canada), and "Rocky Mountain" (Yellowstone). The western coastal sample is differentiated from other western gray wolves on some axes of the principal components analysis (vonHoldt et al. 2011, Supplementary Figure S3). In a neighbor-joining tree and a phylogram (vonHoldt et al. 2011, Supplementary Figure S5), western coastal and

northern Quebec follow Mexican wolf in representing branches basal to other western gray wolves.  $F_{ST}$  among samples representing *C. l. occidentalis* (Alaska, inland Canada, and Yellowstone) range 0.01 to 0.03, while  $F_{ST}$  between these samples and the *C. l. nubilus* sample from coastal British Columbia is greater (range 0.6 to 0.8). vonHoldt et al. (2011) attribute this difference between coastal and inland populations and other patterns of geographic variation within gray wolves to differences in habitat characteristics. Differences in habitat can, however, coincide with subspecies boundaries and can play a role maintaining taxonomic distinctions when ranges of subspecies are contiguous. Explanations based on habitat variation can therefore be consistent with subspecies ranges and boundaries

Wolves in the western range of *C. l. occidentalis* are genetically discontinuous with coastal wolves of British Columbia and southeast Alaska at autosomal microsatellite DNA loci and for mtDNA haplotypes (Weckworth et al. 2005, 2010; Muñoz-Fuentes et al. 2009). The coastal wolves in this area were considered to be *C. l. nubilus* by Nowak (1995, 2002) these genetic data are discussed in more detail in the following section on that subspecies.

The overview of mtDNA variation presented in Table 4 indicates that *C. l. occidentalis* (Interior: Alaska, Yukon, Northwest Territories, Alberta, and Saskatchewan) and historical *C. l. nubilus* share some haplotypes but most, especially for *C. l. occidentalis*, are not shared.

*Canis lupus mackenzii* is treated as a synonym of *C. l. occidentalis* by Nowak (1995). Its distribution is mapped by Goldman (1944) and Hall (1981) as the northern Northwest Territories extending to the Arctic Ocean. The genetic discontinuity at the Mackenzie River indicated by microsatellite DNA near the southwest boundary of the mapped range of *C. l. mackenzii* (Carmichael et al. 2001) raises the possibility of genetic support for recognizing it as a subspecies. Additional microsatellite DNA data covering the eastern portion of the range of *C. l.*



*mackenzii* (Carmichael et al. 2008), which includes the type locality (Bathurst Inlet), does not support subspecies standing because the sampling areas (Cape Bathurst, Bluenose West, and Bathurst) that represent the range of *C. l. mackenzii* do not group together in a neighbor-joining analysis (Carmichael et al. 2008, Figure 3(B)).

The autosomal microsatellite DNA (Carmichael et al. 2007, 2008) and mtDNA data (summarized in Table 4) indicate limited genetic continuity between *C. l. nubilus* and *C. l. occidentalis*. Although there are the exceptions noted above related to sampling areas not being confined to single subspecies, these data are in general agreement with the morphometric support (Skeel and Carbyn 1977, Nowak 1995) for recognizing *C. l. occidentalis*.

*Canis lupus nubilus* (plains wolf). This is the most difficult and complex subspecies to evaluate because it is or was historically in contact with each of the other three *C. lupus* subspecies, *C. lycaon*, and probably *C. rufus*. Some areas included within *C. l. nubilus* range may represent intergrade or contact zones between subspecies. More localized genetic structure generated by habitat and prey preferences (Carmichael et al. 2001, 2007, 2008; Pilot et al. 2006; Musiani et al. 2007) may alternatively obscure or coincide with older patterns of structure that may be of more taxonomic significance. Moreover, this subspecies has suffered extirpation over a great part of its range, including all of the conterminous United States except for northeastern Minnesota and Isle Royale, so genetic data have been limited by the relatively few individuals from museum collections that have been characterized.

The kind of unique and divergent mtDNA haplotype sequences that have been so important in our understanding of *C. lycaon* and *C. l. baileyi* are not as evident in *C. l. nubilus*. The previously noted limitations of the size and scale of sampling of mtDNA for evaluating

subspecies taxonomy are particularly applicable for evaluating *C. l. nubilus*. The most widely-distributed haplotypes, designated lu-28 and lu-32 by Vilà et al. (1999) are widely distributed in both *C. l. nubilus* and *C. l. occidentalis*. Haplotypes from museum specimens indicate that both haplotypes were widely distributed in the western United States prior to extirpation of wolves from those areas (Leonard et al. 2005). Interestingly, lu-32 and a similar haplotype were also found in historical samples from Labrador (Leonard et al. 2005), which supports the similarity of wolves within the wide geographic range attributed by Nowak (1995) to *C. l. nubilus*. It does not, however, appear to be a genetic marker unique to *C. l. nubilus*, as it occurs within the range of *C. l. occidentalis* in Alberta and Northwest Territories. Most haplotypes from historical *C. l. nubilus* have not been found in *C. l. occidentalis* (Table 4, where *C. l. occidentalis* is represented by “Interior” samples from Alaska, Yukon, Northwest Territories, British Columbia, Saskatchewan, and most of Manitoba).

The neighbor-joining analysis of microsatellite DNA data (Carmichael et al. 2008, Figure 3(A)) groups together some northern sampling areas for *C. l. nubilus*, which include Baffin Island, the adjacent mainland, and Atlantic (Newfoundland). The Bathurst sampling area is also included in this group, but as discussed under *C. l. occidentalis*, it straddles Nowak’s (1995) boundary dividing the two subspecies. Unfortunately for taxonomic purposes, this study was designed to explore relationships of Arctic wolves and did not include samples from Ontario or Quebec in the southern Canada range of *C. l. nubilus*.

Nowak (1995) included wolves from the Pacific Northwest of the United States, coastal British Columbia, and southeastern Alaska in *C. l. nubilus*. An affinity between wolves in western coastal areas of Canada and those in southeastern Alaska was shown by morphometric data (Jolicoeur 1959). These coastal populations are closer in morphology to one another than

nearby inland populations (Jolicoeur 1959, Nowak 1983). Nowak (1995) observed that the southeastern Alaska populations are intermediate between *C. l. nubilus* and *C. l. occidentalis*.

Differentiation between coastal and inland wolves in southeastern Alaska has been reported for both microsatellite DNA (Weckworth et al. 2005) and mtDNA data (Weckworth et al 2010). Differentiation between coastal and inland wolves in British Columbia has also been documented using mtDNA (Muñoz-Fuentes et al. 2009). These coastal-inland patterns of divergence lend support to Nowak's (1995) boundary between *C. l. nubilus* and *C. l. occidentalis* in this region. The neighbor-joining tree of Carmichael et al. (2008) places southeast Alaska wolves with populations attributable to *C. l. occidentalis* rather than the distant, eastern *C. l. nubilus* represented in the study. Microsatellite DNA data may not be reliable, however, in situations where there is this type of geographic discontinuity among populations (Paetkau et al. 1997).

Coastal populations of southeast Alaska (Weckworth et al. 2010) and British Columbia (Muñoz-Fuentes et al. 2009) form a distinct phylogroup based on mtDNA, and share the common lu38 and unique lu68 haplotypes (Table 4). This sharing of haplotypes argues against taxonomically distinguishing the southeast Alaska wolves (as *C. l. ligoni*) from the coastal populations of British Columbia.

Although Nowak (1995) assigned the coastal populations of southeast Alaska and British Columbia to *C. l. nubilus*, genetic comparison with inland *C. l. nubilus* populations of the western United States is limited to the relatively small sample of historical individuals for which genetic data (mtDNA) are available (Leonard et al. 2005). The finding of Muñoz-Fuentes (2009) that coastal British Columbia wolves are less differentiated from inland *C. l. occidentalis* ( $\Phi_{ST} = 0.305$ ) than from the historical samples (Leonard et al. 2005) of *C. l. nubilus* from the

conterminous western United States ( $\Phi_{ST} = 0.550$ ) led them to assert that this analysis supported their view that these coastal wolves were evolutionarily distinct from inland wolves, including *C. l. nubilus*. However, the large proportion of unique, and apparently extinct, haplotypes in the historical sample contributes to an exaggerated measure of divergence between the coastal populations and historical inland *C. l. nubilus*. A different picture emerges when examining the phylogenetic relationships of the haplotypes. The most common haplotype (lu38) in coastal British Columbia also occurs in historical Kansas and Nebraska samples (Table 4, Leonard et al. 2005), and nearly all coastal haplotypes are in the same phylogroup as the historical western *C. l. nubilus* haplotypes (Weckworth et al. 2010, Figure 2). These relationships are consistent with the coastal British Columbia and southeast Alaska wolves being a northward extension of *C. l. nubilus*.

The wolves of southeast Alaska are the descendents of a southern Pleistocene refugium or southern clade (*C. l. nubilus*) that migrated north along the west coast as glacial ice retreated inland some 12,000 years ago. The northern continental wolves of the inland localities in this study are an admixture of northern refugium wolves from Beringia (*C. l. occidentalis*) with a substantial contribution from southern clade wolves that migrated north as glacial ice retreated from the Great Plains. Northwest coastal wolves are a vestige of a widespread southern phylogroup that was extirpated during the last century.

The Pacific Coastal region of British Columbia and southeastern Alaska is occupied by very closely-related populations of wolves that share phylogenetic relationships and certain haplotypes with the historical *C. l. nubilus* of inland portions of the western United States. The name *Canis lupus crassodon* (type locality: Tahsis Canal, Vancouver Island) has been used for the wolves of Vancouver Island (Hall 1981), but mtDNA data do not indicate differentiation

between modern samples from Vancouver Island and the coastal mainland of British Columbia (Muñoz-Fuentes et al. 2009), and the extirpation of the original population followed by colonization by wolves of a different mtDNA haplotype (Muñoz-Fuentes et al. 2010) further argues against taxonomic distinction of the modern wolf population of the island. Furthermore, the haplotype lu68 found in historical wolves from Vancouver Island is common in coastal mainland wolves today, so there is no genetic support for taxonomic distinction of the historical population of the island.

The wolf population of coastal British Columbia was probably contiguous with the original populations of coastal Washington and Oregon, which were included by Goldman (1944) with *Canis lupus fuscus*, the type locality of which (near The Dalles, Oregon) was not coastal. Hall and Kelson (1959) included most of coastal British Columbia with the range of this subspecies, which Nowak considered a synonym of *C. l. nubilus*. Bailey (1936) identified coastal wolves of Oregon as *Canis lycaon gigas* (type locality near Vancouver, Washington). Goldman (1944) included this name as a synonym of *Canis lupus fuscus*. Genetic study of historical individuals from western Oregon and Washington would be valuable in evaluating the alternative taxonomic placements based on traditional morphology and morphometrics of the historical population of the Pacific Northwest of the United States.

Another portion of the range whose subspecies assignment is uncertain is the area west of Hudson Bay in Northwest Territories. Skeel and Carbyn (1977) found morphometric affinity with *C. l. occidentalis* from Wood Buffalo National Park, the basis of which Nowak (1995) has questioned on the basis of their inclusion of females with samples of males. The Qamanirjuaq sampling area of the microsatellite DNA study of Carmichael et al. (2008) encompasses an area from Hudson Bay west, and it straddles the boundary between the two subspecies. As a possible

consequence, its position on the neighbor-joining tree (Carmichael et al. 2008, Fig 3(B)) is only weakly supported and therefore provides little reliable information on taxonomic standing.

Hybrid or intergrade zones involving *C. l. nubilus* were discussed in sections on relationships of *C. lycaon* to gray wolves and *C. l. baileyi*. General conclusions on these intergrade zones are repeated here. *Canis lupus nubilus* forms a hybrid zone with *C. lycaon* from eastern Ontario to Minnesota and Manitoba. The hybrid zone includes wolves that have recently re-colonized Wisconsin and the Upper Peninsula of Michigan. There was historical contact between *C. l. nubilus* and *C. l. baileyi*. Information from historical specimens indicates that haplotypes of *C. l. nubilus* occurred as far south as Arizona and *C. l. baileyi* haplotypes occurred as far north as Nebraska. Wolves were subsequently extirpated from the area of this zone, and the only wolves now present near this area are *C. l. baileyi* that have been reintroduced in southern Arizona and New Mexico.

Nowak's (1983, 1995) classification and evolutionary explanation characterizes *C. l. nubilus* as a medium-size wolf that was widespread in North America at the time of arrival of the larger *C. l. occidentalis*. Morphometric analyses by Skeel and Carbyn (1977) provide general support for a comparable distribution of larger and smaller wolves in central Canada. Autosomal microsatellite DNA data (Carmichael et al. 2007, 2008) distinguish *Canis lupus nubilus* from *C. l. occidentalis* in the northeastern portion of its range, and both microsatellite and mtDNA data (Weckworth et al. 2005, 2010; Muñoz-Fuentes et al. 2009, 2010) distinguish its Pacific Coast populations from inland *C. l. occidentalis*. Historical samples of *C. l. nubilus* from the western United States (Leonard et al. 2005, Table 4 of this paper) have several unique and phylogenetically related mtDNA haplotypes. The available genetic information bearing on the

question of subspecies lends general support for *C. l. nubilus* as delineated by Nowak (1995, 2003), at least in the areas covered by those studies.

### **A General Evolutionary Interpretation**

The following evolutionary scenario is presented as an overview of the conclusions of this review in the context of the evolutionary history of modern North American *Canis*. Coyotes, *Canis rufus*, and *Canis lycaon* are modern representatives of a major and diverse clade that evolved within North America, as proposed by Wilson et al. (2000). *Canis lupus* arose in Eurasia and invaded North America at least three separate times, each invasion by one or more different clades of Eurasian *Canis lupus*. The first of these invasions was by the ancestors of *C. l. baileyi*, as suggested by Vilà et al. 1999, followed by the invasion by the ancestors of *C. l. nubilus*, which displaced *C. l. baileyi* in the northern part of its range. While expanding in North America and displacing *C. l. baileyi*, the historical *C. l. nubilus* population gained some mtDNA haplotypes from the latter (Leonard et al. 2005) in a process whereby an invading population is genetically introgressed with local genes. The distribution of *C. l. occidentalis* has the general form of an invading population, and its southward expansion and displacement of *C. l. nubilus* may have continued into historic times. The final invasion, probably post-glacial, was by *C. l. occidentalis*, which displaced *C. l. nubilus* in the northern part of its former range. This final phase was undoubtedly more complex, as the biogeography of Beringia is complex, and at least one Beringian lineage of *C. lupus* became extinct without leaving genetic traces in modern wolves (Leonard et al. 2007). *Canis lupus* is not morphologically or genetically homogeneous or undifferentiated across North America. An interpretation that wolves of these different lineages have mixed in North America to an extent that the only geographic pattern is isolation by

distance is not supported by the geographic distribution of lineage markers. There is geographic structure in genetic composition (Tables 4, 5) that is consistent with multiple invasions of North America from Eurasia. This geographic structure on a continental scale coincides with the general distributions of the three *C. lupus* subspecies recognized in this review.

### **Final Comments and Recommendations**

The taxonomic recommendations and conclusions stated here are intended to represent the most reasonable interpretations based on the available scientific information. A very generalized summary of the findings are provided on a map in Figure 13 of this paper. Some conclusions, such as the taxonomic standing of *Canis lupus baileyi*, are more strongly supported than others. The taxonomic standing for *Canis lupus arctos* is not confirmed, but important limitations in the available data do not permit more definitive statements on its taxonomic status.

It is possible that further research will provide data that would change certain conclusions reached here. Longer sequences of mtDNA (most studies used ~200 to ~400 base pairs) could provide more robust resolution of both extant and historical populations. There are many more specimens from museum and government agency collections that have not yet had DNA characterized. Y-chromosome haplotypes from additional populations of wolves would provide an additional lineage marker to complement mtDNA data. Single nucleotide polymorphisms (SNPs) are now being studied in wolves, but only a small amount of preliminary data on wild populations have been reported (Anderson et al. 2009, Gray et al. 2009). Genomic approaches have potential to provide vast amounts of information on individual specimens, but it is not yet known how they will affect our understanding of relationships among populations and their taxonomic standing.



Even with expanded application of various genetic markers, geographic coverage or spatial sampling patterns may still limit our understanding of crucial areas. There are sampling concerns on both very broad and more localized scales. A notable example on the broadest scale is the original wolf population of the conterminous United States, which was extirpated outside of northeastern Minnesota and Isle Royale. Without genetic study of additional specimens from collections or possibly as yet unstudied or undiscovered natural deposits of bones or other persistent remains of wolves, the coverage of crucial and underrepresented geographic areas will hinder our understanding of the historical relationships of populations. Additional morphometric studies that use more objective methods and explore more alternative taxonomic arrangements have potential for improving our understanding of evolutionary relationships and their taxonomic implications, especially when integrated with genetic data.

The design of spatial sampling is also important on a more local scale (Schwartz and McKelvey 2009, Schwartz and Vucetich 2009). This is particularly important in cases where both isolation by distance and local barriers (ecological or behavioral) to interaction are suspected to be operating, as in the Great Lakes region. Grouping of spatial genetic data by state or province, for example, can obscure finer-scale patterns of contact and interaction between populations with different evolutionary histories. Geographic mapping of haplotypes in the Great Lakes region (Fain et al. 2010, Wheeldon et al. 2010) provides a more objective portrayal of these interactions and facilitates the identification of areas for further investigation.

There is scientific support for the taxa recognized here, but delineation of exact geographic boundaries presents challenges. Rather than sharp lines separating taxa, boundaries should generally be thought of as intergrade zones of variable width. These “fuzzy” boundaries are a consequence of lineages of wolves that evolved elsewhere coming into contact. Historical

or modern boundaries should also not be viewed as static or frozen in any particular time. Our understanding of the historical interactions between subspecies or genetically different populations (e.g., Leonard et al. 2005) is that they are dynamic processes and boundaries can shift over time. Even with the great dispersal capabilities of wolves and their interaction in these intergrade zones, genetic indications of the independent evolution of the wolves here recognized as species or subspecies are still discernable on a continental scale.

As stated in the Scope and Intent section at the beginning of this review, we have not evaluated non-taxonomic alternatives to subspecies classifications of wolves, such as management units or evolutionarily significant units, or the appropriateness of legal protection as distinct vertebrate population segments under the Endangered Species Act (USFWS and NOAA 1995). Even with scientific support of its taxonomic validity, a subspecies may or may not be the most suitable unit for protection or management. Weighing the value of subspecies compared to non-taxonomic units requires the evaluation of specific legal, policy, and management objectives that are decidedly beyond the scope of this review. We are aware that taxonomy can have policy and legal implications, but we have carried out this review following the rule that, “[t]he relationship between conservation and taxonomy must be unidirectional; conservation strategies should be influenced by taxonomy, but taxonomy cannot be influenced by conservation priorities” (Bowen and Carl 1999). This review was prepared to provide objective evaluations of controversial issues in wolf taxonomy for the consideration of those who have the responsibility for using the best available scientific information in concert with legal and policy considerations in developing conservation programs.

This review was initiated because of the wide range of views expressed by different researchers and research groups on some major features of relationships and classification of

North American wolves. We have endeavored to be as comprehensive and objective as possible in developing recommendations based on the total information available today, and have sought to reconcile differing interpretations in the literature whenever possible.

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### **References**

- Adams JR, Kelly BT, Waits LP. 2003. Using faecal DNA sampling and GIS to monitor hybridization between red wolves (*Canis rufus*) and coyotes (*Canis latrans*). *Molecular Ecology* 12:2175-2186.
- Allen GM, Barbour T. 1937. The Newfoundland wolf. *Journal of Mammalogy* 18(2):229-234.

- 2985 American Ornithologists' Union. 1983. Check-list of North American birds. Washington,  
2986 District of Columbia: American Ornithologists' Union.
- 2987 Anderson TM, vonHoldt BM, Candille SI, Musiani M, Greco C, Stahler DR, Smith DW,  
2988 Padhukasahasram B, Randi E, Leonard JA, Bustamante CD, Ostrander EA, Tang H,  
2989 Wayne RK, Barsh GS. 2009. Molecular and evolutionary history of melanism in North  
2990 American gray wolves. *Science* 323:1339-1343
- 2991 Avise JC. 2004. Molecular markers, natural history, and evolution, 2<sup>nd</sup> edition. Sunderland,  
2992 Massachusetts: Sinauer.
- 2993 Avise JC., Walker D, Johns GC. 1998. Speciation durations and Pleistocene effects on  
2994 vertebrate phylogeography. *Proceedings of the Royal Society B* 265:1707-1712.
- 2995 Bailey V. 1936. The mammals and life zones of Oregon. *North American Fauna* Number 55.
- 2996 Baker RJ, Bradley LC, Bradley RD, Dragoo JW, Engstrom MD, Hoffman FS, Jones CA, Reid F,  
2997 Rice DW, Jones C. 2003. Revised checklist of North American mammals north of  
2998 Mexico. Museum of Texas Tech University. Occasional Papers Number 229. December  
2999 1, 2003.
- 3000 Bannasch DL, Bannasch MJ, Ryun JR, Famula TR, Pederson NC. 2005 Y chromosome  
3001 haplotype analysis in purebred dogs. *Mammalian Genome* 16:273-280.
- 3002 Bertorelle G, Excoffier L. 1998. Inferring admixture proportions from molecular data.  
3003 *Molecular Biology and Evolution* 15:1298-1311.
- 3004 Bogan MA, Mehlhop P. 1983. Systematic relationships of gray wolves (*Canis lupus*) in  
3005 southwestern North America. Museum of Southwestern Biology, University of New  
3006 Mexico. Occasional Papers Number 1. 21 pp.

- 3007 Bowen BW, Karl SA. 1999. In war, truth is the first casualty. *Conservation Biology* 13:1013-  
3008 1016.
- 3009 Brewster WG, Fritts SH. 1995. Taxonomy and genetics of the gray wolf in western North  
3010 America: a review. Pages 353-373 in Carbyn LN, Fritts SH, Seip DR editors.  
3011 Proceedings of the Second North American Symposium on Wolves. Edmonton, Alberta:  
3012 Canadian Circumpolar Institute, University of Alberta.
- 3013 Brown WM, George M Jr., Wilson AC. 1979. Rapid evolution of animal mitochondrial DNA.  
3014 Proceedings of the National Academy of Sciences USA 76:1967-1971.
- 3015 Carmichael, LE, Krizan J, Nagy JA, Dumond M, Johnson D, Veitch A, Strobeck C. 2008.  
3016 Northwest passages: conservation genetics of Arctic island wolves. *Conservation*  
3017 *Genetics* 9:879-892.
- 3018 Carmichael LE, Krizan J, Nagy JA, Fuglei E, Dumond M, Johnson D, Veitch A, Berteauz D,  
3019 Strobeck C. 2007. Historical and ecological determinants of genetic structure in arctic  
3020 canids. *Molecular Ecology* 16:3466-3483.
- 3021 Carmichael LE, Nagy JA, Larter NC, Strobeck C. 2001. Prey specialization may influence  
3022 patterns of gene flow in wolves of the Canadian Northwest. *Molecular Ecology* 10:2787-  
3023 2798.
- 3024 Carroll C. 2003. Impacts of landscape change on wolf viability in the northeastern U.S. and  
3025 southeastern Canada; implications for wolf recovery. *Wildlands Project Special Paper*  
3026 No. 5. Richmond, Vermont: Wildlands Project.
- 3027 Carroll C, Phillips MK, Lopez-Gonzales CA, Schumaker NH. 2006. Defining recovery goals  
3028 and strategies for endangered species: the wolf as a case study. *BioScience* 56:25-37.

- 3029 Carroll C, Phillips MK, Schumaker NH, Smith DW. 2003. Impacts of landscape change on  
3030 wolf restoration success: planning a reintroduction program based on static and dynamic  
3031 models. *Conservation Biology* 17:536-548.
- 3032 Cathey JC, Bickham JW, Patton JC. 1998. Introgressive hybridization and non-concordant  
3033 evolutionary history of maternal and paternal lineages in North American deer.  
3034 *Evolution* 52:1224-1229.
- 3035 Coyne JA, Orr HA. 2004. *Speciation*. Sunderland, Massachusetts: Sinauer.
- 3036 Cronin MA, Mech DL. 2009. Problems with the claim of ecotype and taxon status of the wolf  
3037 in the Great Lakes region. *Molecular Ecology* 18:4991-4993.
- 3038 Currat M, Ruedi M, Petit RJ, Excoffier L. 2008. The hidden side of invasions: massive  
3039 introgression by local genes. *Evolution* 62:1908-1920.
- 3040 Dawes PR, Elander M, Ericson M. 1986. The wolf (*Canis lupus*) in Greenland: A historical  
3041 review and present status. *Arctic* 39:119-132.
- 3042 de Queiroz K. 2007. Species concepts and species delimitation. *Systematic Biology* 56:879-  
3043 886.
- 3044 Excoffier L, Smouse PE, Quattro JM. 1992. Analysis of molecular variance inferred from metric  
3045 distances among DNA haplotypes: application to human mitochondrial DNA restriction  
3046 data. *Genetics* 131:479-491.
- 3047 Fain SR, Straughan DJ, Taylor BF. 2010. Genetic outcomes of eastern timber wolf recovery in  
3048 the western Great Lakes States. *Conservation Genetics* 11:1747-1765.
- 3049 Feng J, Lajia C, Taylor DJ, Webster MS. 2001. Genetic distinctiveness of endangered dwarf  
3050 blue sheep (*Pseudois nayaur schaeferi*): Evidence from mitochondrial control region and  
3051 Y-linked intron sequences. *Journal of Heredity* 92:9-15.

- 3052 Fitzpatrick BM, Johnson JR, Kump DK, Shaffer HB, Smith JJ, Voss SR. 2009. Rapid fixation  
3053 of non-native alleles revealed by genome-wide SNP analysis of hybrid tiger salamanders.  
3054 BMC Evolutionary Biology 9:176. doi:10.1186/1471-2148-9-176.
- 3055 Forbes, SH, Boyd DK. 1996. Genetic variation of naturally colonizing wolves in the central  
3056 Rocky Mountains. Conservation Biology 10:1082-1090.
- 3057 Forbes, SH, Boyd DK. 1997. Genetic structure and migration in native and reintroduced Rock  
3058 Mountain wolf populations. Conservation Biology 10:1226-1234).
- 3059 García-Moreno J, Matocq MD, Roy RS, Geffen E, Wayne RK. 1996. Relationships and genetic  
3060 purity of the endangered Mexican wolf based on analysis of microsatellite loci.  
3061 Conservation Biology 10:376-389.
- 3062 Geffen E, Anderson MJ, Wayne RK. 2004. Climate and habitat barriers to dispersal in the  
3063 highly mobile grey wolf. Molecular Ecology 13:2481-2490.
- 3064 Goldman EA. 1937. The wolves of North America. Journal of Mammalogy 18:37-45.
- 3065 Goldman EA. 1944. Classification of wolves. Part II in Young SP, Goldman EA, editors. The  
3066 wolves of North America. Washington, District of Columbia: The American Wildlife  
3067 Institute.
- 3068 Goldstein PZ, DeSalle R, Amato G, Vogler AP. 2000. Conservation genetics at the species  
3069 boundary. Conservation Biology 14:120-131.
- 3070 Good JM, Hird S, Reid N, Demboski JR, Stepan SJ, Martin-Nims TR, Sullivan J. 2008.  
3071 Ancient hybridization and mitochondrial capture between two species of chipmunks.  
3072 Molecular Ecology 17:1313-1327.
- 3073 Grant PR, Grant BR. 1992. Hybridization of bird species. Science 256:193-197.

- 3074 Grant PR, Grant BR. 2006. Species before speciation is complete? *Annals of the Missouri*  
3075 *Botanical Garden* 93:94-102.
- 3076 Gray MM, Granka JM, Bustamente CD, Sutter NB, Boyko AR, Zuh L, Ostrander EA, Wayne  
3077 RK. 2009. Linkage disequilibrium and demographic history of wild and domestic  
3078 canids. *Genetics* 181:1493-1505.
- 3079 Grewal SK, Wilson PJ, Kung TK, Shami K, Theberge MT, Theberge JB, White BN. 2004. A  
3080 genetic assessment of the eastern wolf (*Canis lycaon*) in Algonquin Provincial Park.  
3081 *Journal of Mammalogy* 85:625-632.
- 3082 Hailer F, Leonard JA. 2008. Hybridization among three native North American *Canis* species in  
3083 a region of natural sympatry. *Public Library of Science ONE* 3(10):e333.  
3084 doi:10.1371/journal.pone.0003333.
- 3085 Hall ER. 1981. The mammals of North America. 2<sup>nd</sup> Edition. Volume II. New York: Wiley.
- 3086 Hall ER, Kelson KR. 1952. Comments on the taxonomy and geographic distribution of some  
3087 North American marsupials, insectivores and carnivores. University of Kansas  
3088 Publications, Museum of Natural History, 5:319-341.
- 3089 Hall ER, Kelson KR. 1959. The mammals of North America. Volume II. New York: Ronald  
3090 Press.
- 3091 Hanotte O, Tawah CL, Bradley DG, Okomo M, Verjee Y, Ochieng J, Rege JEO. 2000.  
3092 Geographic distribution and frequency of a taurine *Bos taurus* and an indicine *Bos*  
3093 *indicus* Y specific allele amongst sub-Saharan African cattle breeds. *Molecular Ecology*  
3094 9:387-396.



- 3095 Hedrick PW, Lee RN, Garrigan D. 2002. Major histocompatibility complex variation in red  
3096 wolves: evidence for common ancestry with coyotes and balancing selection. *Molecular*  
3097 *Ecology* 11:1905-1913.
- 3098 Hedrick PW, Lee RN, Parker KM. 2000. Major histocompatibility complex (MHC) variation the  
3099 endangered Mexican wolf and related canids. *Heredity* 85:617-624.
- 3100 Hedrick PW, Miller PS, Geffen E, Wayne RK. 1997. Genetic evaluation of the three captive  
3101 Mexican wolf lineages. *Zoo Biology* 16:47-69.
- 3102 Hendry AP. 2009. Speciation. *Nature* 458:162-164.
- 3103 Hudson RR, Coyne JA. 2002. Mathematical consequences of the genealogical species concept.  
3104 *Evolution* 56:1557-1565.
- 3105 International Commission on Zoological Nomenclature. 1999. International code for zoological  
3106 nomenclature. 4<sup>th</sup> edition. London: The International Trust for Zoological  
3107 Nomenclature. <http://www.iczn.org/iczn/index.jsp>. Accessed June 23, 2009.
- 3108 James FC, McCulloch CE. 1990. Multivariate analysis in ecology and systematics: panacea or  
3109 Pandora's box? *Annual Review of Ecology and Systematics* 21:129-166.
- 3110 Jolicoeur P. 1959. Multivariate geographical variation in the wolf *Canis lupus* L. *Evolution*  
3111 13:283-299.
- 3112 Kays R, Curtis A, Kirchman JJ. 2009. Rapid adaptive evolution of northeastern coyotes via  
3113 hybridization with wolves. *Biology Letters* 6:89-93.
- 3114 Koblmüller S, Nord M, Wayne RK, Leonard JA. 2009a. Origin and status of the Great Lakes  
3115 wolf. *Molecular Ecology* 18:2313-2326.
- 3116 Koblmüller S, Nord M, Wayne RK, Leonard JA. 2009b. More is better. *Molecular Ecology*  
3117 18:4991-4993.

- 3118 Kolenosky GB, Standfield RO. 1975. Morphological and ecological variation among gray  
3119 wolves (*Canis lupus*) of Ontario, Canada. Pages 62-72 in Fox MW, editor. The wild  
3120 canids; their systematic, behavioral ecology and evolution. New York: Van Nostrand  
3121 Reinhold.
- 3122 Kurtén B, Anderson E. 1980. Pleistocene mammals of North America. New York: Columbia  
3123 University Press.
- 3124 Kyle CJ, Johnson AR, Patterson BR, Wilson PJ, Grewal SK, White BN. 2006. Genetic nature  
3125 of eastern wolves: Past, present and future. *Conservation Genetics* 7:273-287.
- 3126 Kyle CJ, Johnson AR, Patterson BR, Wilson PJ, White BN. 2008. The conspecific nature of  
3127 eastern and red wolves: Conservation and management implications. *Conservation*  
3128 *Genetics* 9:699-701.
- 3129 Lance RF, Kennedy ML, Leberg PL. 2000. Classification bias in discriminant function analyses  
3130 used to evaluate putatively different taxa. *Journal of Mammalogy* 81:245-249.
- 3131 Lawrence B, Bossert WH. 1967. Multiple character analysis of *Canis lupus*, *latrans*, and  
3132 *familiaris*, with a discussion of the relationships of *Canis niger*. *American Zoologist*  
3133 7:223-232.
- 3134 Lehman N, Eisenhawer A, Hansen K, Mech LD, Peterson RO, Gogan PJP, Wayne RK. 1991.  
3135 Introgression of coyote mitochondrial DNA into sympatric North American gray wolf  
3136 populations. *Evolution* 45:104-119.
- 3137 Leonard JA, Vilà C, Wayne RK. 2005. Legacy lost: genetic variability and population size of  
3138 extirpated US Grey wolves (*Canis lupus*). *Molecular Ecology* 14:9-17.

- 3139 Leonard JA, Vilà C, Fox-Dobbs K, Koch PL, Wayne RK, Van Valkenburgh, G. 2007.  
3140 Megafaunal extinctions and the disappearance of a specialized wolf ecomorph. *Current*  
3141 *Biology* 17:1146-1150.
- 3142 Leonard JA, Wayne RK. 2008. Great Lakes wolves were not restored. *Biology Letters* 4:95-98.
- 3143 Leonard J.A, Wayne RK. 2009. Wishful thinking: imagining that the current Great Lakes wolf  
3144 is the same entity that existed historically. *Biology Letters* 5:67-68.
- 3145 Maldonado JE, Hertel F, Vilà. 2004. Discordant patterns of morphological variation in  
3146 genetically divergent populations of ornate shrews (*Sorex ornatus*). *Journal of*  
3147 *Mammalogy* 85:886-896.
- 3148 Mank JE, Carson JE, Brittingham MC. 2004. A century of hybridization: Decreasing genetic  
3149 distance between American black ducks and mallards. *Conservation Genetics* 5:395-403.
- 3150 Mayr E. 1942. *Systematics and the origin of species*. New York: Columbia University Press.
- 3151 Mayr E. 1963. *Animal species and evolution*. Cambridge, Massachusetts: Belknap Press.
- 3152 Mayr E. 1969. *Principles of systematic zoology*. New York: McGraw-Hill.
- 3153 Mayr E. 1970. *Populations, species, and evolution*. Cambridge, Massachusetts: Belknap Press.
- 3154 McCay BD, Zink RM. 2010. The causes of mitochondrial DNA gene tree parphyly in birds.  
3155 *Molecular Phylogenetics and Evolution* 54:647-650.
- 3156 McCracken KG, Johnson WP, Sheldon FH. Molecular population genetics, phylogeography,  
3157 and conservation biology of the mottled duck (*Anas fulvigula*). *Conservation Genetics*  
3158 2:87-102.
- 3159 Mech LD. 1974. *Canis lupus*. *Mammalian Species* 37:106.

- 3160 Mech LD. 1987. Age, season, distance, direction, and social aspects of wolf dispersal from a  
3161 Minnesota pack. Pages 55-74 in Chepko-Sade BD and Halpin ZT, editors. Mammalian  
3162 dispersal patterns. Chicago: University of Chicago Press.
- 3163 Mech LD. 2009. Crying wolf: concluding that wolves were not restored. *Biology Letters* 5:65-  
3164 66.
- 3165 Mech LD. 2010. What is the taxonomic identity of Minnesota wolves? *Canadian Journal of*  
3166 *Zoology* 88:129-138.
- 3167 Mech LD, Federoff NE. 2002.  $\alpha_1$ Antitrypsin polymorphism and systematic of North American  
3168 wolves. *Canadian Journal of Zoology* 80:961-963.
- 3169 Mech LD, Frenzel LD. 1971. The possible occurrence of the Great Plains wolf in northeastern  
3170 Minnesota. Pages 60-62 in Mech LD, Frenzel LD editors. *Ecological studies of the*  
3171 *timber wolf in northeastern Minnesota*. Saint Paul, Minnesota: North Central Forest  
3172 Experiment Station.
- 3173 Mech LD, Nowak RM. In review. Use of cranial characters in Minnesota wolf taxonomy.
- 3174 Mech LD, Paul WJ. 2008. Wolf body mass cline across Minnesota related to taxonomy?  
3175 *Canadian Journal of Zoology* 86:933-936.
- 3176 Miller GS Jr. 1912. The names of large wolves of northern and western North America.  
3177 *Smithsonian Miscellaneous Collections* 59(15).
- 3178 Moritz C. Defining evolutionarily significant units for conservation. *Trends in Ecology and*  
3179 *Evolution* 9:373-375.
- 3180 Mulders R. 1997. Geographic variation in the cranial morphology of the wolf (*Canis lupus*) in  
3181 northern Canada. Master of Science Thesis. Sudbury, Ontario: Laurentian University.

- 3182 Muñoz-Fuentes V, Darimont CT, Paquet PC, Leonard JA. 2010. The genetic legacy of  
3183 extirpation and re-colonization in Vancouver Island Wolves. *Conservation Genetics*  
3184 11:547-556.
- 3185 Muñoz-Fuentes V, Darimont CT, Wayne RK, Paquet PC, Leonard JA. 2009. Ecological factors  
3186 drive differentiation in wolves from British Columbia. *Journal of Biogeography*  
3187 36:1516-1531.
- 3188 Murray DL, Waits L. 2007. Taxonomic status and conservation of the endangered red wolf: a  
3189 response to Kyle et al. (2006). *Conservation Genetics* 8:1483-1485.
- 3190 Musiani M, Leonard JA, Cluff HD, Gates CC, Mariani S, Paquet PC, Vilà C, Wayne RK. 2007.  
3191 Differentiation of tundra/taiga and boreal coniferous forest wolves: genetics, coat colour  
3192 and association with migratory caribou. *Molecular Ecology* 16:4149-4170.
- 3193 Nei, M. 1972. Genetic distance between populations. *American Naturalist* 106:283-292.
- 3194 Nei, M. 1978. Estimation of average heterozygosity and genetic distance from a small number  
3195 of individuals. *Genetics* 89:583-590.
- 3196 Nelson EW, Goldman EA. 1929. A new wolf from Mexico. *Journal of Mammalogy* 10:165-  
3197 166.
- 3198 Nevado B, Koblmüller SL, Sturmbauer C, Snoeks J, Usano-Aleman J, Verheyen E. 2009.  
3199 Complete mitochondrial DNA replacement in a Lake Tanganyika cichlid fish. *Molecular*  
3200 *Ecology* 18:4240-4255.
- 3201 Nowak RM. 1979. North American Quaternary *Canis*. Monograph of the Museum of Natural  
3202 History, University of Kansas 6:1-154.

- 3203 Nowak RM. 1983. A perspective on the taxonomy of wolves in North America. Pages 10-19 in  
3204 Carbyn LN, editor. Wolves in Canada and Alaska: their status, biology, and  
3205 management. Edmonton, Alberta: Canadian Wildlife Service.
- 3206 Nowak RM. 1995. Another look at wolf taxonomy. Pages 375-397 in Carbyn LN, Fritts SH,  
3207 Seip DR, editors. Proceedings of the second North American symposium on wolves.  
3208 Edmonton, Alberta: Canadian Circumpolar Institute, University of Alberta.
- 3209 Nowak RM. 2002. The original status of wolves in eastern North America. Southeastern  
3210 Naturalist. 1:95-130.
- 3211 Nowak RM. 2003. Wolf evolution and taxonomy. Pages 239-258 in Mech LD, Boitani L,  
3212 editors. Wolves, behavior, ecology, and conservation. Chicago: University of Chicago  
3213 Press.
- 3214 Nowak RM. 2009. Chapter 15; taxonomy, morphology, and genetics of wolves in the Great  
3215 Lakes region. Pages 233-250 in Wydeven AP, Van Deelen TR, Heske E, editors.  
3216 Recovery of wolves in the Great Lakes region. New York: Springer.
- 3217 Oakleaf JK, Murray DL, Oakleaf JR, Bangs EE, Mack CM, Smith DW, Fontaine JA, Jimenez  
3218 MD, Meier TJ, Niemeyer CC. 2006. Habitat selection by recolonizing wolves in the  
3219 northern Rocky Mountains of the United States. Journal of Wildlife Management  
3220 70:554-563.
- 3221 Olivier M, Breen M, Binns MM, Lust G. 1999. Localization and characterization of nucleotide  
3222 sequences from the canine Y chromosome. Chromosome Research 7:223-233.
- 3223 Olivier M, Lust G. 1998. Two new nucleotide sequences specific for the canine Y chromosome.  
3224 Animal Genetics 29:146-149.

- 3225 Parsons, D. 1996. Case study: the Mexican wolf. Pages 101-123 in Herrera EA, Huenneke LF,  
3226 editors. New Mexico's natural heritage; biological diversity in the land of enchantment.  
3227 New Mexico Journal of Science, Volume 36, November 1996.
- 3228 Patten MA, Unitt P. 2002. Diagnosability versus mean differences of sage sparrow subspecies.  
3229 The Auk 119:26-35.
- 3230 Paetkau D, Waits LP, Clarkson PL, Craighead L, Strobeck C. 1997. An empirical evaluation of  
3231 genetic distance statistics using microsatellite data from bear (Ursidae) populations.  
3232 Genetics 147:1943-1957.
- 3233 Phillips MK, Henry VG, Kelly BT. 2003. Restoration of the red wolf. Pages 272-288 in Mech  
3234 LD, Boitani L, editors. Wolves: behavior, ecology, and conservation. Chicago:  
3235 University of Chicago Press.
- 3236 Pidancier N, Jordan S, Luikart G, Taberlet P. 2006. Evolutionary history of the genus *Capra*  
3237 (Mammalia, Artiodactyla): Discordance between mitochondrial DNA and Y-  
3238 chromosome phylogenies. Molecular Phylogenetics and Evolution 40:739-749.
- 3239 Pilot M, Jędrzewewski W, Branicki W, Sidorovich VE, Jędrzewewski B, Stachura K, Funk SM.  
3240 2006. Ecological factors influence population genetic structure of European grey wolves.  
3241 Molecular Ecology 15:4533-4553.
- 3242 Pocock RI. 1935. The races of *Canis lupus*. Proceedings of the Zoological Society, London,  
3243 Part 3:647-686.
- 3244 Pritchard JK, Stephens M, Donnelly P. 2000. Inference of population structure using multilocus  
3245 genotype data. Genetics 155:945-959.
- 3246 Reich DE, Wayne RK, Goldstein DB. 1999. Genetic evidence for a recent origin by  
3247 hybridization of red wolves. Molecular Ecology 8:139-144.

- 3248 Roy MS, Geffen E, Ostrander D, Wayne RK. 1994. Patterns of differentiation and hybridization  
3249 in North American wolflike canids, revealed by analysis of microsatellite loci. *Molecular*  
3250 *Biology and Evolution* 11:533-570.
- 3251 Roy MS, Geffen E, Smith E, Wayne RK. 1996. Molecular genetics of pre-1940 red wolves.  
3252 *Conservation Biology* 10:1413-1424.
- 3253 Rutledge LY, Bos KJ, Pearce RJ, White BN. 2010a. Genetic and morphometric analysis of  
3254 sixteenth century *Canis* skull fragments: implication for historic eastern and gray wolf  
3255 distribution in North America. *Conservation Genetics* 11:1273-1281.
- 3256 Rutledge LY, Garroway CJ, Loveless KM, Patterson BR. 2010b. Genetic differentiation of  
3257 eastern wolves in Algonquin Park despite bridging gene flow between coyotes and grey  
3258 wolves. *Heredity* 105:520-531.
- 3259 Rutledge LY, Patterson BR, White BN. 2010c. Analysis of *Canis* mitochondrial DNA  
3260 demonstrates high concordance between the control region and ATPase genes. *BMC*  
3261 *Evolutionary Biology* 10:25.
- 3262 Say T. 1823. Chapter IX in Long SH. Account of an expedition from Pittsburgh to the Rock  
3263 Mountains performed in the years 1819 and 1820, Volume I. Philadelphia: Carey and  
3264 Lea.
- 3265 Schmitz, OJ, Kolenosky GB. 1985. Wolves and coyotes in Ontario: morphological relationships  
3266 and origins. *Canadian Journal of Zoology* 63:1130-1137.
- 3267 Schwartz MK, McKelvey KS. 2009. Why sampling scheme matters: the effect of sampling  
3268 scheme on landscape genetic results. *Conservation Genetics* 10:441-452.



- 3269 Schwartz MK, Pilgrim KL, McKelvey KS, Lindquist EL, Claar JJ, Loch S, Ruggiero LF. 2004.  
3270 Hybridization between Canada lynx and bobcats: genetic results and management  
3271 implications. *Conservation Genetics* 5:349-355.
- 3272 Schwartz MK, Vucetich JA. 2009. Molecules and beyond: assessing the distinctness of the  
3273 Great Lakes wolf. *Molecular Ecology* 18:2307-2309.
- 3274 Schwenk K, Brede N, Streit B. 2008. Introduction. Extent, processes and evolutionary impact  
3275 of interspecific hybridization in animals. *Philosophical Transactions of the Royal Society*  
3276 B 363:2805-2811.
- 3277 Seehausen, O. 2006. Conservation: losing biodiversity by reverse speciation. *Current Biology*  
3278 16(9):R334. Skeel MA, Carbyn LN. 1977. The morphological relationship of gray  
3279 wolves (*Canis lupus*) in national parks of central Canada. *Canadian Journal of Zoology*  
3280 55:737-747.
- 3281 Seehausen, O, van Alphen JJM, Witte F. 1997. Cichlid fish diversity threatened by  
3282 eutrophication that curbs sexual selection. *Science* 277:1808-1811.
- 3283 Shami K. 2002. Evaluating the change in distribution of the eastern timber wolf (*Canis lycaon*)  
3284 using the Y-chromosome. MSc thesis. Hamilton, Ontario: McMaster University.
- 3285 Sites JW Jr., Marshall JC. 2004. Operational criteria for delimiting species. *Annual Review of*  
3286 *Ecology and Systematics* 35:199-227.
- 3287 Skeel MA, Carbyn L. 1977. The morphological relationship of gray wolves (*Canis lupus*) in  
3288 national parks of central Canada. *Canadian Journal of Zoology* 55:737-747.
- 3289 Slatkin M. 1985. Rare alleles as indicators of gene flow. *Evolution* 39:53-65.
- 3290 Standfield R. 1970. Some considerations on the taxonomy of wolves in Ontario. Pages 32-38 in  
3291 Jorgensen SE, Faulkner CE, Mech LD, editors. *Proceedings of a symposium on wolf*

3292 management in selected areas of North America. Twin Cities, Minnesota: United States  
3293 Bureau of Sport Fisheries and Wildlife.

3294 Stronen AV, Forbes GJ, Sallows T, Goulet G, Musiani M, Paquet, PC. 2010. Wolf body mass,  
3295 skull morphology, and mitochondrial DNA haplotypes in the Riding Mountain National  
3296 Park Region of Manitoba, Canada. *Canadian Journal of Zoology* 88:496-507.

3297 Sundqvist A-K, Ellegren H, Olivier M, Vilà C. 2001. Y chromosome haplotyping in  
3298 Scandinavian wolves (*Canis lupus*) based on microsatellite markers. *Molecular Ecology*  
3299 10:1959-1966.

3300 Templeton AR. 1989. The meaning of species and speciation: a genetic perspective. Pages 3-27  
3301 in Otte D, Endler JA editors. *Speciation and its consequences*. Sunderland,  
3302 Massachusetts: Sinauer Associates.

3303 [USFWS] U.S. Fish and Wildlife Service. 1990. Red wolf recovery/species survival plan.  
3304 Atlanta, Georgia.

3305 [USFWS and NOAA] U.S. Fish and Wildlife Service and National Oceanic and Atmospheric  
3306 Administration. 1995. Policy regarding the recognition of distinct vertebrate population  
3307 segments under the Endangered Species Act. 61 *Federal Register* 4722-4725.

3308 Vilà C, Amorim IR, Leonard JA, Posada D, Catroviello J, Petrucci-Fonseca F, Crandall KA,  
3309 Ellegren H, Wayne RK. 1999. Mitochondrial DNA phylogeography and population  
3310 history of the grey wolf *Canis lupus*. *Molecular Ecology* 8:2089-2103.

3311 vonHoldt BM, Stahler DR, Smith DW, Earl DA, Pollinger JP, Wayne RK. 2008. The genealogy  
3312 and genetic viability of reintroduced Yellowstone grey wolves. *Molecular Ecology*  
3313 17:252–274.

3314 vonHoldt BM, Stahler DR, Banks EE, Smith DW, Jimenez MD, Mack CM, Niemeyer CC,  
3315 Pollinger JP, Wayne RK. 2010. A novel assessment of population structure and gene

- 3316 flow in grey wolf populations of the Northern Rocky Mountains of the United States.  
3317 Molecular Ecology 19:4412-4427.
- 3318 vonHoldt BM, Pollinger JP, Earl DA, Knowles JC, Boyko AR, Parker H, Geffen E, Pilot M,  
3319 Jedrzejewski W, Jedrzejewski B, Sidorovich V, Creco C, Ettore R, Musiani M, Kays R,  
3320 Bustamante CD, Ostrander EA, Novembre J, Wayne RK. In press. A genome-wide  
3321 perspective on the evolutionary history of enigmatic wolf-like canids. Genome Research  
3322 21:1294-1305.
- 3323 Wayne RK, Jenks SM. 1991. Mitochondrial DNA analysis implying extensive hybridization of  
3324 the endangered red wolf *Canis rufus*. Nature 351:565-568.
- 3325 Wayne RK, Lehman N, Allard MW, Honeycutt RL. 1992. Mitochondrial DNA variability of  
3326 the gray wolf: Genetic consequences of population decline and habitat fragmentation.  
3327 Conservation Biology 6:559-569.
- 3328 Wayne RK, Lehman N, Fuller TK. 1995. Conservation genetics of the gray wolf. Pages 399-  
3329 407 in Carbyn LN, Fritts SH, Seip DR, editors. Proceedings of the second North  
3330 American symposium on wolves. Edmonton, Alberta: Canadian Circumpolar Institute,  
3331 University of Alberta.
- 3332 Wayne RK, Vilà C. 2003. Molecular genetic studies of wolves. Pages 218-238 in Mech LD,  
3333 Boitani L, editors. Wolves: behavior, ecology, and conservation. Chicago: University  
3334 of Chicago Press.
- 3335 Weckworth BV, Talbot SL, Cook JA. 2010. Phylogeography of wolves (*Canis lupus*) in the  
3336 Pacific Northwest. Journal of Mammalogy 91:363-375.

- 3337 Weckworth BV, Talbot S, Sage GK, Person DK, Cook J. 2005. A signal for independent  
3338 coastal and continental histories among North American wolves. *Molecular Ecology*  
3339 14:917-931.
- 3340 Weir BS, Cockerham CC. 1984. Estimating  $F$  statistics for the analysis of population structure.  
3341 *Evolution* 38:1358-1370.
- 3342 Wheeldon T, Patterson BR, White BN. 2010. Sympatric wolf and coyote populations of the  
3343 western Great Lakes region are reproductively isolated. *Molecular Ecology* 19:4428-  
3344 4440.
- 3345 Wheeldon T, White BM. 2009. Genetic analysis of historical western Great Lakes region wolf  
3346 samples reveals early *Canis lupus/lycaon* hybridization. *Biology Letters* 5:101-104.
- 3347 Wiley EO. 1981. *Phylogenetics; the theory and practice of phylogenetic systematics*. New  
3348 York: Wiley.
- 3349 Wilson EO, Brown WL. 1953. The subspecies concept and its taxonomic application.  
3350 *Systematic Zoology* 2:97-111.
- 3351 Wilson DE, Reeder DM, editors. 2005. *Mammal species of the world; a taxonomic and*  
3352 *geographic reference*. 3<sup>rd</sup> edition. Baltimore, Maryland: Johns Hopkins University  
3353 Press.
- 3354 Wilson PJ, Grewal S, Lawford ID, Heal JNM, Granacki AG, Pennock D, Theberge JB, Theberge  
3355 MT, Voigt DR, Waddell W, Chambers RE, Paquet PC, Goulet G, Cluff D, White BN.  
3356 2000. DNA profiles of the eastern Canadian wolf and the red wolf provide evidence for a  
3357 common evolutionary history independent of the gray wolf. *Canadian Journal of*  
3358 *Zoology* 78:2156-2166.

- 3359 Wilson PJ, Grewal S, McFadden T, Chambers RC, White BN. 2003. Mitochondrial DNA  
3360 extracted from eastern North American wolves killed in the 1800s is not of gray wolf  
3361 origin. *Canadian Journal of Zoology* 81:936-940.
- 3362 Wilson PJ, Grewal SK, Mallory FF, White BN. 2009. Genetic characterization of hybrid wolves  
3363 across Ontario. *Journal of Heredity* 2009:100 (Supplement 1):S80-S89.
- 3364 Wright, S. 1951. The genetical structure of populations. *Annals of Eugenics* 15:323-354.
- 3365 Zink RM. 2003. The role of subspecies in obscuring avian biological diversity and misleading  
3366 conservation policy. *Proceedings of the Royal Society of London* 271:561-564.
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- 3368
- 3369 Table captions:
- 3370
- 3371 Table 1. Available names for North American “gray” wolves in order of their publication date  
3372 (does not include coyotes). Based on Goldman (1944), Hall and Kelson (1959) and Hall (1981).
- 3373
- 3374 Table 2. Subspecies of *Canis lupus* recognized by Hall and Kelson (1959) and Hall (1981),  
3375 largely based on Goldman (1944). The five subspecies recognized by Nowak (1995) in his  
3376 revision are in **bold** and each is followed by its synonyms as recognized in his reclassification.
- 3377
- 3378 Table 3. Subspecies of *Canis rufus* (Goldman 1944, Nowak 1979, Hall 1981).
- 3379
- 3380 Table 4. Number of individuals identified for 35 mtDNA-control region sequence haplotypes  
3381 found in wolves from coastal locations in Alaska (AK) and British Columbia (BC) and interior

locations in Alaska, Yukon, Northwest Territories (NWT), British Columbia, Alberta (ALTA), Saskatchewan (SASK), Manitoba (MAN), Minnesota, Wisconsin and Michigan (GLS) and eastern Ontario (EONT). The historical *C. l. nubilus* samples were combined from locations in North Dakota, Colorado, Kansas, Nebraska, Oklahoma and New Mexico; and the historical *C. l. baileyi* samples were combined from locations in Arizona, New Mexico and northern Mexico (Leonard et al 2005).

Table 5. Number of individuals identified for 33 Y-chromosome haplotypes found in wolves from locations in Alaska (AK), Northwest Territories (NWT), British Columbia (BC), Alberta (ALTA), Great Lakes states of Minnesota, Wisconsin and Michigan (GLS), Texas (TX) as well as dog breeds originating in the Americas (NW) and Europe (OW). Haplotypes were derived from the dog Y-chromosome loci MS34A, MS34B, MS41A and MS41B (Olivier & Lust 1998, Sundqvist et al 2001).

Table 6. Genetic distances based on microsatellite DNA between paired samples of eastern wolf (Southern Quebec, Algonquin, western Great Lakes), other wolves, and coyotes. Genetic distances between samples with substantial eastern wolf representation are indicated in bold. Koblmüller et al. (2009a) used the notation  $\theta_{ST}$  to report the coancestry parameter  $\theta$  of Weir and Cockerham (1984), which they consider to be generally comparable to  $F_{ST}$ , the fixation index of Wright (1951).  $D$  is the unbiased genetic distance of Nei (1978).

Table 7. Private alleles among wolf populations with respect to populations covered in each cited source. Values representing eastern wolf are in bold.

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3406 Table 8. Mitochondrial DNA sequence divergences.

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3408 Table 9. Mitochondrial DNA sequence divergences within the coyote clade

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3428 Table 1.

Species or subspecies	Author(s)	Date	Described as:	
			Species	Subspecies
<i>Canis lycaon</i>	Schreber	1775	X	
<i>Canis nubilus</i>	Say	1823	X	
<i>Canis lupus occidentalis</i>	Richardson	1829		X
<i>Canis lupus fuscus</i>	Richardson	1839		X
<i>Canis lupus canadensis</i>	Blainville	1843		X
<i>Lupus gigas</i>	Townsend	1850	X	
<i>Canis lupus</i> var. <i>rufus</i>	Audubon & Bachman	1851		X
<i>Canis occidentalis griseoalbus</i>	Baird	1858		X
<i>Canis pambasileus</i>	Elliot	1905	X	
<i>Canis floridanus</i>	Miller	1912	X	
<i>Canis tundrarum</i>	Miller	1912	X	
<i>Canis nubilus baileyi</i>	Nelson & Goldman	1929		X
<i>Canis occidentalis crassodon</i>	Hall	1932		X
<i>Canis lupus arctos</i>	Pocock	1935		X
<i>Canis lupus orion</i>	Pocock	1935		X
<i>Canis rufus gregoryi</i>	Goldman	1937		X
<i>Canis lupus beothucus</i>	Allen & Barbour	1937		X
<i>Canis lupus labradorius</i>	Goldman	1937		X
<i>Canis lupus ligoni</i>	Goldman	1937		X
<i>Canis lupus youngi</i>	Goldman	1937		X
<i>Canis lupus irremotus</i>	Goldman	1937		X
<i>Canis lupus monstrabilis</i>	Goldman	1937		X
<i>Canis lupus mogollonensis</i>	Goldman	1937		X
<i>Canis tundrarum ungavensis</i>	Comeau	1940		X
<i>Canis lupus alces</i>	Goldman	1941		X
<i>Canis lupus columbianus</i>	Goldman	1941		X
<i>Canis lupus hudsonicus</i>	Goldman	1941		X
<i>Canis lupus bernardi</i>	Anderson	1943		X
<i>Canis lupus mackenzii</i>	Anderson	1943		X
<i>Canis lupus manningi</i>	Anderson	1943		X
<i>Canis lupus knightii</i>	Anderson	1947		X

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3432 Table 2.

Subspecies	Author(s)	Date	Type locality
<i>Canis lupus lycaon</i>	Schreber	1775	Restricted by Goldman (1937) to vicinity of Quebec, Quebec
<i>Canis lupus baileyi</i>	Nelson & Goldman	1929	Colonia Garcia (about 60 miles SW of Casas Grandes), Chihuahua, Mexico (altitude 6,700 feet)
<i>Canis lupus nubilus</i>	Say	1823	Near Blair, Washington County, Nebraska
<i>Canis lupus fuscus</i>	Richardson	1839	Banks of Columbia river below The Dalles, Oregon
<i>Canis lupus crassodon</i>	Hall	1932	Tahsis Canal, Nootka Sound, Vancouver Island, British Columbia
<i>Canis lupus beothucus</i>	Allen & Barbour	1937	Newfoundland
<i>Canis lupus labradorius</i>	Goldman	1937	Fort Chimo, Quebec
<i>Canis lupus ligoni</i>	Goldman	1937	Head of Duncan Canal, Kupreanof Island, Alexander Archipelago, Alaska
<i>Canis lupus youngi</i>	Goldman	1937	Harts Draw, N. slope of Blue Mountains, 20 miles NE Monticello, San Juan County, Utah
<i>Canis lupus irremotus</i>	Goldman	1937	Red Lodge, Carbon County, Montana
<i>Canis lupus monstrabilis</i> <sup>1</sup>	Goldman	1937	Ten miles S of Rankin, Upton County, Texas
<i>Canis lupus mogollonensis</i> <sup>1</sup>	Goldman	1937	S. A. Creek, 10 miles NW Luna, Catron County, New Mexico
<i>Canis lupus hudsonicus</i>	Goldman	1941	Head of Schultz Lake, Keewatin, [now Nunavut], Canada
<i>Canis lupus manningi</i>	Anderson	1943	Hantzsch River, E side Foxe Basin, W side Baffin Island, District of Franklin, Northwest Territories [now Nunavut], Canada
<i>Canis lupus arctos</i>	Pocock	1935	Melville Island, Canadian Arctic
<i>Canis lupus orion</i>	Pocock	1935	Cape York, northwestern Greenland
<i>Canis lupus bernardi</i>	Anderson	1943	Cape Kellett, Banks Island, Northwest Territories, Canada
<i>Canis lupus occidentalis</i>	Richardson	1829	Restricted by Miller (1912) to Fort Simpson, Mackenzie, Canada
<i>Canis lupus griseoalbus</i>	Baird	1858	Restricted by Hall and Kelson (1952) to Cumberland House, Saskatchewan

<i>Canis lupus pambasileus</i>	Elliot	1905	Upper waters of Sushitna River, Region of Mount McKinley
<i>Canis lupus tundrarum</i>	Miller	1912	Point Barrow, Alaska
<i>Canis lupus alces</i>	Goldman	1941	Kachemak Bay, Kenai Peninsula, Alaska
<i>Canis lupus columbianus</i>	Goldman	1941	Wistaria, N side of Ootsa Lake, Coastal District, British Columbia.
<i>Canis lupus mackenzii</i>	Anderson	1943	Imnanuit, W of Kater Point, Bathurst Inlet, District of Mackenzie, Northwest Territories

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<sup>1</sup>Considered synonyms of *Canis lupus baileyi* Goldman, 1937 by Bogan and Mehlhop (1983)

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Table 3.

Subspecies	Author(s)	Date	General range (Nowak, 2002)
<i>Canis rufus rufus</i>	Audubon & Bachman	1851	Central and Gulf coast Texas & Louisiana
<i>Canis rufus floridanus</i>	Miller	1912	United States east of the Mississippi River
<i>Canis rufus gregoryi</i>	Goldman	1937	Northeast Texas to Indiana

3470 Table 4.

Haplotype	Genbank	Coastal (n=277)			Interior (n=966)								Historical samples (n=84)			
		AK	SEAK	BC	AK	Yukon	NWT	BC	ALTA	SASK	MAN	GLS	EONT	<i>C. l. nubilus</i>	<i>C. l. baileyi</i>	<i>C. lycaon</i>
lu38 <sup>a,h</sup> , F <sup>b</sup> , I <sup>b</sup>	AF812731		127 <sup>b</sup>	57 <sup>a</sup>	3 <sup>a</sup>		4 <sup>a</sup>	23 <sup>a</sup> ,5 <sup>b</sup>						3 <sup>h</sup>		
lu48 <sup>h</sup>	AY812733													1 <sup>h</sup>		
lu49 <sup>h</sup>	AY812734													1 <sup>h</sup>		
lu68 <sup>a,i</sup> , H <sup>b</sup>	FN298179		2 <sup>b</sup>	14 <sup>a</sup>										12 <sup>i</sup>		
lu32 <sup>a,h</sup> , W6 <sup>c</sup> , C22 <sup>d,i</sup>	AF005309			4 <sup>a</sup>	1 <sup>a</sup>		25 <sup>a</sup> ,232 <sup>g</sup>	31 <sup>a</sup>	20 <sup>c</sup> ,1 <sup>f</sup>	4 <sup>d</sup>	10 <sup>d</sup> , 1 <sup>i</sup>	38 <sup>c</sup>	4 <sup>c</sup>	3 <sup>h</sup>	2 <sup>h</sup>	
lu53 <sup>h</sup>	AY812738													1 <sup>h</sup>		
lu54 <sup>h</sup>	AY812739													2 <sup>h</sup>		
lu11 <sup>a</sup>	AF005300						2 <sup>a</sup>									
lu28 <sup>a,h</sup> , A <sup>b</sup> , M <sup>b</sup> , W7 <sup>c</sup> , C	AF005308	54 <sup>b</sup>			12 <sup>a</sup> ,41 <sup>b</sup> ,2 <sup>c</sup>	3 <sup>b</sup>	1 <sup>a</sup>	15 <sup>a</sup> ,7 <sup>b</sup>			25 <sup>d</sup>	1 <sup>c</sup>	3 <sup>i</sup>	7 <sup>h</sup>		
lu52 <sup>h</sup>	AY812737													1 <sup>h</sup>		
lu67 <sup>a</sup>	FM201672							1 <sup>a</sup>								
lu29 <sup>a,f</sup> , B <sup>b</sup>	AF005310	18 <sup>b</sup>			5 <sup>a</sup> ,8 <sup>b</sup>	6 <sup>b</sup> ,1 <sup>f</sup>	3 <sup>a</sup> ,57 <sup>g</sup>	2 <sup>a</sup>								
lu30 <sup>a,f</sup> , L <sup>b</sup>	AF005311				4 <sup>a</sup>	2 <sup>b</sup> ,1 <sup>f</sup>		1 <sup>b</sup>								
lu31 <sup>a,f</sup> , K <sup>b</sup> , W1 <sup>c</sup>	AF005312				4 <sup>a</sup> ,3 <sup>b</sup>	1 <sup>f</sup>	4 <sup>a</sup>	11 <sup>a</sup> ,10 <sup>b</sup>	5 <sup>c</sup>							
lu37 <sup>a</sup>	AF812730				2 <sup>a</sup> ,3 <sup>b</sup>											
lu61 <sup>a</sup> , C <sup>b</sup> , J <sup>b</sup> , 16 <sup>d</sup> , W3 <sup>c</sup>	AF812741	1 <sup>b</sup>			3 <sup>a</sup> ,9 <sup>b</sup>		1 <sup>a</sup>			20 <sup>d</sup>	1 <sup>d</sup>					
lu36 <sup>a</sup>	FM201632							2 <sup>a</sup>								
G <sup>b</sup>	GQ376506					1 <sup>b</sup>										
N <sup>b</sup>	GQ376226							1 <sup>b</sup>								
lu33 <sup>h</sup>	AF005313														4 <sup>h</sup>	
lu47 <sup>h</sup>	AY812732														1 <sup>h</sup>	
lu50 <sup>h</sup>	AY812735														6 <sup>h</sup>	
lu51 <sup>h</sup>	AY812736														1 <sup>h</sup>	
C3 <sup>d,e</sup> , C21 <sup>c</sup> , GL2 <sup>k,m</sup>	FJ213916										4 <sup>d</sup> ▲	18 <sup>c</sup> ▲				5 <sup>k</sup> ,2 <sup>m</sup>
C13 <sup>e,j,n</sup> , C12 <sup>c</sup> , GL10 <sup>m</sup>	FJ213915											58 <sup>c</sup> ▲	7 <sup>c</sup> ▲			1 <sup>i</sup> ,1 <sup>m</sup> ,1 <sup>n</sup>
C1 <sup>e,n</sup> , C4 <sup>c</sup> , GL1 <sup>k,m</sup>	FJ213914											1 <sup>c</sup> ▲	28 <sup>c</sup> ▲, 7 <sup>i</sup>			7 <sup>i</sup> ,1 <sup>i</sup> ,5 <sup>k</sup> ,1 <sup>m</sup> ,1 <sup>n</sup>
C9 <sup>c</sup>	AY267726												62 <sup>ca</sup>			1 <sup>i</sup>
C14 <sup>c</sup>	AY267731												52 <sup>ca</sup>			3 <sup>i</sup>
C16 <sup>c</sup>	AY267733												1 <sup>ca</sup>			
C17 <sup>c</sup>	AY267734												10 <sup>ca</sup>			1 <sup>i</sup>
C19 <sup>c</sup>	AY267736												38 <sup>ca</sup>			1 <sup>i</sup>
GL3 <sup>k</sup>	J. Leonard <sup>PC</sup>															1 <sup>k</sup>
GL5 <sup>k,m</sup>	J. Leonard <sup>PC</sup>															1 <sup>k</sup> ,1 <sup>m</sup>
GL6 <sup>k,m</sup>	J. Leonard <sup>PC</sup>															1 <sup>k</sup> ,1 <sup>m</sup>
GL8 <sup>k,m</sup>	J. Leonard <sup>PC</sup>															1 <sup>k</sup> ,1 <sup>m</sup>
Total		73 <sup>b</sup>	129 <sup>b</sup>	75 <sup>a</sup>	34 <sup>a</sup> ,64 <sup>b</sup> ,3 <sup>f</sup>	12 <sup>b</sup> ,3 <sup>f</sup>	40 <sup>a</sup> ,289 <sup>g</sup>	85 <sup>a</sup> ,24 <sup>b</sup>	25 <sup>c</sup> ,1 <sup>f</sup>	24 <sup>d</sup>	1 <sup>i</sup> ,36 <sup>d</sup> ,4 <sup>d</sup> ▲	39 <sup>c</sup> ,77 <sup>c</sup> ▲	3 <sup>i</sup> ,4 <sup>c</sup> ,35 <sup>c</sup> ▲,163 <sup>ca</sup>	19 <sup>h</sup> ,12 <sup>i</sup>	14 <sup>h</sup>	13 <sup>i</sup> ,2 <sup>i</sup> ,14 <sup>k</sup> ,7 <sup>m</sup> ,3 <sup>n</sup>

<sup>a</sup>Muñoz-Fuentes et al (2009a), <sup>b</sup>Weckworth et al (2010), <sup>c</sup>Fain et al (2010), <sup>d</sup>Stronen et al (2010), <sup>e</sup>Grewel et al (2004), <sup>f</sup>Vilá et al (1999), <sup>g</sup>Musiani et al (2007), <sup>h</sup>Leonard et al (2005), <sup>i</sup>Leonard and Wayne (2008), <sup>j</sup>Muñoz-Fuentes et al (2009b), <sup>k</sup>Koblmüller et al (2009), <sup>m</sup>Wheeldon and White (2009)

▲ *C. lycaon* haplotype (<sup>i</sup>Wilson et al 2000, <sup>j</sup>Wilson et al 2003), ▲ *C. latrans* haplotype (Wilson et al 2000), <sup>PC</sup>Sequence not available on GenBank - obtained by personal communication with J. Leonard

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3480 Table 5.

Haplotype	Locality										Totals
	AK <i>C. lupus</i>	NWT <i>C. lupus</i>	BC <i>C. lupus</i>	ALTA <i>C. lupus</i>	GLS <i>C. lupus</i>	GLS <i>C. lycaon</i>	<i>C. l. baileyi</i> <sup>f</sup>	<i>C. rufus</i> <sup>f</sup>	<i>C. latrans</i> TX	<i>C. familiaris</i> NW, OW	
E <sup>e</sup>						29 <sup>e</sup>					29
O <sup>e</sup>						2 <sup>e</sup>					2
R <sup>e</sup>						10 <sup>e</sup>					10
Y <sup>e</sup>						1 <sup>e</sup>					1
X <sup>e</sup>						1 <sup>e</sup>					1
H7 <sup>a</sup> , H1 <sup>d</sup>								1 <sup>d</sup>		202 <sup>a</sup>	203
H15 <sup>d</sup>								4 <sup>d</sup>	2 <sup>d</sup>		6
H28 <sup>d</sup> , G <sup>e</sup>	3 <sup>e</sup>						6 <sup>d</sup>				9
H29 <sup>d</sup>							10 <sup>d</sup>			110 <sup>a</sup>	120
H30 <sup>bc</sup> , I <sup>e</sup>	1 <sup>b</sup> , 1 <sup>e</sup>	32 <sup>c</sup>	5 <sup>e</sup>	12 <sup>e</sup>							51
H31 <sup>bc</sup>		1 <sup>b</sup> , 8 <sup>c</sup>									9
H32 <sup>bc</sup> , Z <sup>e</sup>	3 <sup>b</sup>	2 <sup>b</sup> , 21 <sup>c</sup>			7 <sup>e</sup>						33
H33 <sup>bc</sup> , A <sup>e</sup> , U	3 <sup>b</sup> , 1 <sup>e</sup>	18 <sup>c</sup>	A1 <sup>e</sup> , U2 <sup>e</sup>		A6 <sup>e</sup> , U10 <sup>e</sup>						41
H34 <sup>bc</sup>		6 <sup>b</sup> , 19 <sup>c</sup>									25
H35 <sup>bc</sup> , W <sup>e</sup>		2 <sup>b</sup> , 32 <sup>c</sup>			1 <sup>e</sup>						35
H36 <sup>bc</sup> , F <sup>e</sup>	2 <sup>b</sup> , 1 <sup>e</sup>	20 <sup>c</sup>		1 <sup>e</sup>	4 <sup>e</sup>						28
H37 <sup>b</sup>	2 <sup>b</sup>										2
H38 <sup>bc</sup> , J <sup>e</sup>		1 <sup>b</sup> , 28 <sup>c</sup>	3 <sup>e</sup>	2 <sup>e</sup>	4 <sup>e</sup>						38
H39 <sup>bc</sup> , L <sup>e</sup>	1 <sup>b</sup> , 1 <sup>e</sup>	1 <sup>b</sup>									3
H40 <sup>b</sup>		1 <sup>b</sup>									1
H41 <sup>c</sup>		2 <sup>e</sup>									2
H44 <sup>c</sup>		1 <sup>e</sup>									1
H45 <sup>c</sup>		1 <sup>e</sup>									1
H50 <sup>c</sup>		17 <sup>c</sup>									17
H52 <sup>c</sup>		5 <sup>e</sup>									5
H53 <sup>c</sup>		1 <sup>e</sup>									1
H55 <sup>c</sup> , D <sup>e</sup>		1 <sup>e</sup>			2 <sup>e</sup>						3
H58 <sup>c</sup>		2 <sup>c</sup>									2
H59 <sup>c</sup>		1 <sup>c</sup>									1
B <sup>c</sup>			1 <sup>c</sup>								1
Ff <sup>e</sup>			2 <sup>e</sup>								2
M <sup>e</sup>	1 <sup>e</sup>										1
Total	20	223	14	15	34	43	16	5	2	312	684

<sup>a</sup>Bannasch et al (2005), <sup>b</sup>Sundqvist et al (2006), <sup>c</sup>Musiani et al (2007), <sup>d</sup>Hailer & Leonard (2008), <sup>e</sup>Fain et al (2010), <sup>f</sup>Experimental population

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3485 Table 6.

Comparison	Distance	Measure	Source
Southern Quebec - Minnesota	<b>0.135</b>	<i>D</i>	Roy et al. 1994
Southern Quebec - northern Quebec	0.296	"	"
Southern Quebec - western <i>C. lupus</i>	0.251 to 0.519	"	"
Great Lakes modern - Great Lakes historic	<b>0.057</b>	$\theta$	Koblmüller et al. 2009a
Great Lakes modern - western wolves	0.075	"	"
Great Lakes modern - eastern coyotes	0.142	"	"
Great Lakes modern - western coyotes	0.133	"	"
Algonquin - proximal wolf populations	<b>0.024 to 0.055</b>	$F_{ST}$	Grewal et al. 2004
Algonquin - Abitibi Temiscamingue	0.089	"	"
Algonquin - La Verendrye Reserve	0.091	"	"
Algonquin - northeast Ontario	0.076	"	"
Algonquin - western Great Lakes states	<b>0.164</b>	"	"
W. Great Lakes States - NE Ontario & Quebec	<b>0.068</b>	"	Wheeldon 2009
Algonquin - Frontenac Axis	0.109	"	"
Algonquin - NE Ontario & Quebec	0.135	"	"
Algonquin - Manitoba	0.232	"	"
Algonquin - Northwest Territories	0.238	"	"
Algonquin - Southern Magnetawan	<b>0.022</b>	"	Wilson et al. 2009
Algonquin - Frontenac Axis	0.055	"	"
Algonquin - northwest Ontario	0.071	"	"
Algonquin - northeast Ontario	0.073	"	"
Algonquin - Minnesota	<b>0.089</b>	"	"
Algonquin - Pukaskwa National Park	0.117	"	"
Among Minnesota, Wisconsin, and Michigan	<b>0.006 to 0.016</b>	"	Fain et al. 2010
Western Great Lakes states - western wolves	0.125	"	"
Western Great Lakes states - Wisconsin coyotes	0.159	"	"
Algonquin - northeast Ontario	0.105	"	Rutledge et al. 2010b
Algonquin - Frontenac Axis	0.052	"	"
Frontenac Axis - northeast Ontario	0.120	"	"

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3489 Table 7.

Sample	Number of			Source
	Private alleles	Loci	Individuals in sample	
Southern Quebec	<b>5</b>	10	24	Roy et al. 1994
Minnesota	<b>12</b>	"	20	"
Northern Quebec	1	"	20	"
Alberta	1	"	20	"
Vancouver Island	0	"	20	"
Northwest Territories	3	"	24	"
Kenai, Alaska	0	"	19	"
Western Great Lakes states	<b>14</b>	8	124	Fain et al. 2010
Alberta	2	"	26	"
British Columbia	0	"	41	"
Alaska	2	"	39	"
Algonquin	<b>4</b>	12	128	Rutlege et al. 2010b
Northeast Ontario	5	"	51	
Frontenac Axis	4	"	38	
Alaska and western Canada	5 <sup>a</sup>	11	221	Weckworth et al. 2005
W. Montana (from Alberta founders)	6	10	91	Forbes and Boyd (1996)
<sup>a</sup> Average per population.				

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Table 9.

Comparison:	No. of comparisons	% sequence divergence		Sequence source	Source
	Comparisons	Mean	Range		
Coyote-coyote	3	1.5	0.8 to 2.0	Restriction sites	Lehman et al. 1991
Eastern wolf-coyote	6	1.5	0.9 to 2.0	"	"
Coyote-coyote	10	1.1	0.4 to 1.7	Cytochrome <i>b</i>	Roy et al. 1996
Red wolf-coyote	15	1.2	0.9 to 2.2	"	"
Coyote-coyote	Not reported	1.7	Not reported	Control region	Wilson et al. 2000
Red wolf-coyote	"	2.3	"	"	"
Eastern wolf-coyote	"	3.2	"	"	"
Red wolf-eastern wolf	"	2.1	"	"	"
Coyote-coyote	Not reported	2.8	Not reported	Control region	Adams et al. 2003
Red wolf-coyote	"	3.2	"	"	"

Figure captions:

Figure 1. Ranges of gray wolves in the conterminous United States: (a) historical range; (b) range at time of listing (1978); (c) current range in the Great Lakes states and experimental population areas in the northern Rocky Mountains and for the Mexican wolf (*Canis lupus baileyi*) in the Southwest. Credit: U.S. Fish and Wildlife Service.

Figure 2. Ranges of North American *Canis lupus* subspecies recognize by Nowak (1995, 2002) and of *Canis rufus* (after Nowak 2002).

Figure 3. Subspecies of *Canis lupus* recognized by Goldman (Figure 14 of Goldman 1944 in *The Wolves of North America*, S. P. Young and E. A. Everman, editors, © Wildlife Management Institute; reproduced with permission).

Figure 4. Principal components analysis of gray wolf skulls from central Canada: projection of male groups on first three principal components (Figure 2 of Skeel and Carbyn 1977). Points labeled H, I, and I are from locations within the range of *C. l. nubilus*; L is *C. l. lycaon*; and W, R, J, and P are from within the range of *Canis lupus*. Credit: M. A. Skeel and L. N. Carbyn, The morphological relationship of Gray Wolves (*Canis lupus*) in national parks of central Canada, Canadian Journal of Zoology 55(4):737-747, © 2008 NRC Canada or its licensors. Reproduced with permission.

Figure 5. Principle components plot of skulls from male *Canis lupus* from the southwestern United States. (Figure 2 of Bogan and Mehlhop 1983). B, *C. l. baileyi*; M, *C. l. mogollonensis*; S, *C. l. monstrabilis*. Credit: Museum of Southwestern Biology, University of New Mexico.

Figure 6. Discriminant function analysis of skulls of some North American *Canis lupus* (Figure 7 of Nowak 1995). Axes represent first (horizontal) and second (vertical) canonical variables. Solid lines are limits of the Nowak's southern group (corresponding to *C. l. nubilus*), which is the polygon on the left with center N; and northern group (corresponding to *C. l. occidentalis*), which is the polygon on the right with center O. Dots represent individuals of *C. l. baileyi*. Credit: © Ronald M. Nowak and Canadian Circumpolar Institute Press. Used with permission.

Figure 7. Multi-dimensional scaling plot based on 10 microsatellite DNA loci (Figure 3 of Roy et al. 1996). Confidence ellipses are indicated by dashed (95%) and solid (99%) lines. © John Wiley and Sons. Used with permission.

Figure 8. Neighbor-joining tree based on mtDNA control region sequences of *Canis lupus* from Vilà et al. (1999, Figure 1). North American haplotypes are lu-28, lu-29, lu-30, lu-31, lu-32, lu-33. Others are from Eurasia. The haplotype unique to *C. l. baileyi* is lu-33. © John Wiley and Sons. Used with permission.

Figure 9. Minimum spanning tree for control region haplotypes from eastern wolves (Figure 5(A) of Wilson et al. 2000). *Canis lycaon* haplotypes are C1, C3, C9, and C14; it shares C19 with *C. rufus* and with coyotes. *C. rufus* has haplotypes C2 and C19. The remaining are coyote

3564 haplotypes not found in either wolf. Credit: P. J. Wilson et al., DNA profiles of eastern  
 3565 Canadian wolf and the red wolf provide evidence for a common evolutionary history  
 3566 independent of the gray wolf, Canadian Journal of Zoology 78(12):2156-2166. © 2008 NRC  
 3567 Canada or its licensors. Reproduced with permission.  
 3568  
 3569 Figure 10. Neighbor-joining tree of mtDNA control region haplotypes (Figure 5(B) of Wilson et  
 3570 al. 2000). The long branch extending to the right (C22, C23, C24) represents *Canis lupus*. *Canis*  
 3571 *lycaon* and *Canis rufus* haplotypes are as identified in the caption for Figure 9 of this paper.  
 3572 Remaining haplotypes are coyotes. Scale represents 0.1 (or 10%) sequence divergence. Credit:  
 3573 P. J. Wilson et al., DNA profiles of eastern Canadian wolf and the red wolf provide evidence for  
 3574 a common evolutionary history independent of the gray wolf, Canadian Journal of Zoology  
 3575 78(12):2156-2166. © 2008 NRC Canada or its licensors. Reproduced with permission.  
 3576  
 3577 Figure 11. Distribution of control region mtDNA haplotypes of *Canis lupus* in British Columbia  
 3578 (Figure 3 of Muñoz-Fuentes et al. 2009). © John Wiley and Sons. Used with permission.  
 3579  
 3580 Figure 12. Median-joining network of Y-chromosome haplotypes of western wolves, western  
 3581 Great Lakes states wolves, and Wisconsin coyotes (Figure 4 of Fain et al. 2010). Green = *C.*  
 3582 *lycaon*; yellow = coyotes; orange = *C. lupus*. © Springer. Used with permission.  
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 3584 Figure 13. Summary of major conclusions of this review. Solid lines within North American  
 3585 indicate general boundaries of species and subspecies of *Canis* recognized by Nowak (2002).  
 3586 Dashed lines indicate recommended alterations or qualifications of Nowak's geographic

boundaries of taxa. Geographic positions of these lines are approximate. **A.** *Canis lycaon* is recognized as a species, rather than a subspecies of *C. lupus*. The historical boundary between *C. rufus* and *C. lycaon* is uncertain owing to absence of samples from the potential area of contact. **B.** Range of *C. lycaon* is extended westward; the extended area also includes *C. lupus* and *C. lupus* x *C. lycaon* hybrids. **C.** Extension of historical boundary of *C. l. baileyi* northward based on morphometric and genetic data. **D.** Taxonomic standing of *C. l. arctos* is uncertain. **E.** There are genetic discontinuities between Pacific Coast populations assigned by Nowak to *C. l. nubilus* and adjacent *C. l. occidentalis* populations; they are phylogenetically closest to and the limited number of historical samples of western *C. l. nubilus*. **F.** Current distribution of *C. l. occidentalis* has extended southward through both natural expansion and reintroduction.

3610      Figure 1.

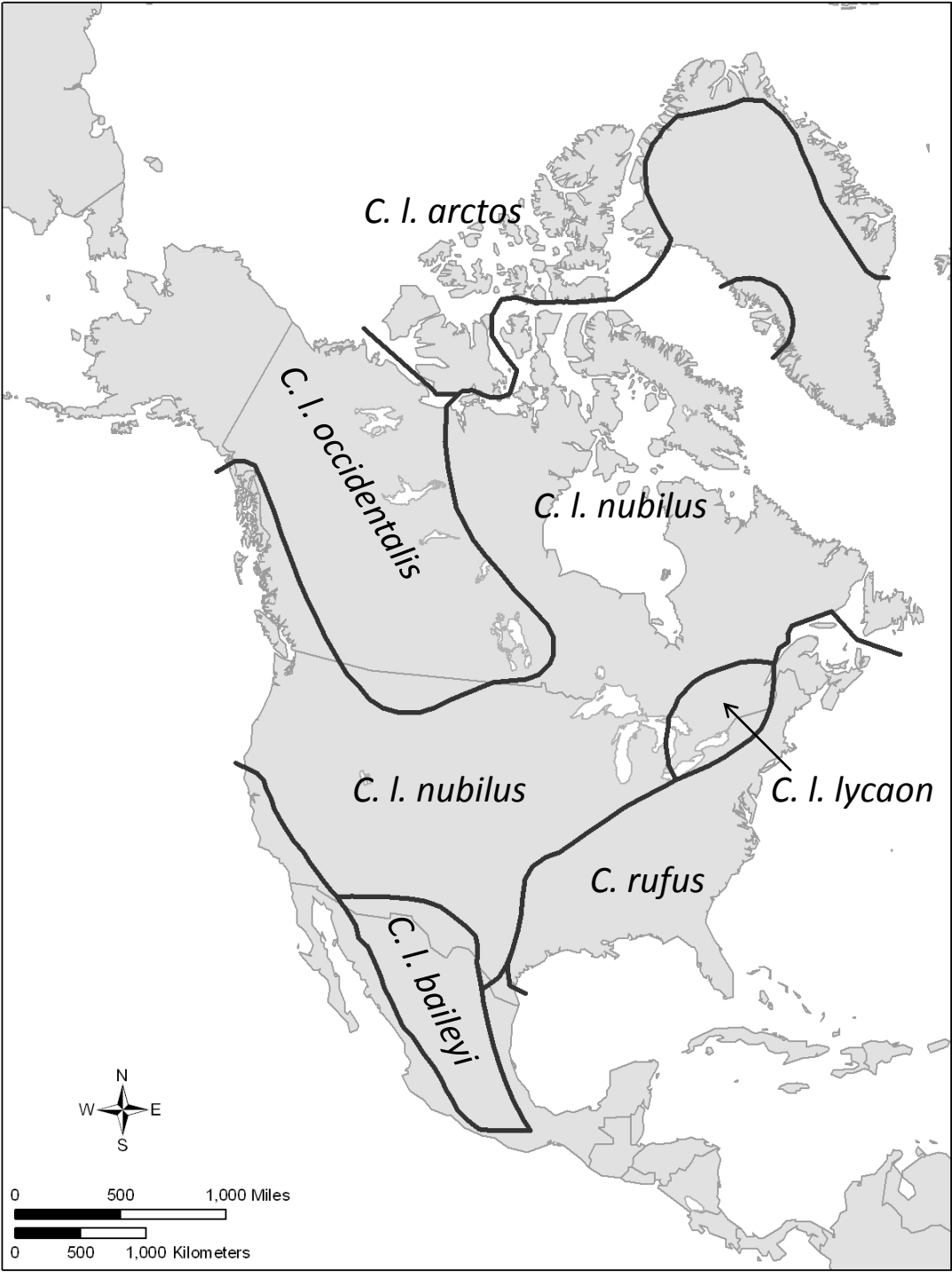


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3614 Figure 2.



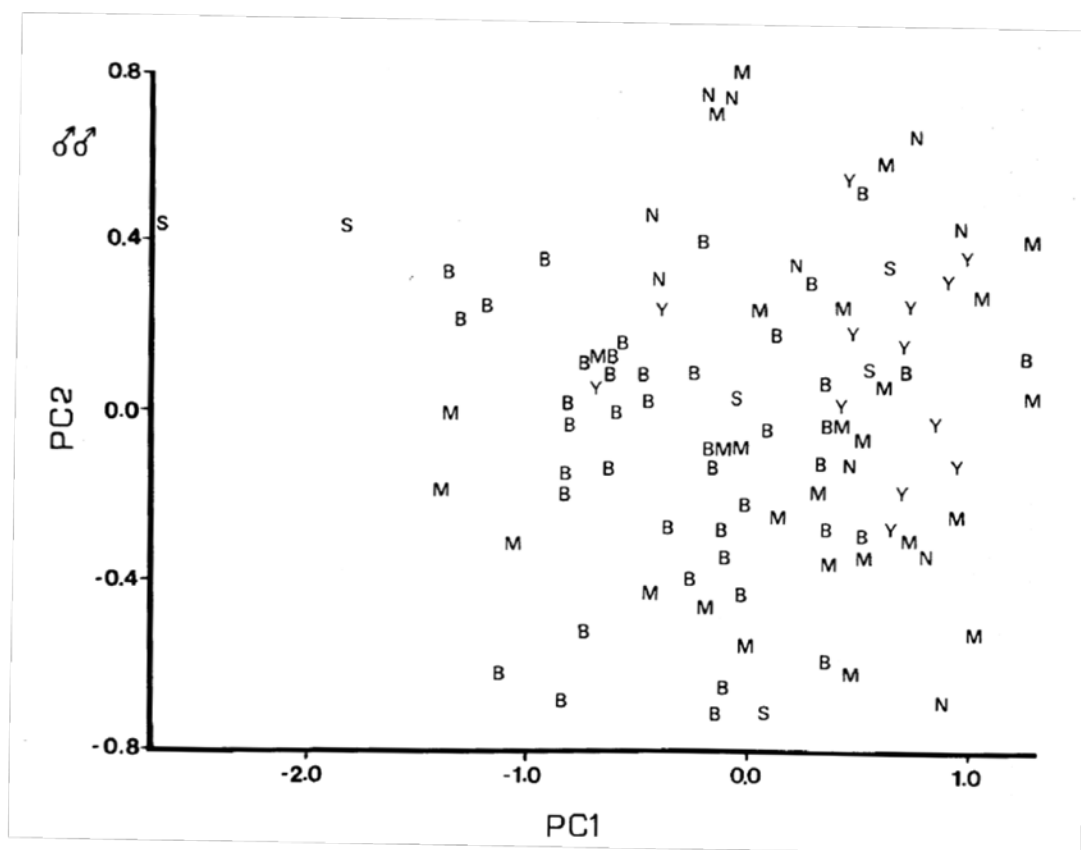
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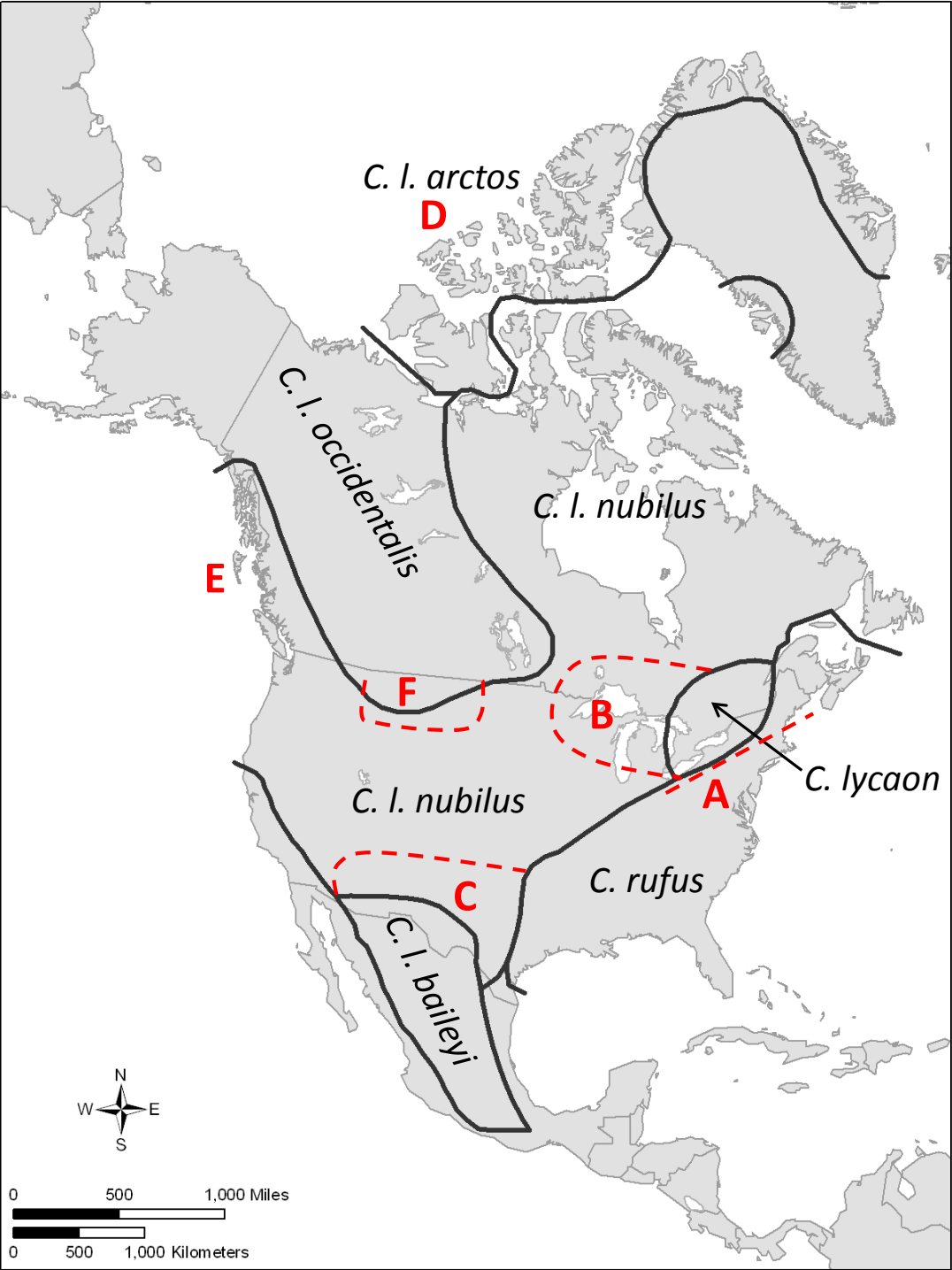
Figure 5.



Figures 6-12 include material under copyright and are not included



3630 Figure 13.



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